Volume 226, Supplement

FEBLAL 226 (Suppl.) 1–88

ISSN 0 014 5793

FEBS LETTERS

MASTER INDEX

VOLUMES 208–226 (1987)

FEBS LETTERS

for the rapid publication of short reports in biochemistry, biophysics and molecular biology

Published by Elsevier
on behalf of the
Federation of European Biochemical Societies

FEBS LETTERS

An international journal established for the most rapid possible publication of essentially final short papers in the field of Biochemistry, Biophysics and Molecular Biology

Published by Elsevier Science Publishers B.V. (Biomedical Division) on behalf of the

FEDERATION OF EUROPEAN BIOCHEMICAL SOCIETIES

A Federation of Biochemical Societies in Austria, Belgium, Bulgaria, Czechoslovakia, Denmark, Federal Republic of Germany, Finland, France, German Democratic Republic, Great Britain, Greece, Hungary, Iceland, Ireland, Israel, Italy, The Netherlands, Norway, Poland, Portugal, Rumania, Spain, Sweden, Switzerland, Turkey, USSR, Yugoslavia.

Editorial policy, subject coverage and submission procedures are described in *Notes to Authors* of this issue.

Submitting authors: Contributors are advised to submit their papers to the Editor who is closest to their field of interest, rather than geographically, to expedite more rapid reviewing.

Editors:

G. SEMENZA (Managing Editor), Eidgenössische Technische Hochschule Zürich, Laboratorium für Biochemie, ETH-Zentrum, Universitätstrasse 16, CH-8092 Zürich, Switzerland
S.P. DATTA (Honorary Chairman), London, England
H.R.V. ARNSTEIN (Reviews Editor), Department of Biochemistry, King's College, Strand, London WC2R 2LS, England

E. BOCK, The Protein Laboratory, University of Copenhagen, 34 Sigurdsgade, DK-2299 Copenhagen N, Denmark
J.E. CELIS, Department of Medical Biochemistry, Ole Worms Allé, Building 170, University Park, Aarhus University, DK-8000 Aarhus, Denmark
F. CHAPEVILLE, Institut Jacques Monod, Université Paris VII — Tour 43, 2, place Jussieu, 75251 Paris Cédex 05, France
P.L. DUTTON, Department of Biochemistry and Biophysics, School of Medicine G3, University of Pennsylvania, Philadelphia, PA 19174, USA
J. HANOUNE, Unité de Recherches, INSERM U-99, Hôpital Henri Mondor, 94010 Créteil, France
H. HOLZER, Biochemisches Institut der Universität, Hermann-Herder-Str. 7, D-7800 Freiburg i.Br., FRG
L.N. JOHNSON, Laboratory of Molecular Biophysics, University of Oxford, Department of Zoology, Rex Richards Building, South Parks Road, Oxford OX1 3QU, England
A. KOTYK, Institute of Physiology, Czechoslovak Academy of Sciences, KRC, Vídeňská 1083, 142 20 Praha 4, Czechoslovakia
A.D. MIRZABEKOV, Institute of Molecular Biology, Academy of Sciences of the USSR, Vavilov str. 32, 117984 Moscow B-334, USSR
Y. NISHIZUKA, Department of Biochemistry, Kobe University School of Medicine, Chuo-ku, Kobe 650, Japan
M.J. OWEN, Laboratory of Lymphocyte Molecular Biology, St. Bartholomew's Hospital, Dominion House, Bartholomew Close, London EC1A 7BE, England
P.J. RANDLE, Nuffield Department of Clinical Biochemistry, John Radcliffe Hospital, Oxford OX3 9DU, England
T.A. RAPOPORT, Zentralinst. Molekularbiologie, Akademie der Wissenschaften der DDR, DDR-1115 Berlin-Buch, GDR
G.C.K. ROBERTS, Department of Biochemistry, University of Leicester, University Road, Leicester LE1 7RH, England
M.L. SALAS, Centro de Biologia Molecular, Universidad Autonóma, Facultad de Ciencias, Canto Blanco, 28049 Madrid, Spain
M. SARASTE, Department of Medical Chemistry, University of Helsinki, Siltavuorenpenger 10, SF 00170 Helsinki, Finland
R. SATO, Osaka University, Institute for Protein Research, 3–2 Yamadoka, Suita, Osaka 565, Japan
N. SHARON, Department of Biophysics, Weizmann Institute of Science, P.O. Box 26, Rehovot 76100, Israel
V.P. SKULACHEV, A.N. Belozersky Laboratory of Molecular Biology and Bioorganic Chemistry, Moscow State University, 119899 Moscow, USSR
A. TREBST, Lehrstuhl für Biochemie der Pflanzen, Ruhr-Universität Bochum, Postfach 10 21 48, 4630 Bochum 1, FRG
I.M. VERMA, The Salk Institute, P.O. Box 85800, San Diego, CA 92138-9216, USA
B. WITKOP, NIADDK, Building 8- B1A-11, National Institutes of Health, Bethesda, MD 20892, USA

Subscription information: The institutional subscription price for 1988, volumes 225–240 (16 volumes in 32 issues) is *f* 3536.00 (US$ 1911.25) including postage, packaging and handling.
Personal subscriptions: FEBS members can order a personal subscription at the reduced rate of *f* 1325.00 (US$ 716.25) for 1988, including postage and handling. Members of the American Society of Biological Chemists and the Japanese Biochemical Society may order a personal subscription at the reduced rate of *f* 1830.00 (US$ 989.25), including postage and handling. The following rules apply: (1) the subscriptions must be prepaid; (2) the order must be sent directly to the Publisher; (3) the copies should not be made available to institutions and laboratories. The Dutch guilder price is definitive. The US$ price is subject to exchange rate fluctuations and is only given as a guide. Journals are sent by surface delivery to all countries, except the following where Surface Airlifted Mail is ensured: USA, Canada, Japan, Australia, New Zealand, P.R. China, India, Israel, South Africa, Malaysia, Singapore, South Korea, Taiwan, Pakistan, Hong Kong, Brazil, Argentina, Mexico and Thailand. Airmail rates for other countries are available on request.
Subscription orders can only be entered by calendar year (January to December) and should be sent to: Elsevier Science Publishers, Journals Department, P.O. Box 211, 1000 AE Amsterdam, The Netherlands; telephone (020) 5803911, telex 18582 ESPA NL; or your usual subscription agent. Claims for missing issues should be made within three months of publication, otherwise they cannot be honoured free of charge. The Publishers reserve the right to issue additional volumes during the course of the year. Such volumes will be invoiced before publication and delivered on receipt of payment. In the United States and Canada: For further information concerning this or any other Elsevier Science Publishers journal, contact Elsevier Science Publishers, Journal Information Center, 52 Vanderbilt Avenue, New York, NY 10017, USA; telephone (212) 370 5520.

Published fortnightly. **Printed in The Netherlands.**
Elsevier Science Publishers B.V., P.O. Box 211, 1000 AE Amsterdam, The Netherlands

FEBS LETTERS

MASTER INDEX TO VOLUMES 208–226

FEBS LETTERS

for the rapid publication of short reports in
biochemistry, biophysics and molecular biology

EDITORS

H.R.V. Arnstein, London, Reviews Editor
E. Bock, Copenhagen
J.E. Celis, Aarhus
F. Chapeville, Paris
S.P. Datta, London, Honorary Chairman
P.L. Dutton, Philadelphia
J. Hanoune, Créteil
H. Holzer, Freiburg i.Br.
L.N. Johnson, Oxford
A. Kotyk, Prague
A.D. Mirzabekov, Moscow
Y. Nishizuka, Kobe
M.J. Owen, London
P.J. Randle, Oxford
T.A. Rapoport, Berlin-Buch
G.C.K. Roberts, Leicester
M.L. Salas, Madrid
M. Saraste, Helsinki
R. Sato, Osaka
G. Semenza, Zürich, Managing Editor
N. Sharon, Rehovot
V.P. Skulachev, Moscow
A. Trebst, Bochum
I.M. Verma, San Diego
B. Witkop, Bethesda

Published by Elsevier Science Publishers B.V.
on behalf of the
Federation of European Biochemical Societies

The exclusive © for all languages and countries is vested in the Federation of European Biochemical Societies

*No part of this volume may be reproduced in any form by print, photoprint, microfilm or any other means without the prior permission of the copyright owner**

No responsibility is assumed by the Publisher for any injury and/or damage to persons or property as a matter of products liability, negligence or otherwise, or from any use or operation of any methods, products, instructions or ideas contained in the material herein. Because of the rapid advances in the medical sciences, the Publisher recommends that independent verification of diagnoses and drug dosages should be made.

Although all advertising material is expected to conform to ethical (medical) standards, inclusion in this publication does not constitute a guarantee or endorsement of the quality or value of such product or of the claims made of it by its manufacturer.

Special regulations for readers in the U.S.A.

This journal has been registered with the Copyright Clearance Center, Inc. Consent is given for copying of articles for personal or internal use, or for the personal use of specific clients. This consent is given on the condition that the copier pays through the Center the per copy fee stated in the code on the first page of each article for copying beyond that permitted by Sections 107 or 108 of the U.S. Copyright Law. The appropriate fee should be forwarded with a copy of the first page of the article to the Copyright Clearance Center, Inc., 27 Congress Street, Salem, MA 01970, U.S.A. If no code appears in an article, the author has not given broad consent to copy and permission to copy must be obtained directly from the author. All articles published prior to 1981 may be copied for a per-copy fee of US $ 2.25, also payable through the Center. (N.B. For review journals this fee is $ 0.25 per copy per page.) This consent does not extend to other kinds of copying, such as for general distribution, resale, advertising and promotion purposes, or for creating new collective works. Special written permission must be obtained from the publisher for such copying.

Special regulations for authors

Upon acceptance of an article by the journal, the author(s) will be asked to transfer copyright of the article to the Federation of European Biochemical Societies. This transfer will ensure the widest possible dissemination of information

* But see item 6 in Notes to Authors

PRINTED IN THE NETHERLANDS

FEBS Letters Volumes 208–226
Master Author Index

A

Aarbakke, G.M. see O.-B. Tysnes (218) 68
Aarsman, A.J., G. Mynbeek, H. van den Bosch, B. Rothhut, B. Prieur, C. Comera, L. Jordan and F. Russo-Marie, Lipocortin inhibition of extracellular and intracellular phospholipases A_2 is substrate concentration dependent (219) 176
Aasa, R., L.-E. Andréasson, G. Lagenfelt and T. Vänngård, A comparison between the multiline EPR signals of spinach and *Anacystis nidulans* and their temperature dependence (221) 245
Aaviksaar, A. see V. Tóugu (225) 77
Abbott, M.S., N. Shavit, S. Selman-Reimer and B.R. Selman, Characterization of nucleotide-binding sites on the chloroplast coupling factor 1 using two photolabile analogs (209) 157
Abbott, W.M. see C. Southan (214) 301
Abdel-Latif, A.A. see P.H. Howe (215) 279
Abdulaev, N.G., I.R. Nabiev, R.G. Efremov and G.D. Chumanov, Retinal Schiff base position relative to the surfaces of photoreceptor disk (213) 113
Abe, E. see J. Abe (226) 58
Abe, E. see T. Hayashi (218) 200
Abe, J., M. Morikawa, K. Miyamoto, S. Kaiho, M. Fukushima, C. Miyaura, E. Abe, T. Suda and Y. Nishii, Synthetic analogues of vitamin D_3 with an oxygen atom in the side chain skeleton: A trial of the development of vitamin D compounds which exhibit potent differentiation-inducing activity without inducing hypercalcemia (226) 58
Abell, C.W. see A. Brossi (214) 291
Abraham, A.K. see G. Burns (208) 217
Abrahamson, M., A. Grubb, I, Olafsson and Å. Lundwall, Molecular cloning and sequence analysis of cDNA coding for the precursor of the human cysteine proteinase inhibitor cystatin C (216) 229
Abramowitz, J. see W.N. Suki (220) 187
Acher, R. see J. Chauvet (217) 180
Acher, R. see M.T. Chauvet (210) 40
Adachi, H. see A. Yamaguchi (218) 126
Adachi, H., T. Ohta and H. Matsuzawa, A water-soluble form of penicillin-binding protein 2 of *Escherichia coli* constructed by site-directed mutagenesis (226) 150
Adam, A. see J.-P. Tenu (220) 93
Adam, Z. and R. Malkin, Reconstitution of isolated Rieske Fe-S protein into a Rieske-depleted cytochrome b_6-f complex (225) 67
Adams, M.J. see M.M. Bhadbhade (211) 243
Adams, R.L.P. and A. Rinaldi, Effect of echinomycin on DNA methylation (215) 266
Adams, S.P. see G.M. Olins (224) 325
Adavani, S.R. see B.A. Hemmings (209) 219
Adler, A. see S.I. Walaas (220) 311
Adrouche, N. see S. Benjannet (224) 142
Aerts, R.J., R.J.W. De Wit and M.M. Van Lookeren Campagne, Cyclic AMP induces a transient alkalinization in *Dictyostelium* (220) 366
Agalarov, S.C. see S.D. Trakhanov (220) 319
Agalarov, S.C. see S.E. Sedelnikova (220) 227
Agarwal, D.P. see T. Braun (215) 233
Agarwal, M.K., B. Hainque, N. Moustaid and G. Lazer, Glucocorticoid antagonists (217) 221
Aggeler, R. see R. Ghosh (222) 154
Agneray, J. see C. Aussel (214) 327
Agostinis, P., J.R. Vandenheede, J. Goris, F. Meggio, L.A. Pinna and W. Merlevede, The ATP,Mg-dependent protein phosphatase: regulation by casein kinase-1 (224) 385
Ahmad, A., R.K. Gordon and P.K. Chiang, A microtechnique for quantification of detergent-solubilized muscarinic and nicotinic acetylcholine receptors using a semiautomated cell harvestor (214) 285
Ahmad, S. and R.A. Jensen, The prephenate dehydrogenase component of the bifunctional T-protein in enteric bacteria can utilize L-arogenate (216) 133
Ahomadegbe, J.-C. see J.-F. Riou (213) 304
Aimoto, S. see Y. Shimonishi (215) 165
Aitken, A. see A.P. Bradford (222) 211
Aitken, A. see C.A. Ellis (218) 238
Aitken, A. see C. Southan (214) 301
Aitken, A. see S.F. Brooks (224) 109
Aizono, Y. see T. Utsumi (216) 99
Akerboom, T.P.M. see J.C. Griffiths (213) 34
Åkerström, G. see P. Nygren (213) 195
Akhrem, A.A. see M.A. Rodionov (223) 402
Akiba, I. see T. Kubo (209) 367
Akiyama, K., T. Nakamura, S. Iwanaga and M. Hara, The chymotrypsin-like activity of human prostate-specific antigen, γ-seminoprotein (225) 168
Akiyama, T. see M. Hoshi (217) 237
Akiyama, T. see Y. Ohta (222) 305
Akkerman, J.W.N. see W. Siffert (212) 123
Akopyanz, N.S. see E.D. Sverdlov (221) 129
Aksenova, N.B. see Yu.S. Borovikov (223) 409
Aktories, K. see J. Vandekerckhove (225) 48
Aktories, K. see S. Rösener (224) 38
Aktories, K., U. Weller and G.S. Chhatwal, *Clostridium botulinum* type C produces a novel ADP-ribosyltransferase distinct from botulinum C2 toxin (212) 109
Akutsu, H. see S. Nishimoto (213) 293
Al-Kabban, M. see J. Reglinski (214) 351
Al-Mohanna, F.A., I. Ohishi and M.B. Hallett, Botulinum C_2 toxin potentiates activation of the neutrophil oxidase: Further evidence of a role for actin polymerization (219) 40
Albagnac, G. see E. Samain (216) 140
Albert, K.A. see S.I. Walaas (220) 311
Albuquerque, E.X. see C. Rapier (212) 292
Albuquerque, E.X. see D.R.E. Macallan (226) 357
Albuquerque, E.X. see Y. Aracava (222) 63
Aldanova, N.A. see Yu.A. Ovchinnikov (217) 269
Alder, L. see H. Kühn (208) 248
Alemany, S. see O.B. da Cruz e Silva (221) 415
Aletsee, M.C. see O.K. Langley (220) 108
Alfani, E. see C. Delfini (210) 17
Alfsen, A. see C. de Paillerets (219) 113
Alhonen-Hongisto, L. see P. Leinonen (215) 68
Alibert, G. see M. Dillenschneider (208) 413
Alitalo, R., The *bcr*-c-*abl* tyrosine kinase activity is extinguished by TPA in K562 leukemia cells (222) 293
Allakhverdiev, S.I., E. Šetlíková, V.V. Klimov and I. Šetlík, In photoinhibited photosystem II particles pheophytin photoreduction remains unimpaired (226) 186
Allan, E.H. see A.E. Fletcher (208) 263
Alleman, J. see J.A.W.H. Vermeulen (219) 426
Allen, A.K. see A.M. Vranken (216) 67
Allen, A.K. see E.J.M. Van Damme (215) 140
Allen, A.K. see W.F. Broekaert (220) 116
Allen, E. see R.I. Norman (212) 127
Allen, J.F. see R.S. Williams (225) 59
Allen, T.M. and A. Chonn, Large unilamellar liposomes with low uptake into the reticuloendothelial system (223) 42
Allende, C.C. see A. Taylor (226) 109
Allende, J.E. see A. Taylor (226) 109
Allende, J.E. see P. Carvallo (215) 109

Allikmets, R.L. see E.D. Sverdlov (217) 275
Allikmets, R.L. see Yu.A. Ovchinnikov (213) 73
Allorio, M. see J.L. Lorenzo (218) 77
Allsopp, M.T.E.P. see I.N.M. Day (222) 139
Ally, S. see D. Katsaros (223) 97
Ally, S. see T. Clair (224) 377
Alon, R. see M.S. Meyer (212) 138
Alpert, B. see S. Pin (208) 325
Alster, D. see P.F. Torrence (212) 267
Altin, J.G. see C. Preston (210) 27
Altin, J.G. see P. Dieter (213) 174
Altmann, F., L. März, S. Stirm and F.M. Unger, Two additional bacteriophage-associated glycan hydrolases cleaving ketosidic bonds of 3-deoxy-D-*manno*-octulosonic acid in capsular polysaccharides of *Escherichia coli* (221) 145
Alvarez, E. see J.G. de Diego (222) 256
Alvarez, J.F. see I. Varela (211) 64
Amano, T. see H. Nakayama (208) 278
Amano, T. see T. Yamakuni (223) 117
Ambler, R.P., A.D. Auffret and P.H. Clarke, The amino acid sequence of the aliphatic amidase from *Pseudomonas aeruginosa* (215) 285
Ameen, M. and P.L. Chang, Pseudo arylsulfatase A deficiency: Biosynthesis of an abnormal arylsulfatase A (219) 130
Amelizad, Z., J.F. Narbonne, C. Borin, L.W. Robertson, A. Periquet and F. Oesch, Effect of unbalanced diets on incorporation of δ-aminolevulinic acid into cytochrome P-450 (220) 231
Amons, R. see G.D.F. Maessen (208) 77
Amons, R. see G.D.F. Maessen (223) 181
Amons, R., Vapor-phase modification of sulfhydryl groups in proteins (212) 68
Amrute, S.B. see V.N. Pandey (213) 204
Andersen, J., A model for the interaction of nucleic acids with transcription factor IIIA (217) 197
Anderson, G.J. and S. Kumar, Transacylase activity of lactating bovine mammary fatty acid synthase (220) 323
Anderson, J.M. and D.J. Goodchild, Transbilayer organization of the main chlorophyll *a*/*b*-protein of photosystem II of thylakoid membranes (213) 29
Anderson, J.M. see M. Spangfort (224) 343
Anderson, L.E., Ribose-5-phosphate isomerase and ribulose-5-phosphate kinase show apparent specificity for a specific ribulose 5-phosphate species (212) 45
Anderson, W.B. see I. Magnaldo (210) 6
Anderson, W.B. see R. Gopalakrishna (225) 233
Andersson, B. see E. Glaser (223) 304
Andersson, B. see M. Spangfort (224) 343
Andersson, B. see P. Mäenpää (215) 31
Andersson, B. see T. Jansen (216) 234
Anderton, B.H. see J.-P. Brion (226) 28
Anderton, B.H. see P.A. Robinson (209) 203
Andreasen, P.A., A. Riccio, K.G. Welinder, R. Douglas, R. Sartorio, L.S. Nielsen, C. Oppenheimer, F. Blasi and K. Danø, Plasminogen activator inhibitor type-1: reactive center and amino-terminal heterogeneity determined by protein and cDNA sequencing (209) 213
Andréasson, L.E. see R. Aasa (221) 245
Andreo, C.S., D.H. Gonzalez and A.A. Iglesias, Higher plant phospho*enol*pyruvate carboxylase: Structure and regulation (213) 1
Andrews, P.C. see T.P. Mommsen (219) 227
Andreyev, A.Yu., T.O. Bondareva, V.I. Dedukhova, E.N. Mokhova, V.P. Skulachev and N.I. Volkov, Carboxyatractylate inhibits the uncoupling effect of free fatty acids (226) 265
Andrianjara, R. see D. Rogez (219) 22
Andus, T., T. Geiger, T. Hirano, H. Northoff, U. Ganter, J. Bauer, T. Kishimoto and P.C. Heinrich, Recombinant human B cell stimulatory factor 2 (BSF-2/IFN-β2) regulates β-fibrinogen and albumin mRNA levels in Fao-9 cells (221) 18
Angeletti, P.U. see P. Conti (225) 103
Ängquist, K.-A. see L. Bläckberg (217) 37
Aniento, F. see E. Roche (221) 231
Anjaneyulu, P.S.R. see R.M. Mogre (221) 408
Ankel-Fuchs, D., R. Böcher, R.K. Thauer, K.M. Noll and R.S. Wolfe, 7-Mercaptoheptanoylthreonine phosphate functions as component B in ATP-independent methane formation from methyl-CoM with reduced cobalamin as electron donor (213) 123
Ankel-Fuchs, D. see A. Kobelt (214) 265
Anni, H. see C. Balny (221) 349
Antia, R. see P. Williamson (219) 316
Antoch, M.P. see V.Yu. Arshavsky (224) 19
Antonenkov, V.D., S.V. Pirozhkov and L.F. Panchenko, On the role of microsomal aldehyde dehydrogenase in metabolism of aldehydic products of lipid peroxidation (224) 357
Antonetti, A. see J. Breton (209) 37
Antoni, F. see L. Buday (223) 15
Antoni, G. see M. Bigio (218) 271
Aoki, S. see Y. Kuroda (224) 137
Aoshima, M. see S. Ohno (222) 279
Aoshima, T. see K. Satoh (216) 7
Aota, S. see T. Gojobori (208) 231
Aoyagi, H. see S. Ono (220) 332
Aoyama, T. see H. Nakayama (208) 278
Apone, S. see D.R. Eyre (220) 337
Appanna, V.D. and C.M. Preston, Manganese elicits the synthesis of a novel exopolysaccharide in an arctic *Rhizobium* (215) 79
Appel, R.G., G.R. Dubyak and M.J. Dunn, Effect of atrial natriuretic factor on cytosolic free calcium in rat glomerular mesangial cells (224) 396
Appelhans, B., B. Ender, G. Sachse, T. Nikiforov, H. Appelhans and W. Ebert, Secretion of antileucoprotease from a human lung tumor cell line (224) 14
Appelhans, H. see B. Appelhans (224) 14
Aquila, H., T.A. Link and M. Klingenberg, Solute carriers involved in energy transfer of mitochondria form a homologous protein family (212) 1
Aracava, Y., S.S. Deshpande, K.L. Swanson, H. Rapoport, S. Wonnacott, G. Lunt and E.X. Albuquerque, Nicotinic acetylcholine receptors in cultured neurons from the hippocampus and brain stem of the rat characterized by single channel recording (222) 63
Arad, T. see J. Piefke (209) 104
Arad, T. see M. Shoham (208) 321
Aragón, J.J., M.-E. Gómez and C. Gancedo, Identification of two forms of 6-phosphofructo-2-kinase in yeast (226) 121
Aragón, M.C., C. Giménez and F. Mayor, Stoichiometry of sodium- and chloride-coupled glycine transport in synaptic plasma membrane vesicles derived from rat brain (212) 87
Arai, H. see S. Ohta (226) 171
Araki, Y. see N. Murazumi (218) 131
Aratani, Y., E. Sugimoto and Y. Kitagawa, Lithium ion reversibly inhibits inducer-stimulated adipose conversion of 3T3-L1 cells (218) 47
Archidiacono, N. see D. Fürst (224) 49
Ardaillou, N. see D. Chansel (220) 247
Ardaillou, R. see D. Chansel (220) 247
Ariki, M., Y. Shichida and T. Yoshizawa, Low temperature spectrophotometry on the photoreaction cycle of sensory rhodopsin (225) 255

Arkowitz, R., M. Hoehn-Berlage and K. Gersonde, The effect of cadmium ions on 2,3-bisphosphoglycerate in erythrocytes studied with ^{31}P NMR (217) 21
Arlaud, G.J., A. Van Dorsselaer, A. Bell, M. Mancini, C. Aude and J. Gagnon, Identification of *erythro-β*-hydroxyasparagine in the EGF-like domain of human C1r (222) 129
Armstrong, A. see A.J. Harmar (208) 67
Arndt, E. see J. Kimura (224) 65
Arora, S.K. see E.J. Kattelman (213) 179
Arroyo, C.M., J.H. Kramer, B.F. Dickens and W.B. Weglicki, Identification of free radicals in myocardial ischemia/reperfusion by spin trapping with nitrone DMPO (221) 101
Arseniev, A.S., A.B. Kuryatov, V.I. Tsetlin, V.F. Bystrov, V.T. Ivanov and Yu.A. Ovchinnikov, ^{19}F NMR study of 5-fluorotryptophan-labeled bacteriorhodopsin (213) 283
Arshavsky, V.Yu., M.P. Antoch and P.P. Philippov, On the role of transducin GTPase in the quenching of a phosphodiesterase cascade of vision (224) 19
Arthur, W.L. see C.A. O'Brian (214) 339
Arystarkhova, E.A. see Yu.A. Ovchinnikov (217) 269
Arzamazova, N.M. see Yu.A. Ovchinnikov (217) 269
Asahi, T. see M. Maeshima (220) 23
Asano, M. see H. Tokuda (215) 335
Asaoka, Y. see U. Kikkawa (217) 227
Asaoka, Y. see U. Kikkawa (223) 212
Ase, K. see U. Kikkawa (223) 212
Ashby, M.K., S.A. Coomber and C.N. Hunter, Cloning, nucleotide sequence and transfer of genes for the B800-850 light harvesting complex of *Rhodobacter sphaeroides* (213) 245
Ashcroft, F.M., M. Kakei, R.P. Kelly and R. Sutton, ATP-sensitive K^+ channels in human isolated pancreatic B-cells (215) 9
Ashcroft, F.M. see M. Kakei (208) 63
Ashcroft, S.J.H. and M. Stubbs, The glucose sensor in HIT cells is the glucose transporter (219) 311
Ashcroft, S.J.H. see M. Kakei (208) 63
Ashcroft, S.J.H. see P. Hammonds (213) 149
Ashcroft, S.J.H. see P. Hammonds (223) 131
Ashford, M.L.J. see N.C. Sturgess (208) 397
Ashkenazi, A., R. Cohen and A. Gertler, Characterization of lactogen receptors in lactogenic hormone-dependent and independent NB2 lymphoma cell lines (210) 51
Ashley, C.C. see A.P. Jackson (216) 35
Ashley, C.C. see M.P. Timmerman (209) 1
Aslanian, D., P. Gróf, M. Négrerie, M. Balkanski and P. Taylor, Raman spectroscopic study on the conformation of 11 S form acetylcholinesterase from *Torpedo californica* (219) 202
Astashkin, A.V. see S.A. Dikanov (224) 75
Astier, C. see J. Olive (208) 308
Astruc, M.E. see F. Beseme (210) 97
Atabekov, J.G. see S.Yu. Morozov (213) 438
Atabekov, J.G. see V.V. Dolja (214) 308
Atabekov, J.G. see V.V. Dolja (214) 313
Atanasov, B.P. see M.I. Dimitrov (226) 17
Atlas, D. see S. Diamant (219) 445
Atreyi, M. see K. Gopalakrishna (215) 95
Attardi, G. see F.G.P. Earley (219) 108
Attfield, P.V., Trehalose accumulates in *Saccharomyces cerevisiae* during exposure to agents that induce heat shock response (225) 259
Aude, C. see G.J. Arlaud (222) 129
Auffret, A.D. see R.P. Ambler (215) 285
Augusteyn, R.C. and J.F. Koretz, A possible structure for α-crystallin (222) 1
Aunis, D. see M. Toutant (215) 339
Aussel, C., D. Desmoulins, J. Agneray and O.G. Ekindjian, Effect of insulin on aminoisobutyric acid uptake by human non-rheumatoid and rheumatoid synovial cells (214) 327
Austen, B. see A. Robinson (218) 63
Authi, K.S., E.J. Hornby, B.J. Evenden and N. Crawford, Inositol 1,4,5-trisphosphate (IP_3) induced rapid formation of thromboxane B_2 in saponin-permeabilised human platelets: mechanism of IP_3 action (213) 95
Avarmaa, R. see K. Mauring (223) 165
Avichezer, D. and N. Gilboa-Garber, PA-II, the L-fucose and D-mannose binding lectin of *Pseudomonas aeruginosa* stimulates human peripheral lymphocytes and murine splenocytes (216) 62
Avraham, B. see S. Diamant (219) 445
Ayad, S., A.P.L. Kwan and M.E. Grant, Partial characterization of type X collagen from bovine growth-plate cartilage: Evidence that type X collagen is processed in vivo (220) 181
Ayala, C. see D. Ukena (209) 122
Ayyer, J. see P. Parrack (212) 297

B

Baader, E. see H.M. Hanauske-Abel (214) 236
Baas, P.D., H. Liewerink, H.A.A.M. van Teeffelen, A.D.M. van Mansfeld, J.H. van Boom and H.S. Jansz, Alteration of the ATG start codon of the A* protein of bacteriophage φX174 into an ATT codon yields a viable phage indicating that A* protein is not essential for φX174 reproduction (218) 119
Baba, S. see M. Nagao (214) 107
Babin, F. see G. Bloch (219) 464
Bachrach, U. see R.B. Frydman (219) 380
Bachs, O. see M.J. Coll (208) 418
Bader, J. see S. Nagata (210) 66
Bagi, G. see G. Premecz (226) 13
Bagnarelli, P. see M. Clementi (221) 11
Bagshaw, C.R. see A.P. Jackson (216) 35
Bailey, A.J. see K. Barnard (212) 49
Baker, J.K. see A. Brossi (223) 77
Baker, jr, J.R. see A.P. Weetman (211) 69
Baklouti, F., Y. Giraud, A. Francina, G. Richard, C. Périer, A. Geyssant, J. Jaubert, C. Brizard and J. Delaunay, Hemoglobin Grange-Blanche [β27(B9) Ala→Val], a new variant with normal expression and increased affinity for oxygen (223) 59
Bakr, S. see J. Bennett (210) 22
Balaban, R.S. see L.A. Katz (221) 270
Baldwin, J.E. and M.J.C. Crabbe, A spectrophotometric assay for deacetoxycephalosporin C synthase (214) 357
Balgavý, P. see L. Horniak (224) 283
Balkanski, M. see D. Aslanian (219) 202
Ballabio, A. see G. Sebastio (208) 460
Ballardie, F.W. see M.E. Hayes (220) 307
Ball, E.H. see G.A. Cates (218) 195
Ball, J.A. see Y.C. Lee (220) 243
Balny, C., H. Anni and T. Yonetani, A stopped-flow study of the reaction of cytochrome *c* peroxidase with hydroperoxides (221) 349
Baltscheffsky, M. see Å. Strid (224) 348
Baltz, T. see F. Jähnig (221) 37
Baluda, M. see C. Kryceve-Martinerie (214) 81
Bamberg, E. see K. Fendler (224) 83

Banerjee, A. and R.F. Luduena, Kinetics of association and dissociation of colchicine-tubulin complex from brain and renal tubulin: Evidence for the existence of multiple isotypes of tubulin in brain with differential affinity to colchicine (219) 103
Banerjee, D. see G.R. Laverdure (222) 261
Bánhegyi, G. see L. Buday (223) 15
Banks, G.R. see A.M. MacNicol (221) 48
Banner, D.W. see K. Petratos (218) 209
Baralle, F.E. see B. Carnemolla (215) 269
Baranov, V.I. see L.A. Ryabova (226) 255
Barbarić, S. see V. Mrša (217) 174
Barber, J., D.J. Chapman and A. Telfer, Characterisation of a PS II reaction centre isolated from the chloroplasts of *Pisum sativum* (220) 67
Barber, J. see K. Gounaris (211) 94
Barber, J. see P.J. Nixon (209) 83
Barber, M.J., B.A. Notton and L.P. Solomonson, Oxidation-reduction midpoint potentials of the molybdenum center in spinach NADH:nitrate reductase (213) 372
Bardat, F. see O. Dogbo (210) 211
Bardin, C.W. see G.L. Hammond (215) 100
Bardosi, A. see D. Fürst (224) 49
Barg, B. see V. Witzemann (223) 104
Barkas, T. see B. Roth (221) 172
Barlas, A., X. Gao and L.M. Greenbaum, Isolation of a thiol-activated T-kininogenase from the rat submandibular gland (218) 266
Barlow, D.J. and P.L. Poole, The hydration of protein secondary structures (213) 423
Bärmann, M. see J. Vandekerckhove (225) 48
Barnard, K., L.J. Gathercole and A.J. Bailey, Basement membrane collagen – evidence for a novel molecular packing (212) 49
Barnett, L.K., C.K. Clugston and G.I. Jenkins, Two phytochrome-mediated effects of light on transcription of genes encoding the small subunit of ribulose-1,5-bisphosphate carboxylase-oxygenase in dark-grown pea *(Pisum sativum)* plants (224) 287
Barone, S. see K.A. Jacobson (225) 97
Barra, D. see M.E. Schininà (221) 87
Barratt, G. see J.-P. Tenu (220) 93
Barret Kalindjian, S. see A.J. Garman (223) 361
Barrett, D.J. see T. van Veen (208) 133
Barrett, M.L. see L.A.J. O'Neill (212) 35
Barrio, J.R. see S.C. Huang (216) 128
Barros-Söderling, J. see Z. Chen (226) 43
Bartels, H., B. Vogt and K. Jungermann, Glycogen synthesis from pyruvate in the periportal and from glucose in the perivenous zone in perfused livers from fasted rats (221) 277
Barton, G.J. see M.J.E. Sternberg (218) 231
Bartosz, G. see A.V. Peskin (219) 212
Bartrons, R. see C. Espinet (209) 254
Bartrons, R. see C. Gallego (222) 167
Barz, W. see K. Tiemann (213) 324
Barz, W. see W. Hinderer (214) 101
Basilico, C. see L. Lania (219) 400
Basset, M. see A. Laurent (226) 324
Batenburg, A.M., J.H. van Esch, J. Leunissen-Bijvelt, A.J. Verkleij and B. de Kruijff, Interaction of melittin with negatively charged phospholipids: consequences for lipid organization (223) 148
Batke, J. see P. Tompa (214) 244
Batliwala, H.F. see R.M. Mogre (221) 408
Bauché, F., A.M. Bourdeaux-Jaubert, Y. Giudicelli and R. Nordmann, Ethanol alters the adenosine receptor-N_i-mediated adenylate cyclase inhibitory response in rat brain cortex in vitro (219) 296
Bauer, J. see T. Andus (221) 18
Bäuerlein, E. see Z. Gao (223) 366
Baumann, M. see L. Keso (215) 105
Baumeister, W. see J.P. Chalcroft (211) 53
Bause, E. see R. Günther (221) 293
Baverel, G. see C. Michoudet (216) 113
Bax, A. see V. Sklenář (208) 94
Bax, A. see V. Sklenář (216) 249
Beabealashvilli, R. see N. Dyatkina (219) 151
Beatty, J.T. see M.E. Forrest (212) 28
Beaumier, L., N. Faucher and P.H. Naccache, Arachidonic acid-induced release of calcium in permeabilized human neutrophils (221) 289
Becker, A. see W. Kühlbrandt (226) 275
Beddell, C.R. see D.K. Stammers (218) 178
Beever, K. see J. Furmaniak (215) 316
Beg, F. see A.P. Bradford (222) 211
Beg, O.U., H. von Bahr-Lindström, Z.H. Zaidi and H. Jörnvall, Characterization of a heterogeneous camel milk whey non-casein protein (216) 270
Beggs, M., H. Patel, J. Espinal and P.J. Randle, Temporal relationships in the effects of protein-free diet on the activities of rat liver branched-chain ketoacid dehydrogenase complex and kinase (215) 13
Beisiegel, U. see J.R. Havinga (216) 275
Belamri, M. see M. Savart (216) 22
Belfrage, P. see H. Olsson (209) 175
Belfrage, P. see J. Donnér (208) 269
Belin, D. see J.-D. Vassalli (214) 187
Belkin, A.M. and V.E. Koteliansky, Interaction of iodinated vinculin, metavinculin and α-actinin with cytoskeletal proteins (220) 291
Bell, A. see G.J. Arlaud (222) 129
Bell, J.D., J.C.C. Brown, J.K. Nicholson and P.J. Sadler, Assignment of resonances for 'acute-phase' glycoproteins in high resolution proton NMR spectra of human blood plasma (215) 311
Bell, J.D., P.J. Sadler, A.F. Macleod, P.R. Turner and A.L. Ville, ^{1}H NMR studies of human blood plasma: Assignment of resonances for lipoproteins (219) 239
Bellatin, J. see P.W. Piper (214) 143
Bellelli, A. see M. Brunori (221) 161
Bellemann, P. see D. Neuser (209) 347
Bellot, R. see J.-M. Sallenave (219) 37
Bembenek, M.E. see A. Brossi (214) 291
Benazzi, L. see P.L. Manachini (214) 305
Benchetrit, T. see C. Gaboriaud (224) 149
Bendicenti di Girolamo, A. see J.A. Lewis (217) 292
Benditt, E.P. see L.H. Ericsson (218) 11
Bendriss, P. see P. Dabadie (226) 77
Benhaim, A., M. Herrou, H. Mittre and P. Leymarie, Effects of phorbol esters on steroidogenesis in small bovine luteal cells (223) 321
Benito, M. see C. Roncero (208) 105
Benjannet, S., R. Leduc, N. Adrouche, J.P. Falgueyret, M. Marcinkiewicz, N.G. Seidah, M. Mbikay, C. Lazure and M. Chretien, Chromogranin B (secretogranin I), a putative precursor of two novel pituitary peptides through processing at paired basic residues (224) 142
Bennett, J., E.K. Shaw and S. Bakr, Phosphorylation of thylakoid proteins and synthetic peptide analogs: Differential sensitivity to inhibition by a plastoquinone antagonist (210) 22
Bennett, J. see H.P. Michel (212) 103
Benning, A.D. see R.J. Howland (208) 128
Benosman, H. see J.P. Gayda (217) 57
Bentley, R. see D.S. Feingold (223) 207
Benveniste, J., D. Nunez, P. Duriez, R. Korth, J. Bidault and J.-C. Fruchart, Preformed PAF-acether and lyso PAF-acether are bound to blood lipoproteins (226) 371

Benvenisty, N. see H. Cohen (223) 347
Beppu, T. see K. Petratos (218) 209
Berenguer, J., F. Rojo, M.A. de Pedro, B. Pfanzagl and W. Löffelhardt, Penicillin-binding proteins in the cyanelles of *Cyanophora paradoxa*, a eukaryotic photoautotroph sensitive to β-lactam antibiotics (224) 401
Bereziat, G. see J. Masliah (222) 11
Bergelson, L.D. see G.I. Muzya (220) 371
Berger, M. see J. Champier (212) 220
Bergman, T. see L. Blomqvist (211) 127
Bergman, T. see L. Hederstedt (213) 385
Berkner, K.L. see E. Boel (219) 181
Berlier, Y.M., G.D. Fauque, J. LeGall, P.A. Lespinat and H.D. Peck, jr, The activation of the periplasmic (NiFe) hydrogenase of *Desulfovibrio gigas* by carbon monoxide (221) 241
Berliner, L.J. see J.A.W.H. Vermeulen (219) 426
Berman, J.M., J.T. Pelton, A.D. Cardin, D.T. Blankenship, C.F. Hassman and T.M.M. Chen, In vitro processing of $rANF_{7-28}$-NH_2 by rat kidney homogenates (220) 214
Berndt, N., D.G. Campbell, F.B. Caudwell, P. Cohen, E.F. da Cruz e Silva, O.B. da Cruz e Silva and P.T.W. Cohen, Isolation and sequence analysis of a cDNA clone encoding a type-1 protein phosphatase catalytic subunit: homology with protein phosphatase 2A (223) 340
Bernini, F. see J.L. Lorenzo (218) 77
Berry, M.N., R.B. Gregory, A.R. Grivell, D.C. Henly, J.W. Phillips, P.G. Wallace and G.R. Welch, Linear relationships between mitochondrial forces and cytoplasmic flows argue for the organized energy-coupled nature of cellular metabolism (224) 201
Berta, P. see M. Dillenschneider (208) 413
Berteloot, A. see C. Malo (220) 201
Berthou, J., D. Migliore-Samour, A. Lifchitz, J. Delettré, F. Floc'h and P. Jollès, Immunostimulating properties and three-dimensional structure of two tripeptides from human and cow caseins (218) 55
Bertina, R.M. see H.K. Ploos van Amstel (222) 186
Bertrand, J.R. see C. Malvy (208) 155
Bertrand, J. see J. Gardette (225) 178
Bertrand, P. see J.P. Gayda (217) 57
Bertrand-Burggraf, E., C. Ling, M. Schnarr, J.F. Lefèvre, J. Pouyet and M. Daune, Fast abortive initiation of uvrA promoter in a supercoiled plasmid studied by stopped-flow techniques (215) 83
Beseme, F., M.E. Astruc, R. Defay and A. Crastes de Paulet, Rat liver cytosol oxysterol-binding protein: Characterization and comparison with the HTC cell protein (210) 97
Besemer, J. see E. Pöschl (226) 96
Bessoule, J.-J., R. Lessire, M. Rigoulet, B. Guerin and C. Cassagne, Fatty acid synthesis in mitochondria from *Saccharomyces cerevisiae* (214) 158
Beyreuther, K. see T. Jansen (216) 234
Bezstarosti, K. see A. Montfoort (226) 101
Bhadbhade, M.M., M.J. Adams, T.G. Flynn and H.R. Levy, Sequence identity between a lysine-containing peptide from *Leuconostoc mesenteroides* glucose-6-phosphate dehydrogenase and an active site peptide from human erythrocyte glucose-6-phosphate dehydrogenase (211) 243
Bhattacharyya, B. see S.N. Maity (218) 102
Bhattacharyya, Nisan and P. Roy, Extrachromosomal DNA from a dicot plant *Vigna radiata* (208) 386
Bialojan, C. see A. Takai (217) 81
Bibb, P.C., D.E. Cochrane and N. Morel-Laurens, Loss of quin 2 accompanies degranulation of mast cells (209) 169
Bichler, V. see R. Brandsch (224) 121
Bidard, J.-N. see A. Lombet (219) 355
Bidault, J. see J. Benveniste (226) 371
Bielka, H. see H.-J. Kärgel (220) 126
Biellmann, J.-F. see K.M. Lee (223) 33
Bienert, M. see H. Repke (221) 236
Bienvenue, A. see M. Vidal (216) 159
Biggs, D.R., R. Welle, F.R. Visser and H. Grisebach, Dimethylallylpyrophosphate: 3,9-dihydroxypterocarpan 10-dimethylallyl transferase from *Phaseolus vulgaris*: Identification of the reaction product and properties of the enzyme (220) 223
Bigio, M., R. Rossi, D. Nucci, G. Antoni, R. Rappuoli and G. Ratti, Conformational changes in diphtheria toxoids: Analysis with monoclonal antibodies (218) 271
Bilgin, S. see S.G. Laychock (218) 7
Billat, C. see P. Mayeux (211) 229
Binas, B. and R. Grosse, Demonstration of an epidermal growth factor-dependent 58 kDa phosphoprotein secreted by rat kidney fibroblasts (213) 164
Binder, T. see W. Rosenthal (211) 137
Binoux, M. see P. Hossenlopp (208) 439
Binoux, M. see Y. Le Bouc (222) 181
Bird, T.A., A.J.H. Gearing and J. Saklatvala, Murine interleukin-1 receptor: differences in binding properties between fibroblastic and thymoma cells and evidence for a two-chain receptor model (225) 21
Birdsall, B., J. De Graw, J. Feeney, S. Hammond, M.S. Searle, G.C.K. Roberts, W.T. Colwell and J. Crase, ^{15}N and ^{1}H NMR evidence for multiple conformations of the complex of dihydrofolate reductase with its substrate, folate (217) 106
Birkett, C.R. see K. Gull (219) 31
Birmingham, J. see I.V. Zlatanov (222) 47
Birnbaumer, L. see J. Codina (216) 104
Birnbaumer, L. see W.N. Suki (220) 187
Bissery, V. see C. Gaboriaud (224) 149
Bizzini, B. see X. Rabasseda (213) 337
Bläckberg, L., K.-A. Ängquist and O. Hernell, Bile salt-stimulated lipase in human milk: evidence for its synthesis in the lactating mammary gland (217) 37
Blackmore, R.S., T. Brittain, P.M.A. Gadsby, C. Greenwood and A.J. Thomson, Electron paramagnetic resonance and magnetic circular dichroism studies of a hexa-heme nitrite reductase from *Wolinella succinogenes* (219) 244
Bladier, D. see N.E. Fink de Cabutti (223) 330
Blake, C.C.F. see K. Harlos (224) 97
Blalock, J.E. see D.J.J. Carr (224) 272
Blanck, J. see G.V. Semisotnov (224) 9
Blankenship, D.T. see J.M. Berman (220) 214
Blanpain, J.-P. see L. de Meis (212) 323
Blasi, F. see P.A. Andreasen (209) 213
Blaustein, R.O., W.J. Germann, A. Finkelstein and B.R. DasGupta, The N-terminal half of the heavy chain of botulinum type A neurotoxin forms channels in planar phospholipid bilayers (226) 115
Blaut, M., V. Müller and G. Gottschalk, Proton translocation coupled to methanogenesis from methanol + hydrogen in *Methanosarcina barkeri* (215) 53
Blázquez, E. see J.G. de Diego (222) 256
Blehm, D.J. see G.M. Olins (224) 325
Bloch, G., J.M. Neumann, F. Babin and T. Huynh-Dinh, Sequence-dependence of the conformational changes induced by the 5-methyl cytosine in synthetic RNA oligomers (219) 464
Block, M.A. see J. Covès (208) 401
Bloemendal, H. see C.E.M. Voorter (221) 249
Blomqvist, L., T. Bergman, M. Thelestam and H. Jörnvall, Characterization of domain borders and of a naturally occurring major fragment of staphylococcal α-toxin (211) 127
Bloom, S.R. see Y.C. Lee (220) 243
Blough, H.A. see M.J. Massare (223) 122
Bloxham, D.P. see Y.C. Hsu (218) 1
Blundell, T.L. see H. Jhoti (219) 419

Blundell, T.L. see L.J. Summers (208) 11
Bober, E. see T. Braun (215) 233
Bocckino, S.B., P.B. Wilson and J.H. Exton, Ca^{2+}-mobilizing hormones elicit phosphatidylethanol accumulation via phospholipase D activation (225) 201
Böcher, R. see D. Ankel-Fuchs (213) 123
Bocian, D.F., N.J. Boldt, B.W. Chadwick and H.A. Frank, Near-infrared-excitation resonance Raman spectra of bacterial photosynthetic reaction centers: Implications for path-specific electron transfer (214) 92
Bock, E., K. Edvardsen, A. Gibson, D. Linnemann, J.M. Lyles and O. Nybroe, Characterization of soluble forms of NCAM (225) 33
Bockaert, J. see M. Toutant (215) 339
Bockaert, J. see M. Toutant (222) 51
Bocquet, J. see G. Landemore (209) 299
Bodner, U. see U. Johanningmeier (211) 221
Boehm, S., A 5 S rRNA-like secondary structure in the 7 SL RNA may define a ribosomal binding site of the signal recognition particle (212) 15
Boehm, S., Similarities between a predicted secondary structure for the M1 RNA ribozyme and the tRNA binding center of 16 S rRNA from *E. coli* (220) 283
Boekema, E.J., J.P. Dekker, M.G. van Heel, M. Rögner, W. Saenger, I. Witt and H.T. Witt, Evidence for a trimeric organization of the photosystem I complex from the thermophilic cyanobacterium *Synechococcus* sp. (217) 283
Boekema, E.J. see M. Rögner (219) 207
Boel, E., K.L. Berkner, B.A. Nexø and T.W. Schwartz, Expression of human pancreatic polypeptide precursors from a dicistronic mRNA in mammalian cells (219) 181
Boelens, R. see J.A.W.H. Vermeulen (219) 426
Bogdanov, A.A. see J. Lasch (214) 13
Bogdanov, A.A. see Yu.F. Drygin (215) 247
Bogdanovskaya, V.A. see A.M. Kuznetsov (215) 219
Boldt, N.J. see D.F. Bocian (214) 92
Boldyrev, A.A. see I.A. Svinukhova (214) 335
Bolla, J.-M., C. Lazdunski, M. Inouye and J.M. Pagès, Export and secretion of overproduced OmpA-β-lactamase in *Escherichia coli* (224) 213
Boller, T. see T. Hottiger (220) 113
Boman, H.G. see D.-A. Lidholm (226) 8
Bomsel, M. see C. de Paillerets (219) 113
Bon, S., J.-Y. Chang and A.D. Strosberg, Identical N-terminal peptide sequences of asymmetric forms and of low-salt-soluble and detergent-soluble amphiphilic dimers of *Torpedo* acetylcholinesterase: Comparison with bovine acetylcholinesterase (209) 206
Bonassi, S. see A. De Ambrosis (225) 120
Bondareva, T.O. see A.Yu. Andreyev (226) 265
Bonicel, J. see P. Rouimi (216) 195
Bonser, R.W., J. Dawson, N.T. Thompson, H.F. Hodson and L.G. Garland, Inhibition of phorbol ester stimulated superoxide production by 1-oleoyl-2-acetyl-*sn*-glycerol (OAG); fact or artefact? (209) 134
Bonser, R.W. see J. Dawson (214) 171
Borchart, U. see H. Schägger (219) 161
Borg, J., J. Toazara, H. Hietter, M. Henry, G. Schmitt and B. Luu, Neurotrophic effect of naturally occurring long-chain fatty alcohols on cultured CNS neurons (213) 406
Borghi, H. see D. Chansel (220) 247
Borin, C. see Z. Amelizad (220) 231
Borovikov, Yu.S., M. Wrotek, N.B. Aksenova, N.N. Lebedeva and I. Kakol, Influence of Mg^{2+} and Ca^{2+} bound to 1,5-IAEDANS-labeled phosphorylated and dephosphorylated heavy meromyosin complexed with F-actin on polarized fluorescence of the fluorophore (223) 409
Borrello, S., T. Galeotti, G. Palombini and G. Minotti, Restoration of hydroperoxide-dependent lipid peroxidation by 3-methylcholanthrene induction of cytochrome P-448 in hepatoma microsomes (209) 305
Borsi, L. see B. Carnemolla (215) 269
Borutaitė, V. see B. Kholodenko (223) 247
Boscá, L. see P. Martín-Sanz (225) 37
Bose, N.K. see L.L. Muldrow (213) 249
Bossa, F. see M.E. Schininà (221) 87
Bost, K.L. see D.J.J. Carr (224) 272
Both, V. see S.V. Shlyapnikov (209) 335
Boulain, J.-C. see O. Trémeau (208) 236
Boulter, D. see J.N. Yarwood (222) 175
Boulter, D. see R.N. Waterhouse (209) 223
Bourdeaux-Jaubert, A.M. see F. Bauché (219) 296
Bovermann, G. see A.M. Gronenborn (215) 88
Box, R.-J. and M. Staehelin, Bromoacetylalprenololmenthane binding to β-receptors modulates the rate of hormone-induced internalization but not desensitization in S49 cells (214) 323
Boyer, J.M. see M. Hirasawa (221) 343
Boyer, P.D. see J.G. Wise (223) 395
Boyer, P.D. see Z. Xue (223) 391
Boys, C.W.G. see K. Harlos (224) 97
Bozou, J.-C., A. Couvineau, C. Rouyer-Fessard, M. Laburthe, J.-P. Vincent and P. Kitabgi, Phorbol ester induces loss of VIP stimulation of adenylate cyclase and VIP-binding sites in HT29 cells (211) 151
Bradford, A.P., A. Aitken, F. Beg, K.G. Cook and S.J. Yeaman, Amino acid sequence surrounding the lipoic acid cofactor of bovine kidney 2-oxoglutarate dehydrogenase complex (222) 211
Bradley, R.J., M.K. Pagala and M.T. Edge, Multiple effects of α-toxins on the nicotinic acetylcholine receptor (224) 277
Bradshaw, C. see T.J. Michalski (226) 72
Bragg, P.D. see E.G. Sedgwick (218) 22
Brain, A.P.R. see R.S. Williams (225) 59
Brammar, W.J., I.G. Charles, M. Matfield, L. Cheng-Pin, R.E. Drew and P.H. Clarke, The nucleotide sequence of the *amiE* gene of *Pseudomonas aeruginosa* (215) 291
Brandan, E. and N.C. Inestrosa, Co-solubilization of asymmetric acetylcholinesterase and dermatan sulfate proteoglycan from the extracellular matrix of rat skeletal muscles (213) 159
Brandsch, R. and V. Bichler, Covalent flavinylation of 6-hydroxy-D-nicotine oxidase involves an energy-requiring process (224) 121
Brann, M.R., R.M. Collins and A. Spiegel, Localization of mRNAs encoding the α-subunits of signal-transducing G-proteins within rat brain and among peripheral tissues (222) 191
Brann, M. see P.M. Murphy (221) 81
Braun, H.P., M.E. Michel-Beyerle, J. Breton, S. Buchanan and H. Michel, Electric field effect on absorption spectra of reaction centers of *Rb. sphaeroides* and *Rps. viridis* (221) 221
Braun, T., E. Bober, S. Singh, D.P. Agarwal and H.W. Goedde, Evidence for a signal peptide at the amino-terminal end of human mitochondrial aldehyde dehydrogenase (215) 233
Bravo, R. see T. Lund (208) 369
Breg, J. see J.A. van Kuik (221) 150
Brennan, S.O., P.M. George and R.E. Jordan, Physiological variant of antithrombin-III lacks carbohydrate sidechain at Asn 135 (219) 431
Brenner, S.L. see M. Coué (213) 316
Bresser, J. see S. Dewhurst (213) 138
Breton, J., J.-L. Martin, J. Petrich, A. Migus and A. Antonetti, The absence of a spectroscopically resolved intermediate state P^+B^- in bacterial photosynthesis (209) 37
Breton, J. see H.P. Braun (221) 221

Bretscher, A. see J. Krizek (225) 269
Brettel, K. see S. Gerken (223) 376
Brew, K. see H. Nagase (222) 83
Brewer, jr, H.B. see S.S. Fojo (213) 221
Bridger, W.A. see T. Lin (224) 322
Brindley, D.N. see A.M. Salter (220) 159
Bringmann, P. and R. Lührmann, Antibodies specific for N^6-methyladenosine react with intact snRNPs U2 and U4/U6 (213) 309
Brion, J.-P., M.E. Cheetham, P.A. Robinson, A.-M. Couck and B.H. Anderton, Isolation of cDNAs coding for epitopes shared by microtubule-associated proteins and neurofibrillary tangles in Alzheimer's disease (226) 28
Brittain, T. see R.S. Blackmore (219) 244
Brizard, C. see F. Baklouti (223) 59
Broekaert, W.F., A.K. Allen and W.J. Peumans, Separation and partial characterization of isolectins with different subunit compositions from *Datura stramonium* seeds (220) 116
Brömme, D., A. Steinert, S. Fittkau and H. Kirschke, Action of rat liver cathepsin B on bradykinin and on the oxidized insulin A-chain (219) 441
Bron, C. and H.R. MacDonald, Identification of the plasma membrane receptor for interleukin-1 on mouse thymoma cells (219) 365
Brooks, B.R. see V. Sklenář (216) 249
Brooks, S.F., F.J. Evans and A. Aitken, The stimulation of phosphorylation of intracellular proteins in GH_3 rat pituitary tumour cells by phorbol esters of distinct biological activity (224) 109
Brossi, A., P. Millet, I. Landau, M.E. Bembenek and C.W. Abell, Antimalarial activity and inhibition of monoamine oxidases A and B by exo-erythrocytic antimalarials: Optical isomers of primaquine, *N*-acylated congeners, primaquine metabolites and 5-phenoxy-substituted analogues (214) 291
Brossi, A. see Q.S. Yu (221) 325
Brossi, A., W. Gessner, C.D. Hufford, J.K. Baker, F. Homo, P. Millet and I. Landau, Photooxidation products of primaquine: Structure, antimalarial activity and hemolytic effects (223) 77
Broude, N.E. see E.D. Sverdlov (217) 275
Broude, N.E. see E.D. Sverdlov (221) 129
Broude, N.E. see N.M. Matveeva (217) 42
Broude, N.E. see Yu.A. Ovchinnikov (213) 73
Brown, A.M. see J. Codina (216) 104
Brown, D.A. see H. Higashida (208) 283
Brown, D.A. see H. Higashida (220) 302
Brown, E., P. Enyedi, M. LeBoff, J. Rotberg, J. Preston and C. Chen, High extracellular Ca^{2+} and Mg^{2+} stimulate accumulation of inositol phosphates in bovine parathyroid cells (218) 113
Brown, E. see M. Oetting (208) 99
Brown, J.C.C. see J.D. Bell (215) 311
Brown, R.A. see P.A. MacLennan (215) 187
Browne, M.J. see I. Dodd (209) 13
Bruccoleri, R.E. see J. Novotný (211) 185
Brun, J.-L. see M. Rousset (208) 34
Brunati, A.M. see F. Meggio (215) 241
Brünger, A.T. see G.M. Clore (213) 269
Bruni, P., V. Vasta and M. Farnararo, Adenylate cyclase stimulating agents and mitogens raise fructose 2,6-bisphosphate levels in human fibroblasts: Evidence for a dual control of the metabolite (222) 27
Brunisholz, R.A. see T.D. Wechsler (210) 189
Brunori, M., A. Bellelli, B. Giardina, S. Condo and M.F. Perutz, Is there a Root effect in *Xenopus* hemoglobin? (221) 161
Brunori, M. see G. Falcioni (221) 355
Brunowsky, W. see E. Pöschl (226) 96
Bruns, C. and D. Marmé, Pertussis toxin inhibits the angiotensin II and serotonin-induced rise of free cytoplasmic calcium in cultured smooth muscle cells from rat aorta (212) 40
Bruns, C., C. Schächtele and D. Marmé, Synthetic diacylglycerols induce a rise of quin2-detectable free intracellular calcium in human platelets (221) 23
Brzezinski, P. see M. Fabian (213) 396
Buchanan, S. see H.P. Braun (221) 221
Buchta, R., M. Pontet and M. Fridkin, Binding of C-reactive protein to human neutrophils (211) 165
Buckland, P.R. see J. Furmaniak (215) 316
Buckle, M., Regulation of ATP hydrolase activity of the F_0-F_1 complex of rat-liver mitochondria during early hepatic regeneration (209) 197
Buday, L., J. Seprődi, G. Farkas, G. Mészáros, T. Romhányi, G. Bánhegyi, J. Mandl, F. Antoni and A. Faragó, Proteolytic activation of protein kinase C in the extracts of cells treated for a short time with phorbol ester (223) 15
Buki, K. see A. Hakam (212) 73
Bülow, L. and K. Mosbach, The expression in *E. coli* of a polymeric gene coding for an esterase mimic catalyzing the hydrolysis of *p*-nitrophenyl esters (210) 147
Bülow, R. see F. Jähnig (221) 37
Bump, N.J. and V.A. Najjar, Tuftsin stimulates growth of HL60 cells (226) 303
Bundle, D.R., J.W. Cherwonogrodzky and M.B. Perry, The structure of the lipopolysaccharide O-chain (M antigen) and polysaccharide B produced by *Brucella melitensis* 16M (216) 261
Burbach, J.P.H. see H.P.J.M. Noteborn (216) 200
Burchell, B. see R.B. Corser (213) 448
Burgess, A.J. see R.I. Norman (212) 127
Burgess, A.W. see R.J. Simpson (224) 128
Burgoyne, L.A. see S.J. Davis (226) 88
Burgoyne, R.D. see I.A. Pearce (223) 143
Burgoyne, R.D. see P.H. Cobbold (211) 44
Burman, K.D. see A.P. Weetman (211) 69
Burns, G., A.K. Abraham and A. Vedeler, Nucleotide binding to elongation factor 2 inactivated by diphtheria toxin (208) 217
Burton, G.W. see K.H. Cheeseman (209) 191
Busby, S. see S. Ponnambalam (212) 21
Busby, S. see S. Ponnambalam (219) 189
Bushfield, M., S.L. Hopple, I.F. Gibson, F.A. Murdoch and D.E. MacIntyre, Effects of protein kinase C activation on human platelet cyclic AMP metabolism (222) 299
Bushueva, T.L. and A.G. Tonevitsky, The effect of pH on the conformation and stability of the structure of plant toxin – ricin (215) 155
Buyssens, S. see H. Höfte (226) 364
Byfield, P.G.H. see J. Worthington (211) 123
Bygrave, F.L. see C. Preston (210) 27
Bygrave, F.L. see P. Dieter (213) 174
Bystrova, M.F. see E.E. Fesenko (219) 224
Bystrov, V.F. see A.S. Arseniev (213) 283

C

Cacan, R. see R. Cecchelli (208) 407
Camara, B. see O. Dogbo (210) 211
Cambray-Deakin, M.A. see I.A. Pearce (223) 143

Campbell, D.G. see N. Berndt (223) 340
Campbell, D.G. see O.B. da Cruz e Silva (221) 415
Campbell, S.F. see J. Singh (224) 161
Camus, J. see M. Waelbroeck (226) 287
Cancelli, A. see P. Conti (225) 103
Candau, P. see F.J. Florencio (223) 37
Cannon, B. see A. Jacobsson (224) 353
Cantatore, P., Z. Flagella, F. Fracasso, A.M.S. Lezza, M.N. Gadaleta and A. de Montalvo, Synthesis and turnover rates of four rat liver mitochondrial RNA species (213) 144
Canters, G.W., The azurin gene from *Pseudomonas aeruginosa* codes for a pre-protein with a signal peptide: Cloning and sequencing of the azurin gene (212) 168
Cantin, M. see J. Gutkowska (214) 17
Cantournet, B., C. Creuzet, O. Komano and J. Loeb, Clathrin β-light chain of rat liver coated vesicles is phosphorylated in vitro and in vivo (220) 143
Capaldi, R.A. see E.P. Gogol (219) 274
Capaldi, R.A. see S. Takamiya (218) 277
Capiod, T., A.C. Field, D.C. Ogden and C.A. Sandford, Internal perfusion of guinea-pig hepatocytes with buffered Ca^{2+} or inositol 1,4,5-trisphosphate mimics noradrenaline activation of K^+ and Cl^- conductances (217) 247
Carchman, R.A. see C.F. Strnad (225) 16
Cardin, A.D. see J.M. Berman (220) 214
Cardinaud, R., Proteolysis rates of a myosin heavy chain site with papain: Evidence for a combined LC2-filament-mediated mechanism (220) 376
Caretta, A. and P.J. Stein, Light- and nucleotide-dependent increase in apparent viscosity in a suspension of retinal disks (219) 97
Carles, C., P. Gueguen and B. Ribadeau-Dumas, C-terminal labelling of β-casein (212) 163
Carling, D., V.A. Zammit and D.G. Hardie, A common bicyclic protein kinase cascade inactivates the regulatory enzymes of fatty acid and cholesterol biosynthesis (223) 217
Carloni, G. see M. Clementi (221) 11
Carlstedt, I. see H. Karlsson (226) 23
Carmena, M.J., J.I. Sancho, M.A. Fernández-Gonzalez, F. Escudero and J.C. Prieto, Somatostatin inhibits VIP- and isoproterenol-stimulated cyclic AMP accumulation in rat prostatic epithelial cells (218) 73
Carne, A. see S.M. Cutfield (214) 57
Carnemolla, B., L. Borsi, L. Zardi, R.J. Owens and F.E. Baralle, Localization of the cellular-fibronectin-specific epitope recognized by the monoclonal antibody IST-9 using fusion proteins expressed in *E. coli* (215) 269
Caro, L.H.P. see X.M. Leverve (219) 455
Caron, M. see N.E. Fink de Cabutti (223) 330
Carper, D., C. Nishimura, T. Shinohara, B. Dietzchold, G. Wistow, C. Craft, P. Kador and J.H. Kinoshita, Aldose reductase and ρ-crystallin belong to the same protein superfamily as aldehyde reductase (220) 209
Carr, D.J.J., B. DeCosta, A.E. Jacobson, K.L. Bost, K.C. Rice and J.E. Blalock, Immunoaffinity-purified opiate receptor specifically binds the δ-class opiate receptor ligand, *cis*-(+)-3-methylfentanylisothiocyanate, SUPERFIT (224) 272
Carrasco, L. see E. Feduchi (214) 153
Carreras, J. see C. Espinet (209) 254
Carreras, J. see C. Gallego (222) 167
Carroll, M.C. see B. Uring-Lambert (217) 65
Carvalho, J.F. see E. Rondinelli (208) 379
Carvallo, P. and J.E. Allende, Interaction of protein synthesis initiation factor 2 from *Xenopus laevis* oocytes with GDP and GTP analogs (215) 109
Casas-Finet, J.R., M.I. Khamis, A.H. Maki and J.W. Chase, Tryptophan 54 and phenylalanine 60 are involved synergistically in the binding of *E. coli* SSB protein to single-stranded polynucleotides (220) 347
Casas-Finet, J.R. see M.I. Khamis (211) 155
Cascales, M. see P. Martín-Sanz (225) 37
Cash, D.J. and K. Subbarao, Two desensitization processes of GABA receptor from rat brain: Rapid measurements of chloride ion flux using quench-flow techniques (217) 129
Casley, D.J. see A.E. Fletcher (208) 263
Casnellie, J.E. see R.E. Thom (222) 104
Cass, A.E.G., D.W. Ribbons, J.T. Rossiter and S.R. Williams, Biotransformation of aromatic compounds: Monitoring fluorinated analogues by NMR (220) 353
Cassagne, C. see J.-J. Bessoule (214) 158
Castagna, M. see F. Heymans (218) 35
Castellani, A.A. see M. Galliano (208) 364
Castignetti, D. and J. Smarrelli, jr, Siderophores, the iron nutrition of plants, and nitrate reductase (209) 147
Castle, N.A. and P.N. Strong, Identification of two toxins from scorpion *(Leiurus quinquestriatus)* venom which block distinct classes of calcium-activated potassium channel (209) 117
Cates, G.A., D.W. Litchfield, S. Narindrasorasak, D. Nandan, E.H. Ball and B.D. Sanwal, Phosphorylation of a gelatin-binding protein from L6 myoblasts by protein kinase C (218) 195
Caudwell, F.B. see N. Berndt (223) 340
Cavaggioni, A., R.T. Sorbi, J.N. Keen, D.J.C. Pappin and J.B.C. Findlay, Homology between the pyrazine-binding protein from nasal mucosa and major urinary proteins (212) 225
Cavieres, J.D., The molecular size required varies according to the reaction step round the sodium pump cycle (225) 145
Cawston, T.E. and E. Mercer, Preferential binding of collagenase to α_2-macroglobulin in the presence of the tissue inhibitor of metalloproteinases (209) 9
Cecchelli, R., R. Cacan and A. Verbert, Mechanism of UDP-sugar transport into intracellular vesicles: Occurrence of UDP-GlcNAc/UDP and UDP-Gal/UDP antiports (208) 407
Cederbaum, A.I. see G. Krikun (208) 292
Cedergren, R., B.F. Lang and D. Gravel, A mechanism for the RNA-catalyzed formation of 5′-phosphates: The origin of nucleases (226) 63
Cederholm-Williams, S.A. see R.J.P. Williams (209) 111
Celis, A. see J.E. Celis (220) 1
Celis, A. see J.E. Celis (223) 237
Celis, J.E. and P. Madsen, Increased nuclear cyclin/PCNA antigen staining of non S-phase transformed human amnion cells engaged in nucleotide excision DNA repair (209) 277
Celis, J.E., G.P. Ratz and A. Celis, Progressin: a novel proliferation-sensitive and cell cycle-regulated human protein whose rate of synthesis increases at or near the G_1/S transition border of the cell cycle (223) 237
Celis, J.E., P. Madsen, A. Celis, H.V. Nielsen and B. Gesser, Cyclin (PCNA, auxiliary protein of DNA polymerase δ) is a central component of the pathway(s) leading to DNA replication and cell division (220) 1
Cerf, R. see D. Rogez (219) 22
Chackalaparampil, I. see G.R. Laverdure (222) 261
Chader, G.J. see T. van Veen (208) 133
Chadwick, B.W. see D.F. Bocian (214) 92
Chalcroft, J.P., H. Engelhardt and W. Baumeister, Structure of the porin from a bacterial stalk (211) 53
Chambaz, E. see J. Fauvel (221) 397
Chambaz, J. see J. Masliah (222) 11

Champier, J., B. Claustrat, G. Sassolas and M. Berger, Detection and enzymatic deglycosylation of a glycosylated variant of prolactin in human plasma (212) 220
Champness, J.N. see D.K. Stammers (218) 178
Chan, C.-T.J. see J. Worthington (211) 123
Chan, L. see C. Yang (224) 261
Chance, B., Early reduction of cytochrome *c* in hypoxia (226) 343
Chance, B. see M. Engelhard (222) 275
Chance, M. see M. Engelhard (222) 275
Chandrasekhar, B., Mrigank and V. Kothekar, On the possible mechanism of recognition of DNA base sequence by steroid hormones (225) 151
Chang, C. see M. Clementi (221) 11
Chang, J.-Y. see S. Bon (209) 206
Chang, J.-Y. see V. Steiner (222) 6
Chang, P.L. see M. Ameen (219) 130
Chang, W.-P. see S.-H. Chiou (221) 134
Chang, W. see N.J. Philp (225) 127
Chansel, D., J.-P. Morin, H. Borghi, N. Ardaillou and R. Ardaillou, Angiotensin I-converting enzyme in isolated human glomeruli (220) 247
Chao, C.C.-K. and S. Lin-Chao, Loss of inducible photorepair in a frog cell line hypersensitive to solar UV light (225) 133
Chap, H. see J. Fauvel (216) 45
Chap, H. see J. Fauvel (221) 397
Chapman, B.E. see P.W. Kuchel (219) 5
Chapman, D.J. see J. Barber (220) 67
Chapman, K. see A.J. Harmar (208) 67
Charles, I.G. see W.J. Brammar (215) 291
Charnay, Y. see J.-M. Zajac (216) 118
Chase, J.W. see J.R. Casas-Finet (220) 347
Chase, J.W. see M.I. Khamis (211) 155
Chauvet, J., M.-T. Chauvet and R. Acher, Conformation limited proteolysis in the common neurophysin-copeptin precursor shown by trypsin-Sepharose chromatographic proteolysis (217) 180
Chauvet, J. see M.T. Chauvet (210) 40
Chauvet, M.-T. see J. Chauvet (217) 180
Chauvet, M.T., Y. Rouillé, J. Chauvet and R. Acher, Guinea pig neurohypophysial hormones: Peculiar processing of the three-domain vasopressin precursor (210) 40
Chazin, W.J., M. Rance and P.E. Wright, Complete assignment of lysine resonances in ^{1}H NMR spectra of proteins as probes of surface structure and dynamics (222) 109
Che, Y.W. see F. Guerrieri (213) 67
Cheek, T.R. see P.H. Cobbold (211) 44
Cheeseman, K.H., M. Collins, S. Maddix, A. Milia, K. Proudfoot, T.F. Slater, G.W. Burton, A. Webb and K.U. Ingold, Lipid peroxidation in regenerating rat liver (209) 191
Cheetham, M.E. see J.-P. Brion (226) 28
Chekalin, S.V., Y.A. Matveetz, A.Y. Shkuropatov, V.A. Shuvalov and A.P. Yartzev, Femtosecond spectroscopy of primary charge separation in modified reaction centers of *Rhodobacter sphaeroides* (R-26) (216) 245
Chen, A. and S.S. Wong, Differentiation of creatine kinase MB and IgA-linked BB isoenzymes on electrophoresis (214) 192
Chen, C. see E. Brown (218) 113
Chen, H.Y. see R.C. Gorewit (225) 238
Chen, K.-X., N. Gresh, B. Pullman, Groove selectivity in the interaction of 9-aminoacridine-4-carboxamide antitumor agents with DNA (224) 361
Chen, S.-W. see S.-H. Chiou (209) 107
Chen, S.-W. see S.-H. Chiou (221) 134
Chen, T.M.M. see J.M. Berman (220) 214
Chen, Z.G. see Y.X. Zhu (208) 253
Chen, Z., V. Mutt, J. Barros-Söderling and H. Jörnvall, Isolation and structural characterization of porcine coupling factor 6 from intestinal tissues (226) 43
Cheng, C.Y. see G.L. Hammond (215) 100
Cheng-Pin, L. see W.J. Brammar (215) 291
Chernousov, M.A., A.I. Faerman, M.G. Frid, O.Yu. Printseva and V.E. Koteliansky, Monoclonal antibody to fibronectin which inhibits extracellular matrix assembly (217) 124
Chernov, B.K. see S.Yu. Morozov (213) 438
Chernov, I.P. see E.D. Sverdlov (212) 233
Chernyak, B.V., V.F. Dukhovich and E.Yu. Khodjaev, The effect of the natural protein inhibitor on H^+-ATPase in hepatoma 22^a mitochondria (215) 300
Cherwonogrodzky, J.W. see D.R. Bundle (216) 261
Chetverikov, A.G. see S.A. Dikanov (224) 75
Chhatwal, G.S. see K. Aktories (212) 109
Chhatwal, G.S. see S. Rösener (224) 38
Chiang, P.K. see A. Ahmad (214) 285
Chiarugi, V. see G. Laffi (220) 217
Chiba, T. see A. Hata-Tanaka (214) 279
Chidgeavadze, Z. see N. Dyatkina (219) 151
Childs, R.A. see C. Southan (214) 301
Chindemi, P.A. see E. Regoeczi (222) 271
Chino, N. see Y. Hirata (219) 369
Chiou, S.-H., S.-W. Chen and T.-B. Lo, The amino-terminal sequences of four major carp γ-crystallin polypeptides and their homology with frog and calf γ-crystallins (209) 107
Chiou, S.-H., W.-P. Chang, C.-H. Lo and S.-W. Chen, Sequence comparison of γ-crystallins from the reptilian and other vertebrate species (221) 134
Chiva, M., H.F. Kasinsky and J.A. Subirana, Characterization of protamines from four avian species (215) 237
Cho-Chung, Y.S. see D. Katsaros (223) 97
Cho-Chung, Y.S. see T. Clair (224) 377
Chomutov, R.M. see J. Šimúth (218) 163
Chonn, A. see T.M. Allen (223) 42
Chrétien, M. see N.G. Seidah (211) 144
Chretien, M. see S. Benjannet (224) 142
Christ, B. and K. Jungermann, Sub-compartmentation of the 'cytosolic' glucose 6-phosphate pool in cultured rat hepatocytes (221) 375
Christie, D.L., K.M. Dziegielewska, R.M. Hill and N.R. Saunders, Fetuin: the bovine homologue of human α_2HS glycoprotein (214) 45
Christophe, J. see J.-P. Dehaye (219) 451
Christophe, J. see M. Waelbroeck (226) 287
Chroneos, Z.C. see Q.A. Vu (220) 79
Chumanov, G.D. see N.G. Abdulaev (213) 113
Chung, F.-Z., K.-U. Lentes, J. Gocayne, M. Fitzgerald, D. Robinson, A.R. Kerlavage, C.M. Fraser and J.C. Venter, Cloning and sequence analysis of the human brain β-adrenergic receptor: Evolutionary relationship to rodent and avian β-receptors and porcine muscarinic receptors (211) 200
Cid, A. see P. Eraso (224) 193
Cid, A. see R. Serrano (208) 143
Cincolà, G. see G. Falcioni (221) 355
Clair, T., S. Ally, P. Tagliaferri, R.K. Robins and Y.S. Cho-Chung, Site-selective cAMP analogs induce nuclear translocation of the R^a cAMP receptor protein in Ha-MuSV-transformed NIH/3T3 cells (224) 377
Clair, T. see D. Katsaros (223) 97
Clark, E.A. see M. Patarroyo (210) 127
Clark, jr, W.H. see J. Thurin (208) 17
Clark, M.G. see E.A. Richter (217) 232
Clarke, C. see A. Stock (220) 8
Clarke, P.H. see R.P. Ambler (215) 285
Clarke, P.H. see W.J. Brammar (215) 291
Clarke, S. see A. Stock (220) 8
Claustrat, B. see J. Champier (212) 220
Clayton, C.L. see B.W. Wren (225) 82
Cleland, P.J.F. see E.A. Richter (217) 232

Clemance, M. see J.-P. Tenu (220) 93
Clementi, M., I. Testa, A. Festa, P. Bagnarelli, C. Chang and G. Carloni, Differential response of the human hepatoma-derived cell line HA22T/VGH to polypeptide mitogens (221) 11
Clore, G.M., M. Nilges, A.T. Brünger, M. Karplus and A.M. Gronenborn, A comparison of the restrained molecular dynamics and distance geometry methods for determining three-dimensional structures of proteins on the basis of interproton distances (213) 269
Clore, G.M. see A.M. Gronenborn (215) 88
Clore, G.M. see H.R. MacDonald (209) 295
Clore, G.M. see M. Nilges (219) 11
Clore, G.M. see P. Wingfield (215) 160
Clouse, J.A. see D.P. Ringer (224) 59
Clugston, C.K. see L.K. Barnett (224) 287
Coates, A.R.M. see M.J.E. Sternberg (218) 231
Cobbold, P.H., T.R. Cheek, K.S.R. Cuthbertson and R.D. Burgoyne, Calcium transients in single adrenal chromaffin cells detected with aequorin (211) 44
Cochrane, D.E. see P.C. Bibb (209) 169
Codina, J., D. Grenet, A. Yatani, L. Birnbaumer and A.M. Brown, Hormonal regulation of pituitary GH_3 cell K^+ channels by G_k is mediated by its α-subunit (216) 104
Codina, J. see W.N. Suki (220) 187
Cohen, H., N. Benvenisty and L. Reshef, Fate of polyoma origin of replication after its direct introduction into mice (223) 347
Cohen, J. see H.L. Sham (220) 299
Cohen, M.E. see G.W.G. Sharp (221) 309
Cohen, P. see N. Berndt (223) 340
Cohen, P.T.W. see E.F. da Cruz e Silva (220) 36
Cohen, P.T.W. see N. Berndt (223) 340
Cohen, P.T.W. see O.B. da Cruz e Silva (221) 415
Cohen, P.T.W. see O.B. da Cruz e Silva (226) 176
Cohen, R. see A. Ashkenazi (210) 51
Coll, M.J., J. Serratosa, O. Bachs, C.G. Gahmberg and C. Enrich, Calmodulin may decrease cell surface sialic acid and be involved in the expression of fibronectin during liver regeneration (208) 418
Collazos, J.M. and A. Sanchez, cAMP reduces the affinity of Ca^{2+}-triggered secretion in platelets (215) 183
Collins, J.H. see J.P. Rieker (212) 154
Collins, J.H. see J.P. Rieker (223) 262
Collins, M. see K.H. Cheeseman (209) 191
Collins, R.M. see M.R. Brann (222) 191
Colom, J. see J.A. Subirana (220) 193
Colonna, R. see E. Papini (215) 73
Colson, A.M. see J.-P. di Rago (208) 208
Colton, C.A. and D.L. Gilbert, Production of superoxide anions by a CNS macrophage, the microglia (223) 284
Coluccio, L.M. see J. Krizek (225) 269
Colwell, W.T. see B. Birdsall (217) 106
Comera, C. see A.J. Aarsman (219) 176
Comera, C. see B. Rothhut (219) 169
Cominelli, F. see G. Laffi (220) 217
Condo, S. see M. Brunori (221) 161
Conlon, J.M., E. Dafgård, S. Falkmer and L. Thim, The primary structure of ratfish insulin reveals an unusual mode of proinsulin processing (208) 445
Conlon, J.M., L. O'Toole and L. Thim, Primary structure of glucagon from the gut of the common dogfish (*Scyliorhinus canicula*) (214) 50
Conquet, F. see J.-P. Lavergne (216) 83
Consalvi, V. see R. Scandurra (212) 79
Conti, P., M. Reale, A. Cancelli and P.U. Angeletti, Lipoxin A augments release of thromboxane from human polymorphonuclear leukocyte suspensions (225) 103
Cook, K.G. see A.P. Bradford (222) 211
Cookson, J. see M.J.E. Sternberg (218) 231
Coomber, S.A. see M.K. Ashby (213) 245
Cooney, D. see A.K. Keenan (217) 287
Cooper, C.E. and P. Nicholls, Activity of proteoliposomes containing cytochrome oxidase in the submitochondrial orientation (223) 155
Cooper, C.S. see P.R. Tempest (209) 357
Cooper, D.R., C.M. de Ruiz Galaretta, L.F. Fanjul, L. Mojsilovic, M.L. Standaert, R.J. Pollet and R.V. Farese, Insulin but not phorbol ester treatment increases phosphorylation of vinculin by protein kinase C in BC3H-1 myocytes (214) 122
Corkey, B.E. see M. Prentki (220) 103
Cornish, A. see R.P. Mason (216) 4
Coronado, R. see H.H. Valdivia (226) 280
Corps, A.N. see G. Yonuschot (213) 401
Corser, R.B., M.W.H. Coughtrie, M.R. Jackson and B. Burchell, The molecular basis of the inherited deficiency of androsterone UDP-glucuronyltransferase in Wistar rats (213) 448
Corsini, A. see J.L. Lorenzo (218) 77
Cortes, R. see S. Pin (208) 325
Costa, T. see F.-J. Klinz (224) 43
Costa, T. see Y. Shimohigashi (222) 71
Cotton, N.P.J., J.F. Myatt and J.B. Jackson, The dependence of the rate of transhydrogenase on the value of the protonmotive force in chromatophores from photosynthetic bacteria (219) 88
Couck, A.-M. see J.-P. Brion (226) 28
Couderc, J. see O. Trémeau (208) 236
Coué, M., S.L. Brenner, I. Spector and E.D. Korn, Inhibition of actin polymerization by latrunculin A (213) 316
Coughtrie, M.W.H. see R.B. Corser (213) 448
Coulaud, D. see J.-F. Riou (213) 304
Courtney, M. see L.H. Tessier (208) 183
Couvineau, A. see J.-C. Bozou (211) 151
Covès, J., M.A. Block, J. Joyard and R. Douce, Solubilization and partial purification of UDP-galactose:diacylglycerol galactosyltransferase activity from spinach chloroplast envelope (208) 401
Covington, M. see J.J. Huang (223) 294
Cox, B.S. see M.F. Tuite (225) 205
Cox, R.A. see A.M. MacNicol (221) 48
Cox, R.A. see M.J. Monteiro (217) 260
Crabbe, M.J.C. see J.E. Baldwin (214) 357
Craft, C. see D. Carper (220) 209
Cragoe, jr, E.J. see L. Hunyady (225) 72
Craig, S. see P. Wingfield (211) 179
Crase, J. see B. Birdsall (217) 106
Crasnier, M., A method to exchange alkali light chains on myosin subfragment 1 (211) 31
Crastes de Paulet, A. see F. Beseme (210) 97
Crawford, N. see K.S. Authi (213) 95
Crema, A.L. see C. Delfini (210) 17
Crepin, K.M. see M.I. Darville (224) 317
Cressent, M. see S. Minvielle (223) 63
Creuzet, C. see B. Cantournet (220) 143
Criado, M. and B.U. Keller, A membrane fusion strategy for single-channel recordings of membranes usually non-accessible to patch-clamp pipette electrodes (224) 172
Crichton, R.R. see Y. Jin (215) 41
Crielaard, W., M.G.L. Elferink, K.J. Hellingwerf, W.N. Konings, J.F. Myatt and J.B. Jackson, Concurrent measurements of the inhibition of ATP synthesis and alanine transport in intact cells of *Rhodobacter capsulatus* (225) 6
Critchley, C. see C. Preston (210) 27
Crochet, J. see C. Kryceve-Martinerie (214) 81
Crofts, A.R. see G. Venturoli (219) 477
Cross, K.E. see R.A. Cross (219) 306

Cross, R.A., K.E. Cross and J.V. Small, Salt dependent dimerisation of caldesmon (219) 306
Croy, R.R.D. see J.N. Yarwood (222) 175
Cryer, G.D. see C.E. Dempsey (218) 173
Crystal, R.G. see L.-H. Tessier (208) 183
Cullum, J. see M. Young (212) 10
Curi, R. see B. Leighton (225) 93
Curran, B. see P.W. Piper (220) 177
Curtis, A. see A.J. Harmar (208) 67
Cusinato, F. see E. Papini (215) 73
Cutfield, J.F. see S.M. Cutfield (214) 57
Cutfield, S.M., A. Carne and J.F. Cutfield, The amino-acid sequences of sculpin islet somatostatin-28 and peptide YY (214) 57
Cuthbertson, K.S.R. see P.H. Cobbold (211) 44
Czichos, J. see B. Ehlers (225) 53
Czyrski, J.A. and W. Gawlikowski, Activity of human, bovine and porcine platelet-derived growth factor in a radioreceptor assay with human placental membrane protein (219) 331

D

Dabadie, P., P. Bendriss, P. Erny and J.-P. Mazat, Uncoupling effects of local anesthetics on rat liver mitochondria (226) 77
Dabrowski, J. see E. Romanowska (211) 175
Da Cruz e Silva, E.F. and P.T.W. Cohen, Isolation and sequence analysis of a cDNA clone encoding the entire catalytic subunit of phosphorylase kinase (220) 36
Da Cruz e Silva, E.F. see N. Berndt (223) 340
Da Cruz e Silva, O.B. and P.T.W. Cohen, A second catalytic subunit of type-2A protein phosphatase from rabbit skeletal muscle (226) 176
Da Cruz e Silva, O.B., S. Alemany, D.G. Campbell and P.T.W. Cohen, Isolation and sequence analysis of a cDNA clone encoding the entire catalytic subunit of a type-2A protein phosphatase (221) 415
Da Cruz e Silva, O.B. see N. Berndt (223) 340
Dacremont, G. see C. Van den Branden (222) 21
Dafgård, E. see J.M. Conlon (208) 445
Daiyasu, H. see S. Koyama (209) 265
Dalrymple, B., The phage Mu repressor c and IS30 transposase proteins are significantly related (208) 7
Dalvit, C., L. Tennant and P.E. Wright, ^{1}H resonances of proximal histidine in CO complexes of hemoglobins provide a sensitive probe of coordination geometry (213) 289
Daly, D.T. see D. Ukena (215) 203
Daly, J.W. see D. Ukena (209) 122
Daly, J.W. see D. Ukena (215) 203
Daly, J.W. see K.A. Jacobson (225) 97
Damante, G., S. Filetti and B. Rapoport, The functional activity of the rat c-Ha-ras promoter requires the coordinate involvement of multiple elements (225) 264
Damen, H.C.M. see D.H. Joziasse (221) 139
Danchin, A. see G. Lenzen (219) 254
Dancshazy, Z., R. Govindjee, B. Nelson and T.G. Ebrey, A new intermediate in the photocycle of bacteriorhodopsin (209) 44
Dandona, P. see B. Pearce (211) 73
Danielius, R.V., K. Satoh, P.J.M. van Kan, J.J. Plijter, A.M. Nuijs and H.J. van Gorkom, The primary reaction of photosystem II in the D1–D2–cytochrome *b*-559 complex (213) 241
Dann, J.G. see D.K. Stammers (218) 178
Danø, K. see P.A. Andreasen (209) 213
Danpure, C.J. see P.J. Wise (222) 17
Danson, M.J. see L.D. Smith (225) 277
Dargent, B., J. Rosenbusch and F. Pattus, Selectivity for maltose and maltodextrins of maltoporin, a pore-forming protein of *E. coli* outer membrane (220) 136
Darszon, A. see A. Guerrero (220) 295
Darszon, A. see M. González-Martínez (218) 247
Darville, M.I., K.M. Crepin, J. Vandekerckhove, J. Van Damme, J.N. Octave, M.H. Rider, M.J. Marchand, L. Hue and G.G. Rousseau, Complete nucleotide sequence coding for rat liver 6-phosphofructo-2-kinase/fructose-2,6-bisphosphatase derived from a cDNA clone (224) 317
Das, M.R. see A.N. Hegde (217) 74
DasGupta, B.R. see R.O. Blaustein (226) 115
Dasgupta, D. see P. Parrack (212) 297
Daumas, P. see Y. Trudelle (216) 11
Daune, M. see E. Bertrand-Burggraf (215) 83
Davidson, J.S. see C.E. Smith (225) 247
Davies, G.J. see J.A. Littlechild (225) 123
Davies, W. see P.W. Piper (220) 177
Davis, D.J. see M. Hirasawa (221) 343
Davis, E.J. see P.W.D. Scislowski (224) 177
Davis, G. see P. Nicotera (209) 139
Davis, J.S., L.A. West, L.L. Weakland and R.V. Farese, Human chorionic gonadotropin activates the inositol 1,4,5-trisphosphate-Ca^{2+} intracellular signalling system in bovine luteal cells (208) 287
Davis, S.J. and L.A. Burgoyne, The DNase I generated disomal series is coherent to 16*N*: Implications for coiling models of chromatin structure (226) 88
Dawson, J., N.T. Thompson, R.W. Bonser, H.F. Hodson and L.G. Garland, Decrease of cellular ATP by dihexanoylglycerol may limit responses to protein kinase C activation (214) 171
Dawson, J. see R.W. Bonser (209) 134
Dawson, J. see R.W. Randall (214) 167
Day, I.N.M. and R.J. Thompson, Molecular cloning of cDNA coding for human PGP 9.5 protein: A novel cytoplasmic marker for neurones and neuroendocrine cells (210) 157
Day, I.N.M., M.T.E.P. Allsopp, D.C.M. Moore and R.J. Thompson, Sequence conservation in the 3′-untranslated regions of neurone-specific enolase, lymphokine and protooncogene mRNAs (222) 139
De Almeida Soares, C.M. see E. Rondinelli (208) 379
De Ambrosis, A., N. Ferrari, S. Bonassi and G. Vidali, Nucleosomal repeat length in active and inactive genes (225) 120
De Blasi, A. see M. Fratelli (212) 149
De Block, J. and W. De Potter, The cell free interaction between chromaffin granules and plasma membranes: An in vitro model for exocytosis? (222) 358
De Boer, E. see R. Wever (216) 1
De Bruijn, S.M. see J.J.S. van Rensen (226) 347
De Caro, A. see P. Rouimi (216) 195
De Chaffoy de Courcelles, D., P. Roevens, H.Van Belle and F.De Clerck, The synergistic effect of serotonin and epinephrine on the human platelet at the level of signal transduction (219) 283
De Clerck, F. see D. de Chaffoy de Courcelles (219) 283
De Diego, J.G., E. Alvarez and E. Blázquez, Characterization of glucacon receptors in Golgi fractions of fetal rat liver (222) 256
De Foresta, B., M. Rogard and J. Gallay, Adenylate cyclase of bovine adrenal cortex plasma membranes: Divergence between corticotropin and fluoride combined effects with forskolin (216) 107
De Graw, J. see B. Birdsall (217) 106
De Haas, G.H. see S.-Y. Mao (211) 83

De Jong, W.W. see C.E.M. Voorter (221) 249
De Jong-Brink, M. see D.H. Joziasse (221) 139
De Kok, A. and A.J.W.G. Visser, Flavin binding site differences between lipoamide dehydrogenase and glutathione reductase as revealed by static and time-resolved flavin fluorescence (218) 135
De Kroon, T. see J. Tommassen (221) 226
De Kruijff, B. see A.M. Batenburg (223) 148
De Kruijff, B. see K. Nicolay (209) 33
De Meis, L., Effects of organic solvents and orthophosphate on the ATPase activity of F1 ATPase (213) 333
De Meis, L., J.-P. Blanpain and A. Goffeau, $P_i \rightleftharpoons$ATP exchange in the absence of proton gradient by the H^+-ATPase from yeast plasma membranes (212) 323
De Meis, L. see V.L. Goncalves de Moraes (222) 163
De Montalvo, A. see P. Cantatore (213) 144
De Moura-Neto, R.S. see E. Rondinelli (208) 379
De Narco, A. see J.A.W.H. Vermeulen (219) 426
De Nobel, H. see W. Homan (215) 323
De Pagter-Holthuizen, P., M. Jansen, F.M.A. van Schaik, R. van der Kammen, C. Oosterwijk, J.L. Van den Brande and J.S. Sussenbach, The human insulin-like growth factor II gene contains two development-specific promoters (214) 259
De Pagter-Holthuizen, P. see J.W.M. Höppener (215) 122
De Paillerets, C., M. Bomsel, H. Weintraub, D. Pépin and A. Alfsen, Clustering in coated vesicles of polyunsaturated phospholipids segregated from plasma and Golgi membranes of adrenocortical cells (219) 113
De Pedro, M.A. see J. Berenguer (224) 401
De Potter, W. see J. De Block (222) 358
De Ruiz Galaretta, C.M. see D.R. Cooper (214) 122
De Vellis, J. see S. Kumar (208) 151
De Wachter, R. see H. Van den Eynde (213) 301
De Wachter, R. see H. Van den Eynde (217) 191
De Wit, R.J.W. see R.J. Aerts (220) 366
De Young, M.B. and A. Scarpa, Extracellular ATP induces Ca^{2+} transients in cardiac myocytes which are potentiated by norepinephrine (223) 53
Dean, R.T., A mechanism for accelerated degradation of intracellular proteins after limited damage by free radicals (220) 278
Dean, R.T. see G.S. Vince (216) 253
Deber, C.M. see G.A. Woolley (224) 337
DeCosta, B., C. George, R.B. Rothman, A.E. Jacobson and K.C. Rice, Synthesis and absolute configuration of optically pure enantiomers of a κ-opioid receptor selective agonist (223) 335
DeCosta, B. see D.J.J. Carr (224) 272
Decottignies, P. see J.-P. Jacquot (209) 87
Dedukhova, V.I. see A.Yu. Andreyev (226) 265
Dees, C. see A.K. Keenan (217) 287
Defay, R. see F. Beseme (210) 97
Dehaye, J.-P., J. Winand and J. Christophe, Scorpion venom inhibits carbamylcholine-induced ^{86}rubidium efflux from rat parotid acini (219) 451
Dekker, J.P. see E.J. Boekema (217) 283
Dekker, J.P. see I. Witt (221) 260
Dekker, J.P. see M. Rögner (219) 207
Del Rey, F. see A.R. Nebreda (220) 27
Delaage, M. see A.M. Garrigues (224) 267
Delain, E. see F. Pochon (217) 101
Delain, E. see J. Dexpert (225) 223
Delamourd, L. see A. Ghazi (209) 325
Delaunay, J. see F. Baklouti (223) 59
Delauney, A. see J.N. Yarwood (222) 175
Delcros, J.G., A.M. Roch, V. Thomas, S. El Alaoui, J.P. Moulinoux and G. Quash, Protein-bound polyamines in the plasma of mice grafted with the Lewis lung carcinoma (220) 236
Delehaye, M.C. see S. Minvielle (223) 63
Delettré, J. see J. Berthou (218) 55
Delfini, C., A.L. Crema, E. Alfani, E. Lo Presti, T. Eremenko and P. Volpe, DNA methylases separated through the HeLa cell cycle methodology show allosteric properties (210) 17
Deli, E. see Z. Kiss (213) 365
Deli, E., Z. Kiss, E. Wilson, J.D. Lambeth and J.F. Kuo, Immunocytochemical localization of protein kinase C in resting and activated human neutrophils (221) 365
Delihas, N., Unusual 5 S ribosomal RNAs: An analysis of individual segments can reveal phylogenetic relatedness (221) 189
Demeneix, B. and N.J. Grant, α-Melanocyte stimulating hormone promotes neurite outgrowth in chromaffin cells (226) 337
Dementiev, A.A. see S.V. Shlyapnikov (209) 335
Demeter, S., P.J. Neale and A. Melis, Photoinhibition: Impairment of the primary charge separation between P-680 and pheophytin in photosystem II of chloroplasts (214) 370
Dempsey, C.E., G.D. Cryer and A. Watts, The interaction of amino-deuteromethylated melittin with phospholipid membranes studied by deuterium NMR (218) 173
Dencher, N.A. see S. Grzesiek (208) 337
Denis, H. see A. Viel (223) 232
DerVartanian, D.V. see E. Samain (216) 140
DerVartanian, D.V. see M. -C. Liu (218) 227
Deshpande, S.S. see Y. Aracava (222) 63
Deshusses, J. see J.-P. Patthey (210) 137
Desmond, H., S. Pauwels, A. Varro, H. Gregory, J. Young and G.J. Dockray, Isolation and characterization of the intact gastrin precursor from a gastrinoma (210) 185
Desmoulins, D. see C. Aussel (214) 327
Deutzmann, R. see K. Mann (218) 167
Devaux, F. see A. Zachowski (223) 315
Dewhurst, S., J. Bresser, M. Stevenson, K. Sakai, M.J. Evinger-Hodges and D.J. Volsky, Susceptibility of human glial cells to infection with human immunodeficiency virus (HIV) (213) 138
Dewhurst, S., M. Stevenson and D.J. Volsky, Expression of the T4 molecule (AIDS virus receptor) by human brain-derived cells (213) 133
Dexpert, J., E. Delain, B. Piriou and F. Pochon, The covalent and non-covalent binding modes of elastase with α_2-macroglobulin influence the conformation of the protease (225) 223
Di Cola, D. and P. Sacchetta, Irreversible inactivation of calcium-dependent proteinases from rat liver by biological disulfides (210) 81
Di Rago, J.P., X. Perea and A.-M. Colson, DNA sequence analysis of diuron-resistant mutations in the mitochondrial cytochrome *b* gene of *Saccharomyces cerevisiae* (208) 208
Diamant, S., B. Avraham and D. Atlas, Neomycin inhibits K^+- and veratridine-stimulated noradrenaline release in rat brain slices and rat brain synaptosomes (219) 445
Dickens, B.F. see C.M. Arroyo (221) 101
Dickey, B.F., H.Y. Pyun, K.C. Williamson and J. Navarro, Identification and purification of a novel G protein from neutrophils (219) 289
Didierjean, L. see C.P. Schelling (214) 21
Didsbury, J.R. and R. Snyderman, Molecular cloning of a new human G protein: Evidence for two $G_{i\alpha}$-like protein families (219) 259
Didsbury, J.R., Y.-S. Ho and R. Snyderman, Human G_i protein α-subunit: deduction of amino acid structure from a cloned cDNA (211) 160
Dieter, P., J.G. Altin and F.L. Bygrave, Possible involvement of prostaglandins in vasoconstriction induced by zymosan and arachidonic acid in the perfused rat liver (213) 174
Dietrich, W. see G.W. Mayr (213) 278

Dietzchold, B. see D. Carper (220) 209
Dikanov, S.A., A.V. Astashkin, Yu.D. Tsvetkov, M.G. Goldfeld and A.G. Chetverikov, Detection of deuterium nuclei in the immediate surroundings of P700 centers of plant photosynthesis by electron spin echo modulation (224) 75
Dillenschneider, M., A. Hetherington, A. Graziana, G. Alibert, P. Berta, J. Haiech and R. Ranjeva, The formation of inositol phosphate derivatives by isolated membranes from *Acer pseudoplatanus* is stimulated by guanine nucleotides (208) 413
Diller, R., M. Stockburger, D. Oesterhelt and J. Tittor, Resonance Raman study of intermediates of the halorhodopsin photocycle (217) 297
Dimitriadis, E. see M. Dunlop (220) 84
Dimitrov, M.I., C.A. Egorov, A.A. Donchev and B.P. Atanasov, Complete amino acid sequence of poplar plastocyanin *b* (226) 17
Dimitrov, S.I. see V.L. Makarov (212) 263
Dimroth, P. see A. Hoffmann (220) 121
Djé, M.K. see A. Viel (223) 232
Dobson, C.M. and L.-Y. Lian, A ^{31}P MAS NMR study of cytidine 2′-phosphate bound to ribonuclease A in the crystalline state (225) 183
Dockray, G.J. see H. Desmond (210) 185
Dodd, I., S. Jalalpour, W. Southwick, P. Newsome, M.J. Browne and J.H. Robinson, Large scale, rapid purification of recombinant tissue-type plasminogen activator (209) 13
Dogbo, O., F. Bardat, J. Quennemet and B. Camara, Metabolism of plastid terpenoids: in vitro inhibition of phytoene synthesis by phenethyl pyrophosphate derivatives (210) 211
Doi, M., M. Tanaka, T. Ishida and M. Inoue, The three-dimensional similarity between a dimeric antiparallel extended structure and a β-turn folded form of enkephalin (213) 265
Doi, M. see M.D. Song (221) 167
Dolja, V.V. and J.G. Atabekov, The structure of barley stripe mosaic virus double-stranded RNAs (214) 313
Dolja, V.V., D.P. Grama, S.Yu. Morozov and J.G. Atabekov, Potato virus X-related single- and double-stranded RNAs: Characterization and identification of terminal structures (214) 308
Dollet, M. see J.-F. Riou (213) 304
Donchev, A.A. see M.I. Dimitrov (226) 17
Donnér, J., H. Eriksson and P. Belfrage, The acute GH action in rat adipocytes is associated with enhanced phosphorylation of a 46 kDa plasma membrane protein enriched by GH-Sepharose (208) 269
Donowitz, M. see G.W.G. Sharp (221) 309
Dooley, D.M., M.A. McGuirl, J. Peisach and J. McCracken, The generation of an organic free radical in substrate-reduced pig kidney diamine oxidase-cyanide (214) 274
Dorée, M. see A. Laurent (226) 324
Dorman, C.J. see J.S. Evans (208) 211
Doronin, S.V., O.I. Lavrik, G.A. Nevinsky and V.N. Podust, The efficiency of dNTP complex formation with human placenta DNA polymerase α as demonstrated by affinity modification (216) 221
Douce, R. see J. Covès (208) 401
Douglas, K. see J.-P. Tenu (220) 93
Douglas, R. see P.A. Andreasen (209) 213
Douste-Blazy, L. see J. Fauvel (216) 45
Douste-Blazy, L. see J. Fauvel (221) 397
Downer, R.G.H. see A.P. Jahagirdar (219) 83
Doyen, N. see M. Ekker (222) 337
Drachev, L.A., A.D. Kaulen, V.P. Skulachev and V.V. Zorina, Protonation of a novel intermediate P is involved in the MbR step of the bacteriorhodopsin photocycle (209) 316
Drachev, L.A., A.D. Kaulen, V.P. Skulachev and V.V. Zorina, The mechanism of H^+ transfer by bacteriorhodopsin: The properties and the function of intermediate P (226) 139
Drachev, L.A., M.D. Mamedov and A.Yu. Semenov, The antimycin-sensitive electrogenesis in *Rhodopseudomonas sphaeroides* chromatophores (213) 128
Dratz, E.A. see K.R. Parker (211) 35
Dreher, M.L. see R.R. Mattingly (223) 11
Dreusicke, D. and G.E. Schulz, The glycine-rich loop of adenylate kinase forms a giant anion hole (208) 301
Drew, R.E. see W.J. Brammar (215) 291
Dreyer, D. see Y. Le Bouc (222) 181
Driessen, A.J.M. see M. Opekarová (213) 45
Drygin, Yu.F., M.V. Sapotsky and A.A. Bogdanov, Radish mosaic virus VPg: Characteristics and linkage with virion RNAs (215) 247
DuBow, M.S. see D.B. Levin (222) 199
Dubacq, J.P. see A. Oursel (219) 393
Dubin, M., L. Grinblat, S.H. Fernandez Villamil and A.O.M. Stoppani, Nitrofuran inhibition of microsomal lipid peroxidation (220) 197
Dubyak, G.R. see R.G. Appel (224) 396
Ducastaing, A. see M. Savart (216) 22
Duchatelle, P. and M. Joffre, Ca^{2+}-dependent-chloride and potassium currents in rat Leydig cells (217) 11
Duclohier, H. see G. Molle (224) 208
Duine, J.A. see R.A. van der Meer (221) 299
Dukhovich, V.F. see B.V. Chernyak (215) 300
Dulobova, I.E. see E.D. Sverdlov (217) 275
Dulubova, I.E. see Yu.A. Ovchinnikov (213) 73
Dumont, J.E. see I. Graff (210) 204
Dunahay, T.G., G. Schuster and L.A. Staehelin, Phosphorylation of spinach chlorophyll-protein complexes: CPII*, but not CP29, CP27, or CP24, is phosphorylated in vitro (215) 25
Duncan, T.M., D. Parsonage and A.E. Senior, Structure of the nucleotide-binding domain in the β-subunit of *Escherichia coli* F_1-ATPase (208) 1
Dunlop, M., E. Dimitriadis and R.G. Larkins, Acute changes in *myo*-inositol uptake and $^{22}Na^+$ flux in murine neuroblastoma cells (N1E-115) following insulin (220) 84
Dunn, M.J. see R.G. Appel (224) 396
Dunne, M.J. and O.H. Petersen, Intracellular ADP activates K^+ channels that are inhibited by ATP in an insulin-secreting cell line (208) 59
Duque-Magalhães, M.C. and J.M. Gualberto, Regulation of mitochondrial proteolysis: Selective degradation of inner membrane polypeptides (210) 142
Duriez, P. see J. Benveniste (226) 371
Dy, M. see P.P. Kamoun (226) 285
Dyatkina, N., S. Minassian, M. Kukhanova, A. Krayevsky, M. von Janta-Lipinsky, Z. Chidgeavadze and R. Beabealashvilli, Properties of 2′,3′-dideoxy-2′,3′-dehydrothymidine 5′-triphosphate in terminating DNA synthesis catalyzed by several different DNA polymerases (219) 151
Dyer, T.A. see P.J. Nixon (209) 83
Dyer, T.A. see S.M. Hird (209) 181
Dzhandzhugazyan, K.N. see Yu.A. Ovchinnikov (217) 111
Dziegielewska, K.M. see D.L. Christie (214) 45
Dzoljic, M.R. see G.R. Elliott (217) 6

E

Earley, F.G.P., S.D. Patel, C.I. Ragan and G. Attardi, Photolabelling of a mitochondrially encoded subunit of NADH dehydrogenase with [^{3}H]dihydrorotenone (219) 108
Ebels, I. see H.P.J.M. Noteborn (216) 200
Eberhardt, I. see L. Kiesel (217) 85
Eberl, G. and K. Schnell, Ca^{2+} uptake and IP_3-induced Ca^{2+} release in permeabilized humanlymphocytes (222) 349
Eberle, A.N. see T. Scimonelli (226) 134
Ebert, B. see G.V. Semisotnov (224) 9
Ebert, W. see B. Appelhans (224) 14
Ebinuma, H. see F. Takaiwa (221) 43
Ebrey, T.G. see Z. Dancshazy (209) 44
Eckerskorn, C. and M. Klingenberg, In the uncoupling protein from brown adipose tissue the C-terminus protrudes to the c-side of the membrane as shown by tryptic cleavage (226) 166
Eckhardt-Wallasch, H. see H.G. Kress (221) 28
Eckstein, F. see J.M. Fernandez (216) 89
Edge, M.T. see R.J. Bradley (224) 277
Edvardsen, K. see E. Bock (225) 33
Edwards, C.R.W. see A.J. Harmar (208) 67
Edwards, C. see K. Enomoto (223) 82
Edzes, H.T. see D.H. Joziasse (221) 139
Efimov, A.V., Pseudo-homology of protein standard structures formed by two consecutive β-strands (224) 372
Efremov, R.G. see N.G. Abdulaev (213) 113
Egea, G. see X. Guitart (219) 219
Egea, G. see X. Rabasseda (213) 337
Egestad, B. see H.-U. Marschall (213) 411
Egorov, C.A. see M.I. Dimitrov (226) 17
Ehlers, B., J. Czichos and P. Overath, Repression and reactivation of the variant surface glycoprotein gene in *Trypanosoma brucei* (225) 53
Ehnholm, C. see L. Keso (215) 105
Eichner, W. see J. Hoppe (223) 243
Eide, B. see P.M. Murphy (221) 81
Ekindjian, O.G. see C. Aussel (214) 327
Ekker, M., N. Doyen, M. Leblond-Francillard and F. Rougeon, A mouse renin promoter containing the conserved decanucleotide element binds the same B-cell factors as an authentic immunoglobulin heavy chain promoter (222) 337
El Alaoui, S. see J.G. Delcros (220) 236
El-Sabban, M. see G.W.G. Sharp (221) 309
Elder, D.E. see J. Thurin (208) 17
Elferink, M.G.L. see W. Crielaard (225) 6
Eliopoulos, E. see D.K. Stammers (218) 178
Ellermann, J., A. Kobelt, A. Pfaltz and R.K. Thauer, On the role of *N*-7-mercaptoheptanoyl-*O*-phospho-L-threonine (component B) in the enzymatic reduction of methyl-coenzyme M to methane (220) 358
Elliott, G.R., M.J. van Batenburg and M.R. Dzoljic, Enhanced prostaglandin E_2 and thromboxane B_2 release from resident peritoneal macrophages isolated from morphine-dependent rats (217) 6
Ellis, C.A., A. Aitken, K. Takayama and N. Qureshi, Substitution of phosphatidylserine by lipid A in the activation of purified rabbit brain protein kinase C (218) 238
Ellis, R.J. see R.S. Williams (225) 59
Elola, M.T. see N.E. Fink de Cabutti (223) 330
Emori, Y. see K. Ishidoh (226) 33
Emori, Y. see K. Suzuki (220) 271
Emori, Y. see S. Imajoh (215) 274
Enami, I., K. Satoh and S. Katoh, Crosslinking between the 33 kDa extrinsic protein and the 47 kDa chlorophyll-carrying protein of the PS II reaction center core complex (226) 161
Ender, B. see B. Appelhans (224) 14
Engelbrecht, S. and W. Junge, Fragmentation of chloroplast coupling factor in dependence of bound nucleotides: Preparation of a reconstitutionally active form of subunit δ (219) 321
Engelhard, M., B. Hess, M. Chance and B. Chance, X-ray absorption studies on bacteriorhodopsin (222) 275
Engelhardt, H. see J.P. Chalcroft (211) 53
Engels, F., H. Willems and F.P. Nijkamp, Cyclooxygenase-catalyzed formation of 9-hydroxylinoleic acid by guinea pig alveolar macrophages under non-stimulated conditions (209) 249
England, P.J. see J.P. Huggins (217) 32
Engström, L. see F.A. Nettelblad (214) 249
Enomoto, K., K. Furuya, T. Maeno, C. Edwards and T. Oka, Mechanically induced electrical responses in murine mammary epithelial cells in primary culture (223) 82
Enrich, C. see M.J. Coll (208) 418
Enyedi, P. see E. Brown (218) 113
Epand, R.M. and S.-W. Hui, Effect of electrostatic repulsion on the morphology and thermotropic transitions of anionic phospholipids (209) 257
Epe, B., P. Woolley and H. Hornig, Competition between tetracycline and tRNA at both P and A sites of the ribosome of *Escherichia coli* (213) 443
Eraso, P., A. Cid and R. Serrano, Tight control of the amount of yeast plasma membrane ATPase during changes in growth conditions and gene dosage (224) 193
Eraso, P. and C. Gancedo, Activation of yeast plasma membrane ATPase by acid pH during growth (224) 187
Erdmann, V.A. see R.K. Hartmann (218) 215
Erecińska, M. and D. Nelson, Amino acid neurotransmitters in the CNS: Relationships between net uptake and exchange in rat brain synaptosomes (213) 61
Eremenko, T. see C. Delfini (210) 17
Ericsson, L.H., N. Eriksen, K.A. Walsh and E.P. Benditt, Primary structure of duck amyloid protein A: The form deposited in tissues may be identical to its serum precursor (218) 11
Ericsson, L.H. see D.R. Eyre (220) 337
Eriksen, N. see L.H. Ericsson (218) 11
Eriksson, H. see J. Donnér (208) 269
Erneux, C. see I. Graff (210) 204
Ernst, M. and E.R. Froesch, Triiodothyronine stimulates proliferation of osteoblast-like cells in serum-free culture (220) 163
Ernst-Fonberg, M.L., M. McGee Tucker and I.B. Fonberg, The amphiphilicity of ACP helices: a means of macromolecular interaction? (215) 261
Erny, P. see P. Dabadie (226) 77
Errasfa, M. see B. Rothhut (219) 169
Errington, J., Two separable functional domains in the σ-subunit of RNA polymerase in *Bacillus subtilis*? (224) 257
Esch, F. see P. Whiting (219) 459
Escher, C. see J.-C. Wagner (218) 31
Escoffier, A. see A. Oursel (219) 393
Escudero, F. see M.J. Carmena (218) 73
Escurat, M. see A. Lomri (222) 311
Esnouf, M.P. see K. Harlos (224) 97
Esnouf, P. see R.J.P. Williams (209) 111
Espinal, J. see M. Beggs (215) 13
Espinet, C., R. Bartrons and J. Carreras, Fructose 2,6-bisphosphate and glucose 1,6-bisphosphate in erythrocytes during chicken development (209) 254
Estes, J.E. see L.A. Selden (217) 89
Estrela, J.M. see C. Roncero (208) 105

Estruch, F. see J.E. Pérez-Ortín (208) 31
Etchebest, C. see Y. Trudelle (216) 11
Etheredge, J.L. see D.P. Ringer (224) 59
Etienne, J. see J. Masliah (222) 11
Etzold, T., C.C. Fritz, J. Schell and P.H. Schreier, A point mutation in the chloroplast 16 S rRNA gene of a streptomycin resistant *Nicotiana tabacum* (219) 343
Evans, A.T., E. Formukong and F.J. Evans, Activation of phospholipase A_2 by cannabinoids: Lack of correlation with CNS effects (211) 119
Evans, F.J. see A.T. Evans (211) 119
Evans, F.J. see S.F. Brooks (224) 109
Evans, J.S., B.A. Levine, I.P. Trayer, C.J. Dorman and C.F. Higgins, Sequence imposed structural constraints in the TonB protein of *E. coli* (208) 211
Evans, M.C.W. see R.W. Mansfield (220) 74
Evenden, B.J. see K.S. Authi (213) 95
Evinger-Hodges, M.J. see S. Dewhurst (213) 138
Exton, J.H. see S.B. Bocckino (225) 201
Eyre, D.R., S. Apone, J.-J. Wu, L.H. Ericsson and K.A. Walsh, Collagen type IX: evidence for covalent linkages to type II collagen in cartilage (220) 337

F

Fabian, M., P.-E. Thörnström, P. Brzezinski and B.G. Malmström, Two-electron reduction is required for rapid internal electron transfer in resting, pulsed and oxygenated cytochrome *c* oxidase (213) 396
Fabri, L.J. see R.J. Simpson (224) 128
Faerman, A.I. see M.A. Chernousov (217) 124
Fairwell, T., P. Julià, R. Kaiser, B. Holmquist, X. Parés, B.L. Vallee and H. Jörnvall, Acetylated N-terminal structures of class III alcohol dehydrogenases: Differences among the three enzyme classes (222) 99
Falasca, A. see E.J. Wawrzynczak (219) 51
Falcioni, G., G. Cincolà and M. Brunori, Glutathione peroxidase and oxidative hemolysis in trout red blood cells (221) 355
Falgueyret, J.P. see S. Benjannet (224) 142
Falk, P. see G. Larson (214) 41
Falkmer, S. see J.M. Conlon (208) 445
Falloon, J., H. Malech, G. Milligan, C. Unson, R. Kahn, P. Goldsmith and A. Spiegel, Detection of the major pertussis toxin substrate of human leukocytes with antisera raised against synthetic peptides (209) 352
Falus, A. see L. Fesus (224) 104
Fang, P.F. see Y.X. Zhu (208) 253
Fang, Y.-I. see T. Isobe (223) 92
Fanjul, L.F. see D.R. Cooper (214) 122
Faragó, A. see L. Buday (223) 15
Farese, R.V. see D.R. Cooper (214) 122
Farese, R.V. see J.S. Davis (208) 287
Farkas, G. see L. Buday (223) 15
Farkas, T. see G. Premecz (226) 13
Farnararo, M. see P. Bruni (222) 27
Farnsworth, D.E. see R. Feyereisen (222) 345
Farr, G.W. see H. Sternlicht (214) 226
Farrar, W.L. see M.R. Ruff (211) 17
Faucher, N. see L. Beaumier (221) 289
Faulhammer, H.G. and R.L. Joshi, Structural features in aminoacyl-tRNAs required for recognition by elongation factor Tu (217) 203
Faulhammer, H.G. see R.L. Joshi (208) 189
Fauque, G.D., I. Moura, J.J.G. Moura, A.V. Xavier, N. Galliano and J. LeGall, Isolation and characterization of a rubredoxin and a flavodoxin from *Desulfovibrio desulfuricans* Berre-Eau (215) 63
Fauque, G.D. see Y.M. Berlier (221) 241
Fauvel, J., J.-P. Salles, V. Roques, H. Chap, H. Rochat and L. Douste-Blazy, Lipocortin-like anti-phospholipase A_2 activity of endonexin (216) 45
Fauvel, J., P. Vicendo, V. Roques, J. Ragab-Thomas, C. Granier, I. Vilgrain, E. Chambaz, H. Rochat, H. Chap and L. Douste-Blazy, Isolation of two 67 kDa calcium-binding proteins from pig lung differing in affinity for phospholipids and in anti-phospholipase A_2 activity (221) 397
Favaudon, V. see F. Pochon (217) 101
Fedi, S. see G. Laffi (220) 217
Feduchi, E. and L. Carrasco, Adenovirus infection reverses the antiviral state induced by human interferon (214) 153
Feeney, J. see B. Birdsall (217) 106
Fehr, S., R. Ivell, R. Koll, D. Schams, M. Fields and D. Richter, Expression of the oxytocin gene in the large cells of the bovine corpus luteum (210) 45
Feiler, U. see W. Nitschke (218) 283
Feingold, D.S. and R. Bentley, Conformational aspects of the reaction mechanisms of polysaccharide lyases and epimerases (223) 207
Feinstein, M.B. see F. O'Rourke (214) 176
Feizi, T. see C. Southan (214) 301
Feldman, D.H., B.M. Olivera and D. Yoshikami, Omega *Conus geographus* toxin: a peptide that blocks calcium channels (214) 295
Felipo, V. and S. Grisolía, Precursors of mitochondrial proteins are degraded in the cytosol at different rates (209) 227
Felipo, V. and S. Grisolía, Retention of actin synthesis in liver under conditions that inhibit synthesis of almost all other proteins (210) 173
Felix, A.M. see A.A. Haritos (218) 107
Fellmann, P. see A. Zachowski (223) 315
Fendler, K., E. Grell and E. Bamberg, Kinetics of pump currents generated by the Na^+,K^+-ATPase (224) 83
Fernandez, J.M., M. Lindau and F. Eckstein, Intracellular stimulation of mast cells with guanine nucleotides mimic antigenic stimulation (216) 89
Fernández-Gonzalez, M.A. see M.J. Carmena (218) 73
Fernandez Villamil, S.H. see M. Dubin (220) 197
Fernig, D.G. and R.J. Mayer, Degradation of nuclear proteins: studies on transplanted B82 cell karyoplast proteins (210) 165
Fernlund, P. see S. Gershagen (220) 129
Ferrara, P., F. Pecceu, E. Marchese, N. Vita, W. Roskam and J. Lupker, Characterization of recombinant glycosylated human interleukin 2 produced by a recombinant plasmid transformed CHO cell line (226) 47
Ferrari, N. see A. De Ambrosis (225) 120
Ferri, G. see M. Galliano (208) 364
Ferro, M. see D. Fürst (224) 49
Fesenko, E.E. see S.S. Kolesnikov (222) 37
Fesenko, E.E., V.I. Novoselov and M.F. Bystrova, The subunits of specific odor-binding glycoproteins from rat olfactory epithelium (219) 224
Festa, A. see M. Clementi (221) 11
Fesus, L., V. Thomazy and A. Falus, Induction and activation of tissue transglutaminase during programmed cell death (224) 104

Fetisova, Z.G., A.M. Freiberg and K.E. Timpmann, Investigations by picosecond polarized fluorescence spectrochronography of structural aspects of energy transfer in living cells of the green bacterium *Chlorobium limicola* (223) 161
Feve, B. and J. Pairault, Dexamethasone-dependent expression of β^{1-24} corticotropin stimulated adenylate cyclase during adipose conversion of 3T3-F442A cells (219) 56
Fewson, C.A. see G.A. Nimmo (213) 18
Feyereisen, R. and D.E. Farnsworth, Inhibition of insect juvenile hormone synthesis by phorbol 12-myristate 13-acetate (222) 345
Field, A.C. see T. Capiod (217) 247
Field, R.A. see N. Foote (214) 347
Fields, M. see S. Fehr (210) 45
Filetti, S. see G. Damante (225) 264
Filipski, J., Correlation between molecular clock ticking, codon usage, fidelity of DNA repair, chromosome banding and chromatin compactness in germline cells (217) 184
Findlay, J.B.C. see A. Cavaggioni (212) 225
Fink, G. see A.J. Harmar (208) 67
Fink, G. see J. Simpson (217) 62
Fink, P.S., T. Whitford, M. Leffak and L.J. Prochaska, Answer (220) 263
Fink, P.S., T. Whitford, M. Leffak and L.J. Prochaska, Homology between bacterial DNA and bovine mitochondrial DNA encoding cytochrome *c* oxidase subunit III (214) 75
Fink de Cabutti, N.E., M. Caron, R. Joubert, M.T. Elola, D. Bladier and J. Herkovitz, Purification and some characteristics of a β-galactoside binding soluble lectin from amphibian ovary (223) 330
Finkelstein, A. see R.O. Blaustein (226) 115
Finn, P.W. see R.V. Fishleigh (214) 219
Fisher, S.C. see A.M. Salter (220) 159
Fishleigh, R.V., B. Robson, J. Garnier and P.W. Finn, Studies on rationales for an expert system approach to the interpretation of protein sequence data: Preliminary analysis of the human epidermal growth factor receptor (214) 219
Fishleigh, R.V. see C.A. Morrison (214) 65
Fittkau, S. see D. Brömme (219) 441
Fitzgerald, M. see F.-Z. Chung (211) 200
Flagella, Z. see P. Cantatore (213) 144
Flentje, U. see W. Hinderer (214) 101
Fletcher, A.E., E.H. Allan, D.J. Casley and T.J. Martin, Atrial natriuretic factor receptors and stimulation of cyclic GMP formation in normal and malignant osteoblasts (208) 263
Flippen-Anderson, J.L. see Q.S. Yu (221) 325
Flitter, W.D. see K.M. Morehouse (222) 246
Floc'h, F. see J. Berthou (218) 55
Florencio, F.J., S. Marqués and P. Candau, Identification and characterization of a glutamate dehydrogenase in the unicellular cyanobacterium *Synechocystis* PCC 6803 (223) 37
Flügel, R.M. see B. Maurer (222) 286
Flynn, T.G. see M.M. Bhadbhade (211) 243
Fojo, S.S., S.W. Law and H.B. Brewer, jr, The human preproapolipoprotein C-II gene: Complete nucleic acid sequence and genomic organization (213) 221
Földes, I. see G. Premecz (226) 13
Foley, M. see I.V. Zlatanov (222) 47
Follmann, H. see H.P.C. Hogenkamp (219) 197
Follmann, H. see J. Harder (222) 171
Fonberg, I.B. see M.L. Ernst-Fonberg (215) 261
Foote, N., P.M.A. Gadsby, R.A. Field, C. Greenwood and A.J. Thomson, A comparison by magnetic circular dichroism of compound X and compound II of horseradish peroxidase (214) 347
Forbes, I.J. see J.K. French (212) 242
Foreman, R.C. and J.D. Judah, The processing and secretion of rat serum albumin by oocytes from *Xenopus laevis* (219) 75
Foreman, R.C., Disruption of the Lys-290–Glu-342 salt bridge in human α_1-antitrypsin does not prevent its synthesis and secretion (216) 79
Formukong, E. see A.T. Evans (211) 119
Forrest, M.E. and J.T. Beatty, Purification of *Rhodobacter capsulatus* RNA polymerase and its use for in vitro transcription (212) 28
Forsgren, M., B. Råden, M. Israelsson, K. Larsson and L.-O. Hedén, Molecular cloning and characterization of a full-length cDNA clone for human plasminogen (213) 254
Forssmann, W.G. see M. Gagelmann (225) 251
Fortina, M.G. see P.L. Manachini (214) 305
Fouladgar, H. see D. Rogez (219) 22
Fracasso, F. see P. Cantatore (213) 144
Francina, A. see F. Baklouti (223) 59
Franco, L. see J.E. Pérez-Ortín (208) 31
Frank, G. see T.D. Wechsler (210) 189
Frank, H.A. see D.F. Bocian (214) 92
Franke, J.E. see G.P. Palace (215) 58
Fraser, C.M. see F.-Z. Chung (211) 200
Fratelli, M. and A. De Blasi, Agonist-induced α_1-adrenergic receptor changes: Evidence for receptor sequestration (212) 149
Fredholm, B.B. see C. Nordstedt (220) 57
Freiberg, A.M. see Z.G. Fetisova (223) 161
French, J.K., N.P. Hurst, P.D. Zalewski, L. Valente and I.J. Forbes, Calcium ionophore A23187 enhances human neutrophil superoxide release, stimulated by phorbol dibutyrate, by converting phorbol ester receptors from a low- to high-affinity state (212) 242
Frère, J.-M. see L. Varetto (225) 218
Frerman, F.E. see M.K. Johnson (226) 129
Frid, M.G. see M.A. Chernousov (217) 124
Fridkin, M. see R. Buchta (211) 165
Friedman, J.E. see P.I. Lelkes (208) 357
Fritz, C.C. see T. Etzold (219) 343
Fritzsche, T.M. see A. Wolf (212) 203
Froesch, E.R. see M. Ernst (220) 163
Fromageot, P. see O. Trémeau (208) 236
Fromme, P., P. Gräber and J. Salnikow, Isolation and identification of a fourth subunit in the membrane part of the chloroplast ATP-synthase (218) 27
Fruchart, J.-C. see J. Benveniste (226) 371
Fruth, U. see H.-G. Simon (223) 352
Frydman, R.B., O. Ruiz, M. Kreisel and U. Bachrach, Oxidation of *N*-alkyl and *C*-alkylputrescines by diamine oxidases (219) 380
Füglistaller, P. see F. Suter (217) 279
Fujii, M., M. Nakamura, K. Ohtani, K. Sugamura and Y. Hinuma, 12-*O*-Tetradecanoylphorbol-13-acetate induces the enhancer function of human T-cell leukemia virus type I (223) 299
Fujii, T. see U. Kikkawa (217) 227
Fujii, T. see U. Kikkawa (223) 212
Fujii, T. see Y. Ono (226) 125
Fujii, Y. see K. Satoh (216) 7
Fujisawa, H. see T. Ichimura (219) 79
Fujita, J. see Y. Seino (223) 74
Fujita, Y. see T. Tobimatsu (222) 56
Fukazawa, C., K. Udaka, A. Murayama, W. Higuchi and A. Totsuka, Expression of soybean glycinin subunit precursor cDNAs in *Escherichia coli* (224) 125
Fukuda, K. see T. Kurosaki (214) 253
Fukuda, K. see T. Tobimatsu (222) 56
Fukui, S. see A. Kurosaka (215) 137
Fukui, S. see T. Itoh (219) 339
Fukumoto, H. see Y. Seino (223) 74
Fukumoto, Y. see K. Kariya (217) 69
Fukumoto, Y. see T. Tsuda (208) 39

Fukusen, N. see H. Kido (223) 223
Fukushima, M. see J. Abe (226) 58
Fukuzaki, H. see K. Kariya (217) 69
Fukuzawa, H., T. Yoshida, T. Kohchi, T. Okumura, Y. Sawano and K. Ohyama, Splicing of group II introns in mRNAs coding for cytochrome b_6 and subunit IV in the liverwort *Marchantia polymorpha* chloroplast genome: Exon specifying a region coding for two genes with the spacer region (220) 61
Fuller, C.M. see A.P. Morris (211) 195
Fumagalli, R. see J.L. Lorenzo (218) 77
Funakoshi, I. see A. Kurosaka (215) 137
Funatsu, G. see T. Utsumi (216) 99
Furmaniak, J., F.A. Hashim, P.R. Buckland, V.B. Petersen, K. Beever, R.D. Howells and B. Rees Smith, Photoaffinity labelling of the TSH receptor on $FRTL_5$ cells (215) 316
Fürst, D., R. Nave, M. Osborn, K. Weber, A. Bardosi, N. Archidiacono, M. Ferro, V. Romano and G. Romeo, Nebulin and titin expression in Duchenne muscular dystrophy appears normal (224) 49
Furukawa, S. see Y. Furukawa (208) 258
Furukawa, Y., S. Furukawa, F. Ikeda, E. Satoyoshi and K. Hayashi, Aliphatic side chain of catecholamine potentiates the stimulatory effect of the catechol part on the synthesis of nerve growth factor (208) 258
Furuki, Y. see T. Yamamoto (219) 326
Furuya, K. see K. Enomoto (223) 82
Futai, M. see S.-Y. Hsu (218) 222
Futai, M. see T. Noumi (213) 381
Fyles, J.M. see P.M. Jones (219) 139

G

Gaboriaud, C., V. Bissery, T. Benchetrit and J.P. Mornon, Hydrophobic cluster analysis: an efficient new way to compare and analyse amino acid sequences (224) 149
Gacesa, P., Alginate-modifying enzymes: A proposed unified mechanism of action for the lyases and epimerases (212) 199
Gadaleta, M.N. see P. Cantatore (213) 144
Gadsby, P.M.A. see N. Foote (214) 347
Gadsby, P.M.A. see R.S. Blackmore (219) 244
Gaffner, R. see P. Moonen (226) 314
Gagelmann, M., D. Hock and W.G. Forssmann, Relaxation of smooth muscle by cardiodilatin/atrial natriuretic peptide is inhibited by cAMP-dependent phosphorylation (225) 251
Gagnon, J. see G.J. Arlaud (222) 129
Gahmberg, C.G. see M.J. Coll (208) 418
Gahmberg, C.G. see M. Patarroyo (210) 127
Gal, A., Y. Shahak, G. Schuster and I. Ohad, Specific loss of LHCII phosphorylation in the *Lemna* mutant 1073 lacking the cytochrome b_6/f complex (221) 205
Galaktionov, S.G. see M.A. Rodionov (223) 402
Galassi, C. see S. Lamponi (216) 265
Galeotti, T. see S. Borrello (209) 305
Gallacher, D.V. see A.P. Morris (211) 195
Gallay, J. see B. de Foresta (216) 107
Gallego, C., R. Bartrons and J. Carreras, Fructose 2,6-bisphosphate and glucose 1,6-bisphosphate in rabbit erythroid cells during differentiation (222) 167
Galliano, M., L. Minchiotti, P. Iadarola, M. Stoppini, G. Ferri and A.A. Castellani, The molecular defect of albumin Tagliacozzo: 313 Lys→Asn (208) 364
Galliano, N. see G.D. Fauque (215) 63
Gallop, P. see H.M. Hanauske-Abel (214) 236
Gallozzi, E. see A.M. Vaccaro (216) 190
Galván, A. see C. Osuna (211) 41
Gamblin, S.J. see J.A. Littlechild (225) 123
Gancedo, C. see J.J. Aragón (226) 121
Gancedo, C. see P. Eraso (224) 187
Ganguly, S., Iodination-induced alterations in biochemical properties of human placental insulin receptor (224) 198
Ganguly, U. see R. Ray (213) 81
Ganter, U.M. see K. Gerwert (213) 39
Ganter, U. see T. Andus (221) 18
Gao, X. see A. Barlas (218) 266
Gao, Z. and E. Bäuerlein, Identifying subunits of ATP synthase $TF_0 \cdot F_1$ in contact with phospholipid head groups: α-Subunits are labelled selectively by a new photoreactive phospholipid designed for hydrophilic photolabelling (223) 366
Garber, M.B. see S.D. Trakhanov (220) 319
Garber, M.B. see S.E. Sedelnikova (220) 227
Gardeström, P., Adenylate ratios in the cytosol, chloroplasts and mitochondria of barley leaf protoplasts during photosynthesis at different carbon dioxide concentrations (212) 114
Gardette, J., D. Margelin, J.C. Maziere, J. Bertrand and J. Picard, Effect of dibutyryl cyclic AMP and theophylline on lipoprotein lipase secretion by human monocyte-derived macrophages (225) 178
Gardinier, M.V. see W.B. Macklin (223) 417
Gardner, J.O. see J. Simpson (217) 62
Gárdos, G. see L. Hunyady (225) 72
Gariglio, P. see J. Silva (214) 71
Garland, L.G. see J. Dawson (214) 171
Garland, L.G. see R.W. Bonser (209) 134
Garland, L.G. see R.W. Randall (214) 167
Garland, P.B. see I.V. Zlatanov (222) 47
Garland, W.G., K.A. Louie, A.T. Matheson and A. Liljas, The complete amino acid sequence of the ribosomal 'A' protein (L12) from *Bacillus stearothermophilus* (220) 43
Garman, A.J. and S. Barret Kalindjian, The preparation and properties of novel reversible polymer-protein conjugates: 2-ω-Methoxypolyethylene (5000) glycoxymethylene-3-methylmaleyl conjugates of plasminogen activators (223) 361
Garnier, J. see R.V. Fishleigh (214) 219
Garrigues, A.M., F. Gehelmann, J.M. Girault, M. Delaage and J. Labouesse, Haloperidol-succinylglycyl[^{125}I]iodotyrosine, a novel iodinated ligand for dopamine D_2 receptors (224) 267
Garty, N.B. and Y. Salomon, Stimulation of partially purified adenylate cyclase from bull sperm by bicarbonate (218) 148
Gasa, S. see Y. Uehara (208) 352
Gasnier, B., D. Scherman and J.-P. Henry, Inactivation of the catecholamine transporter during the preparation of chromaffin granule membrane 'ghosts' (222) 215
Gatehouse, J.A. see J.N. Yarwood (222) 175
Gatehouse, J.A. see R.N. Waterhouse (209) 223
Gathercole, L.J. see K. Barnard (212) 49
Gause, G.G. see L.J. Summers (208) 11
Gausing, K. see P.S. Nielsen (225) 159
Gavrilova, E.F. see A.M. Kuznetsov (215) 219
Gawler, D. and M.D. Houslay, Insulin stimulates a novel GTPase activity in human platelets (216) 94
Gawlikowaki, W. see J.A. Czyrski (219) 331
Gayda, J.P., T. Yagi, H. Benosman and P. Bertrand, EPR redox study of cytochrome c_3 from *Desulfovibrio vulgaris* Miyazaki (217) 57
Gearing, A.J.H. see T.A. Bird (225) 21
Geddes, A.J. see D.K. Stammers (218) 178
Gehelmann, F. see A.M. Garrigues (224) 267

Geiger, J.D. and J.I. Nagy, Lack of adenosine deaminase deficiency in the mutant mouse *wasted* (208) 431
Geiger, T. see T. Andus (221) 18
Geisler, N., J. Vandekerckhove and K. Weber, Location and sequence characterization of the major phosphorylation sites of the high molecular mass neurofilament proteins M and H (221) 403
Gelfand, V.I. see S.A. Kuznetsov (212) 145
Genest, J. see J. Gutkowska (214) 17
Gensburger, C., G. Labourdette and M. Sensenbrenner, Brain basic fibroblast growth factor stimulates the proliferation of rat neuronal precursor cells in vitro (217) 1
Gentilini, P. see G. Laffi (220) 217
George, C. see B. DeCosta (223) 335
George, P.M. see S.O. Brennan (219) 431
Gerisch, G. see M. Westphal (209) 92
Gerken, S., K. Brettel, E. Schlodder and H.T. Witt, Direct observation of the immediate electron donor to chlorophyll-a_{II}^{+} (P-680) in oxygen-evolving photosystem II complexes: Resolution of nanosecond kinetics in the UV (223) 376
Gerken, S. see I. Witt (221) 260
Germann, W.J. see R.O. Blaustein (226) 115
Gershagen, S., P. Fernlund and Å. Lundwall, A cDNA coding for human sex hormone binding globulin: Homology to vitamin K-dependent protein S (220) 129
Gershman, L.C. see L.A. Selden (217) 89
Gersonde, K. see R. Arkowitz (217) 21
Gertler, A. see A. Ashkenazi (210) 51
Gerwert, K., U.M. Ganter, F. Siebert and B. Hess, Only water-exposed carboxyl groups are protonated during the transition to the cation-free blue bacteriorhodopsin (213) 39
Geschwind, J.-F. see M. Prentki (220) 103
Gesser, B. see J.E. Celis (220) 1
Gessner, W. see A. Brossi (223) 77
Gestautas, J. see R.P. Magnusson (208) 391
Gevondyan, N.M. see Yu.A. Ovchinnikov (217) 269
Gewitz, H.S. see J. Piefke (209) 104
Geyssant, A. see F. Baklouti (223) 59
Ghanem, N. see S. Huck (208) 221
Ghazi, A., L. Delamourd and E. Shechter, Absence of a unique relationship between active transport of lactose and protonmotive force in *E. coli* (209) 325
Ghiggino, K.P. see J.R. Wardlaw (223) 20
Ghosh, R. and R. Aggeler, Effect of lipid fluidity upon the activity and structure of the 39 kDa porin from *Enterobacter cloacae* 908S (222) 154
Ghuysen, J.-M. see L. Varetto (225) 218
Giardina, B. see M. Brunori (221) 161
Gibson, A. see E. Bock (225) 33
Gibson, I.F. see M. Bushfield (222) 299
Gierschik, P. and K.H. Jakobs, Receptor-mediated ADP-ribosylation of a phospholipase C-stimulating G protein (224) 219
Gierschik, P. see P.M. Murphy (221) 81
Gierse, J.K. see G.M. Olins (224) 325
Gigova, L. see I. Ivanov (210) 56
Gilbert, D.L. see C.A. Colton (223) 284
Gilboa-Garber, N. see D. Avichezer (216) 62
Giles, I.G. see Y.C. Hsu (218) 1
Gilkes, N.R. see M.L. Langsford (225) 163
Gilmanshin, R.I. and O.B. Ptitsyn, An early intermediate of refolding α-lactalbumin forms within 20 ms (223) 327
Gilroy, S., W.A. Hughes and A.J. Trewavas, Calmodulin antagonists increase free cytosolic calcium levels in plant protoplasts in vivo (212) 133
Giménez, C. see M.C. Aragón (212) 87
Ginns, E.I. see M.R. Ruff (211) 17
Ginsberg, J., P.G. Murray, J.E. Parente and K. Wong, Translocation of protein kinase C in porcine thyroid cells following exposure to thyrotropin (226) 223
Girard, F. see Y. Le Bouc (222) 181
Giraud, Y. see F. Baklouti (223) 59
Girault, J.M. see A.M. Garrigues (224) 267
Gishizky, M.L. and G.M. Grodsky, Differential kinetics of rat insulin I and II processing in rat islets of Langerhans (223) 227
Giudicelli, Y. see F. Bauché (219) 296
Gladilin, K.L. see Y.A. Motorin (220) 363
Glas-Albrecht, R. see B. Stecher (223) 25
Glaser, E., B. Hamasur, B. Norling and B. Andersson, Activation of F_1-ATPase isolated from potato tuber mitochondria (223) 304
Glaser, E. see B. Norling (223) 309
Glazer, A.N. see F. Suter (217) 279
Glennon, M.C. see M. Prentki (220) 103
Gliemann, J. and L. Sottrup-Jensen, Rat plasma α_1-inhibitor$_3$ binds to receptors for α_2-macroglobulin (221) 55
Glossmann, H. see J. Striessnig (212) 247
Glukhova, M.A., A.E. Kabakov, O.I. Ornatsky, T.D. Vasilevskaya, V.E. Koteliansky and V.N. Smirnov, Immunoreactive forms of caldesmon in cultivated human vascular smooth muscle cells (218) 292
Gnuchev, N.V. see L.P. Sashchenko (226) 261
Gocayne, J. see F.-Z. Chung (211) 200
Godfroid, J.-J. see F. Heymans (218) 35
Goedde, H.W. see T. Braun (215) 233
Goetz, J. see B. Uring-Lambert (217) 65
Goffeau, A. see L. de Meis (212) 323
Gogol, E.P., U. Lücken and R.A. Capaldi, The stalk connecting the F_1 and F_0 domains of ATP synthase visualized by electron microscopy of unstained specimens (219) 274
Going, J. see A.J. Harmar (208) 67
Gojobori, T., S. Aota, T. Inoue and K. Shimotohno, A sequence homology between the pX genes of HTLV-I/II and the murine IL-3 gene (208) 231
Gold, M.H. see K. Miki (210) 199
Golderer, G., P. Loidl and P. Gröbner, Histone acetyltransferase activity during the cell cycle (222) 322
Goldfeld, M.G. see S.A. Dikanov (224) 75
Goldsmith, P. see J. Falloon (209) 352
Goldsmith, P. see P.M. Murphy (221) 81
Goldstein, B.N. and A.N. Ivanova, Hormonal regulation of 6-phosphofructo-2-kinase/fructose-2,6-bisphosphatase: kinetic models (217) 212
Goldstein, S. see C. Mazière (218) 243
Golovanova, N.K. see G.I. Muzya (220) 371
Gómez, M.-E. see J.J. Aragón (226) 121
Gómez-Márquez, J. and F. Segade, Prothymosin α is a nuclear protein (226) 217
Goncalves de Moraes, V.L. and L. de Meis, ATP synthesis by the $(Na^+ + K^+)$-ATPase in the absence of an ionic gradient: Effects of organic solvent (222) 163
Gonzalez, D.H. see C.S. Andreo (213) 1
Gonzalez-Carrero, M. see J.L. Ramos (226) 241
González-Martínez, M. and A. Darszon, A fast transient hyperpolarization occurs during the sea urchin sperm acrosome reaction induced by egg jelly (218) 247
Goodchild, D.J. see J.M. Anderson (213) 29
Gopalakrishna, K., A. Kalia, A.K. Royyuru, Mrigank, M. Atreyi, M.V.R. Rao and V. Kothekar, Theoretical study on binding of Hoechst 33258 with oligonucleotides (215) 95
Gopalakrishna, R. and W.B. Anderson, Susceptibility of protein kinase C to oxidative inactivation: loss of both phosphotransferase activity and phorbol diester binding (225) 233
Gopher, A. see B. Kalderon (213) 209

Gopher, A. see B. Kalderon (220) 259
Goping, I.S. see G.L. Hammond (215) 100
Gordon, R.K. see A. Ahmad (214) 285
Gorewit, R.C., H.Y. Chen and J.J. Kopchick, Growth hormone gene expression in myoepithelial cells directed by various eucaryotic transcriptional regulatory sequences (225) 238
Goris, J. see P. Agostinis (224) 385
Gorter, G. see W. Siffert (212) 123
Gosselin, G. see P.F. Torrence (212) 267
Goto, K. see S. Ohta (226) 171
Gotoh, K. see W. Sakamoto (219) 437
Gottschalk, G. see M. Blaut (215) 53
Gounaris, K., U. Pick and J. Barber, Stoichiometry and turnover of photosystem II polypeptides (211) 94
Govindjee, R. see Z. Dancshazy (209) 44
Gowda, S. see R.V. Guntaka (221) 332
Gräber, P. see P. Fromme (218) 27
Graber, P. see P. Wingfield (215) 160
Grabner, M. see J. Striessnig (212) 247
Graceffa, P., Evidence for interaction between smooth muscle tropomyosin and caldesmon (218) 139
Graff, I., J. Mockel, E. Laurent, C. Erneux and J.E. Dumont, Carbachol and sodium fluoride, but not TSH, stimulate the generation of inositol phosphates in the dog thyroid (210) 204
Grama, D.P. see V.V. Dolja (214) 308
Grange, R.W. see M.E. Houston (219) 469
Granier, C. see J. Fauvel (221) 397
Grant, M.E. see S. Ayad (220) 181
Grant, N.J. see B. Demeneix (226) 337
Gras, U. see B. Stecher (223) 25
Gratzl, M. see O.K. Langley (220) 108
Gravel, D. see R. Cedergren (226) 63
Gray, D.W.R. see P. Hammonds (223) 131
Gray, J.C. see S.M. Hird (209) 181
Gray, K.A. see M. Hirasawa (221) 343
Grazi, E. see V. Lanzara (221) 387
Graziana, A. see M. Dillenschneider (208) 413
Green, M.J., H.A.O. Hill and D.G. Tew, The rate of oxygen consumption and superoxide anion formation by stimulated human neutrophils: The effect of particle concentration and size (216) 31
Greenbaum, L.M. see A. Barlas (218) 266
Greenberg, J.R. and L.I. Slobin, Eukaryotic elongation factor Tu is present in mRNA-protein complexes (224) 54
Greenwood, C. see N. Foote (214) 347
Greenwood, C. see R.S. Blackmore (219) 244
Gregory, H. see H. Desmond (210) 185
Gregory, R.B. see M.N. Berry (224) 201
Grell, E. see K. Fendler (224) 83
Grenet, D. see J. Codina (216) 104
Gresh, N. see K.-X. Chen (224) 361
Griffiths, J.C., H. Sies, P.J. Meier and T.P.M. Akerboom, Inhibition of taurocholate efflux from rat hepatic canalicular membrane vesicles by glutathione disulfide (213) 34
Griffor, M.C. see E.C. Wiener (224) 33
Grigoriadis, D.E. see H.B. Niznik (209) 71
Grimm, R., F. Lottspeich and W. Rüdiger, Heterogeneity of the amino acid sequence of phytochrome from etiolated oat seedlings (225) 215
Grimmelikhuijzen, C.J.P. and A. Groeger, Isolation of the neuropeptide pGlu-Gly-Arg-Phe-amide from the pennatulid *Renilla köllikeri* (211) 105
Grinblat, L. see M. Dubin (220) 197
Grinstein, S. see O.D. Rotstein (215) 223
Grinstein, S. see P.E. Nasmith (221) 95
Grisebach, H. see D.R. Biggs (220) 223
Grishanova, N.P. see V.Z. Paschenko (214) 28
Grishin, A.V. see E.D. Sverdlov (217) 275
Grishin, A.V. see E.D. Sverdlov (221) 129
Grisolía, S. see E. Roche (221) 231
Grisolía, S. see J.L. Vargas (224) 182
Grisolía, S. see R. Wallace (208) 427
Grisolía, S. see V. Felipo (209) 227
Grisolía, S. see V. Felipo (210) 173
Gritsan, N.P. see D.G. Sushkov (225) 139
Grivell, A.R. see M.N. Berry (224) 201
Gröbner, P. see G. Golderer (222) 322
Grodsky, G.M. see M.L. Gishizky (223) 227
Groeger, A. see C.J.P. Grimmelikhuijzen (211) 105
Gróf, P. see D. Aslanian (219) 202
Groh, M.M.J. see A. Montfoort (226) 101
Gronenborn, A.M., G. Bovermann and G.M. Clore, A ^{1}H-NMR study of the solution conformation of secretin: Resonance assignment and secondary structure (215) 88
Gronenborn, A.M. see G.M. Clore (213) 269
Gronenborn, A.M. see H.R. MacDonald (209) 295
Gronenborn, A.M. see M. Nilges (219) 11
Gronenborn, A.M. see P. Wingfield (215) 160
Grootveld, M. see B. Halliwell (213) 9
Grootveld, M. see B. Halliwell (213) 15
Grootveld, M. see P.C. Moorhouse (213) 23
Gross, B. see H.-J. Kärgel (220) 126
Gross, M. see T. Wagner (212) 317
Grosse, R., R. Schön, P. Jantscheff and G. Pasternak, Immunological and biochemical aspects of malignant transformation and cell proliferation: A report of a meeting held at Berlin-Buch, German Democratic Republic, September 2–4, 1986 (210) 123
Grosse, R. see B. Binas (213) 164
Grubb, A. see M. Abrahamson (216) 229
Grundström, T. see J. Sanchez (208) 194
Gryshin, A.V. see Yu.A. Ovchinnikov (213) 73
Grzesiek, S. and N.A. Dencher, Time-course and stoichiometry of light-induced proton release and uptake during the photocycle of bacteriorhodopsin (208) 337
Grzonka, Z. see E. Kojro (212) 83
Gu, Z.-W. see C. Yang (224) 261
Gualberto, J.M. see M.C. Duque-Magalhães (210) 142
Gualerzi, C.O. see M. Lammi (215) 115
Gubanov, V.V. see Yu.A. Ovchinnikov (223) 169
Gudmundsson, G.H. see D.-A. Lidholm (226) 8
Gueguen, P. see C. Carles (212) 163
Guerin, B. see J.-J. Bessoule (214) 158
Guerrero, A., J.A. Sánchez and A. Darszon, Single-channel activity in sea urchin sperm revealed by the patch-clamp technique (220) 295
Guerrieri, F., R. Scarfò, F. Zanotti, Y.W. Che and S. Papa, Regulatory role of the ATPase inhibitor protein on proton conduction by mitochondrial H^+-ATPase complex (213) 67
Guerritore, A. see S. Lamponi (216) 265
Guibbolini, M.E. and B. Lahlou, Neurohypophyseal peptide inhibition of adenylate cyclase activity in fish gills: The effect of environmental salinity (220) 98
Guitart, X., G. Egea, C. Solsona and J. Marsal, Botulinum neurotoxin inhibits depolarization-stimulated protein phosphorylation in pure cholinergic synaptosomes (219) 219
Gulayeva, L.F. see Ya.Yu. Woldman (212) 53
Gull, K., M. Wilcox and C.R. Birkett, Immunological evidence for a *Physarum* β-tubulin polypeptide possessing an α-tubulin-like carboxyl terminus (219) 31
Gulyaeva, N.V., A.B. Obidin and B.S. Marinov, Modulation of superoxide dismutase by electron donors and acceptors (211) 211
Gunge, N. see S. Oshiro (220) 383
Guntaka, R.V., S. Gowda, H. Wagner and D. Simon, Methylation of the enhancer region of avian sarcoma virus long terminal repeat suppresses transcription (221) 332
Günther, H. see S. Nagata (210) 66

Günther, R., E. Bause and L. Jaenicke, UDP-L-arabinose-hydroxyproline-*O*-glycosyltransferases in *Volvox carteri* (221) 293
Günzler, V. see H.M. Hanauske-Abel (214) 236
Guo, S.Y. see Y.X. Zhu (208) 253
Guschlbauer, W. see A. Kriebardis (213) 297
Guschlbauer, W. see R. Lauster (220) 167
Gutkowska, J., M. Cantin, J. Genest and P. Sirois, Release of immunoreactive atrial natriuretic factor from the isolated perfused rat lung (214) 17
Gutowicz, J. see K. Michalak (219) 233
Gutteridge, J.M.C., A method for removal of trace iron contamination from biological buffers (214) 362
Gutteridge, J.M.C. see P.C. Moorhouse (213) 23
Gylfe, E. see P. Nygren (213) 195

H

Ha, J.-M. see T. Hiwasa (211) 23
Haak, N.P. see H.S. van Walraven (208) 138
Haemmerli, S.D., H.E. Schoemaker, H.W.H. Schmidt and M.S.A. Leisola, Oxidation of veratryl alcohol by the lignin peroxidase of *Phanerochaete chrysosporium*: Involvement of activated oxygen (220) 149
Haenni, A.L. see R.L. Joshi (208) 189
Hafner, R.P., Thyroid hormone uptake into the cell and its subsequent localisation to the mitochondria (224) 251
Haga, K. see T. Kubo (209) 367
Haga, T. see T. Kubo (209) 367
Haiech, J. see M. Dillenschneider (208) 413
Hainque, B. see M.K. Agarwal (217) 221
Haith, C.E. see W.J. Jackson (215) 171
Haka-Risku, T. see R. Soininen (225) 188
Hakam, A., J. McLick, K. Buki and E. Kun, Catalytic activities of synthetic octadeoxyribonucleotides as coenzymes of poly(ADP-ribose) polymerase and the identification of a new enzyme inhibitory site (212) 73
Hakkola, E. see T. Kärki (215) 228
Halban, P.A. see C.J. Rhodes (215) 179
Hale, R.S., K.N. Jordan and P.F. Leadlay, A small, discrete acyl carrier protein is involved in de novo fatty acid biosynthesis in *Streptomyces erythraeus* (224) 133
Hales, C.N. see N.C. Sturgess (208) 397
Hall, A. see R.M. O'Brien (217) 253
Hall, B.S., A.V. Hoffbrand and R.G. Wickremasinghe, Two major tyrosine protein kinases of resting human T lymphocytes are down-regulated following mitotic stimulation (223) 6
Hallett, M.B. see F.A. Al-Mohanna (219) 40
Halliwell, B. and M. Grootveld, The measurement of free radical reactions in humans: Some thoughts for future experimentation (213) 9
Halliwell, B., M. Wasil and M. Grootveld, Biologically significant scavenging of the myeloperoxidase-derived oxidant hypochlorous acid by ascorbic acid: Implications for antioxidant protection in the inflamed rheumatoid joint (213) 15
Halliwell, B. see P.C. Moorhouse (213) 23
Hamamori, Y. see T. Tsuda (208) 39
Hamasur, B. see B. Norling (223) 309
Hamasur, B. see E. Glaser (223) 304
Hamelin, J. see N.G. Seidah (211) 144
Hammond, G.L., D.A. Underhill, C.L. Smith, I.S. Goping, M.J. Harley, N.A. Musto, C.Y. Cheng and C.W. Bardin, The cDNA-deduced primary structure of human sex hormone-binding globulin and location of its steroid-binding domain (215) 100
Hammond, S. see B. Birdsall (217) 106
Hammonds, P., P.N. Schofield and S.J.H. Ashcroft, Glucose regulates preproinsulin messenger RNA levels in a clonal cell line of simian virus 40-transformed B cells (213) 149
Hammonds, P., P.N. Schofield, S.J.H. Ashcroft, R. Sutton and D.W.R. Gray, Regulation and specificity of glucose-stimulated insulin gene expression in human islets of Langerhans (223) 131
Hanai, R., A. Suyama and A. Wada, Vestiges of lost introns in the thermal stability map of DNA (226) 247
Hanauske-Abel, H.M., G. Tschank, V. Günzler, E. Baader and P. Gallop, Pyrroloquinoline quinone and molecules mimicking its functional domains: Modulators of connective tissue formation? (214) 236
Hane, M. see Y. Shimonishi (215) 165
Hanley, M.R. see M.J.O. Wakelam (210) 181
Hanley, M.R. see R.R. Mattingly (223) 11
Hannah-White, C. see G.W.G. Sharp (221) 309
Hansen, C.A. see S.K. Joseph (219) 125
Hansen, H.S. see J.R. Nielsen (218) 93
Hansen, M.T. see A.J. Moody (212) 302
Hansen, M.T. see L. Thim (212) 307
Hansson, G.C. see H. Karlsson (226) 23
Hara, M. see K. Akiyama (225) 168
Harada, N. see M. Ohshima (225) 243
Harder, J. and H. Follmann, Characterization of the free radical in a plant ribonucleotide reductase (222) 171
Hardesty, B. see W. Kudlicki (215) 16
Hardie, D.G. see D. Carling (223) 217
Haritos, A.A., P.P. Yialouris, E.P. Heimer, A.M. Felix and M.A. Rosemeyer, On the molecular size of thymosins (218) 107
Harley, M.J. see G.L. Hammond (215) 100
Harlos, K., C.W.G. Boys, S.K. Holland, M.P. Esnouf and C.C.F. Blake, Structure and order of the protein and carbohydrate domains of prothrombin fragment 1 (224) 97
Harmar, A.J., A. Armstrong, J.C. Pascall, K. Chapman, R. Rosie, A. Curtis, J. Going, C.R.W. Edwards and G. Fink, cDNA sequence of human β-preprotachykinin, the common precursor to substance P and neurokinin A (208) 67
Harris, B.A. see T. Rall (224) 365
Harris, N. see J.N. Yarwood (222) 175
Harrison, T.M. see R.I. Norman (212) 127
Hartmann, B., J. Ramstein and M. Leng, Slow exchanging protons in the Z-form of G-C and A-C alternating polymers by using a rapid dialysis method (225) 11
Hartmann, R.K., D.W. Vogel, B. Kröger, N. Ulbrich and V.A. Erdmann, Studies on rDNA from the extreme thermophilic eubacterium *Thermus thermophilus* HB8: The 23 S rDNA region D (218) 215
Hartshorn, R.T., A.G. Mauk, M.R. Mauk and G.R. Moore, NMR study of the interaction between cytochrome b_5 and cytochrome *c*: Observation of a ternary complex formed by the two proteins and $[Cr(en)_3]^{3+}$ (213) 391
Hartzell, P. see P. Nicotera (209) 139
Hase, S. see S. Koyama (209) 265
Hasegawa, T. see I. Maruyama (220) 89
Hasegawa-Sasaki, H. see T. Sasaki (218) 87
Hashim, F.A. see J. Furmaniak (215) 316
Hashimoto, E. see A. Takeda (210) 169
Hashimoto, S., R. Nakajima, I. Yamazaki, Y. Tatsuno and T. Kitagawa, Oxygen exchange between the Fe(IV)=O heme and bulk water for the A_2 isozyme of horseradish peroxidase (208) 305

Hasselbach, W. and A. Migala, Activation and inhibition of the calcium gate of sarcoplasmic reticulum by high-affinity ryanodine binding (221) 119
Hassinen, I.E. see T. Kärki (215) 228
Hassman, C.F. see J.M. Berman (220) 214
Hata-Tanaka, A., T. Chiba and K. Kakinuma, ESR signals from stimulated and resting porcine blood neutrophils (214) 279
Hatanaka, M. see E. Takano (208) 199
Hatanaka, M. see M. Maki (223) 174
Hattori, T. see M. Shimada (221) 327
Hauck, M. see E. Romanowska (211) 175
Haug, B.L., J.T. Sibley and J.S. Lee, Increased synthesis of poly(ADP-ribose) in isolated liver nuclei from autoimmune NZB/NZW mice (215) 252
Hauptmann, G. see B. Uring-Lambert (217) 65
Hauptmann, G. see J.A. Schifferli (213) 415
Hauska, G. see W. Nitschke (213) 453
Hauska, G. see W. Nitschke (218) 283
Havinga, J.R., P. Lohse and U. Beisiegel, Immunoblotting and ligand blotting of the low-density lipoprotein receptor of human liver, HepG2 cells and HeLa cells (216) 275
Havlíček, J. see L. Morávek (208) 435
Hawrylak, K. and R.A. Stinson, Tetrameric alkaline phosphatase from human liver is converted to dimers by phosphatidylinositol phospholipase C (212) 289
Hayase, Y. see H. Inoue (215) 327
Hayashi, H., M.K. Owada, S. Sonobe, T. Kakunaga, H. Kawakatsu and J. Yano, A 32-kDa protein associated with phospholipase A_2-inhibitory activity from human placenta (223) 267
Hayashi, K. see M. Ohta (222) 79
Hayashi, K. see S. Inoue (218) 17
Hayashi, K. see Y. Furukawa (208) 258
Hayashi, T., S. Yamada, C. Miyaura, H. Tanaka, K. Yamamoto, E. Abe, H. Takayama and T. Suda, Phagocytic cells metabolize 25-hydroxyvitamin D_3 to 10-oxo-19-nor-25-hydroxyvitamin D_3 and a new metabolite, 8α,25-dihydroxy-9,10-seco-4,6,10(19)-cholestatrien-3-one (218) 200
Hayes, M.E., D.J. O'Donoghue, F.W. Ballardie and E.B. Mawer, Peritonitis induces the synthesis of 1α,25-dihydroxyvitamin D_3 in macrophages from CAPD patients (220) 307
Heber, U. see M.E. Salvucci (221) 215
Hedén, L.-O. see M. Forsgren (213) 254
Hederstedt, L., T. Bergman and H. Jörnvall, Processing of *Bacillus subtilis* succinate dehydrogenase and cytochrome *b*-558 polypeptides: Lack of covalently bound flavin in the *Bacillus* enzyme expressed in *Escherichia coli* (213) 385
Hegde, A.N. and M.R. Das, *ras* proteins enhance the phosphorylation of a 38 kDa protein (p38) in rat liver plasma membrane (217) 74
Heimer, E.P. see A.A. Haritos (218) 107
Heinrich, P.C. see T. Andus (221) 18
Heinrichs, M. and H. Schönert, Identification of different quaternary structures of beef heart cytochrome-*c* oxidase by two-dimensional polyacrylamide gel electrophoresis (223) 255
Heitz, F. see Y. Trudelle (216) 11
Heizmann, C.W. see C.P. Schelling (214) 21
Hekman, M. see A.K. Keenan (217) 287
Heldt, H.W. see S. Krömer (226) 352
Helland, D.E., R. Male and K. Kleppe, Separation of damage specific DNA endonuclease activities present in calf thymus (213) 215
Hellingwerf, K.J. see W. Crielaard (225) 6
Hemmings, B.A., M. Schwarz, S.R. Adavani and D.A. Jans, Expression cloning of a cDNA encoding the type II regulatory subunit of the cAMP-dependent protein kinase (209) 219
Hemmings, B.A. see S. Oehen (220) 47
Hempel, J., J.-O. Höög and H. Jörnvall, Mitochondrial aldehyde dehydrogenase: Homology of putative targeting sequence to that of carbamyl phosphate synthetase I revealed by correlation of cDNA and protein data (222) 95
Hendy, G.N. see N.G. Seidah (211) 144
Henly, D.C. see M.N. Berry (224) 201
Henne, V., A. Piiper and H.-D. Söling, Inositol 1,4,5-trisphosphate and 5′-GTP induce calcium release from different intracellular pools (218) 153
Henning, U. see G. Ried (223) 387
Henry, J.-P. see B. Gasnier (222) 215
Henry, M. see J. Borg (213) 406
Henschen, A. see E. Töpfer-Petersen (226) 38
Henseling, J. and K.-H. Röhm, Unusual solvent isotope effects on the aminoacylase-catalyzed hydrolysis of acetylamino acids (219) 27
Herbst, D. see H.A. Weich (213) 89
Herkovitz, J. see N.E. Fink de Cabutti (223) 330
Herlyn, M. see J. Thurin (208) 17
Hermann, P. see E. Kojro (212) 83
Hernell, O. see L. Bläckberg (217) 37
Herranz, J. see M.A. Jiménez (221) 320
Herrmann, R.G. see T. Jansen (216) 234
Herrou, M. see A. Benhaim (223) 321
Hers, H.-G. see E. Mertens (221) 124
Hervé, P. see A. Zachowski (223) 315
Herz, A. see Y. Shimohigashi (222) 71
Hess, B. see K. Gerwert (213) 39
Hess, B. see M. Engelhard (222) 275
Hetherington, A. see M. Dillenschneider (208) 413
Heymans, F., C.D. Silva, N. Marrec, J.-J. Godfroid and M. Castagna, Alkyl analogs of diacylglycerol as activators of protein kinase C (218) 35
Hibino, T., S. Izaki and M. Izaki, Purification of epidermal plasminogen activator inhibitor (208) 273
Hicke, B.J. see J.G. Wise (223) 395
Hidaka, Y. see Y. Shimonishi (215) 165
Hietter, H. see J. Borg (213) 406
Higashida, H. and D.A. Brown, Bradykinin inhibits potassium (M) currents in N1E-115 neuroblastoma cells: Responses resemble those in NG108-15 neuroblastoma × glioma hybrid cells (220) 302
Higashida, H. and D.A. Brown, Membrane current responses to intracellular injections of inositol 1,3,4,5-tetrakisphosphate and inositol 1,3,4-trisphosphate in NG108-15 hybrid cells (208) 283
Higashiyama, H. and F. Tokunaga, Cause of the blue shift of the absorption spectrum of tetranitromethane-treated bacteriorhodopsin (218) 287
Higgins, C.F. see J.S. Evans (208) 211
Higuchi, T. see M. Shimada (221) 327
Higuchi, T. see S. Kawai (210) 61
Higuchi, T. see T. Umezawa (218) 255
Higuchi, W. see C. Fukazawa (224) 125
Hikichi, K. see M. Ikura (219) 17
Hilditch, P., H. Thomas and L.J. Rogers, Two processes for the breakdown of the Q_B protein of chloroplasts (208) 313
Hill, H.A.O. see M.J. Green (216) 31
Hill, R.M. see D.L. Christie (214) 45
Hiltunen, J.K. see T. Kärki (215) 228
Hinderer, W. see K. Tiemann (213) 324
Hinderer, W., U. Flentje and W. Barz, Microsomal isoflavone 2′- and 3′-hydroxylases from chickpea (*Cicer arietinum* L.) cell suspensions induced for pterocarpan phytoalexin formation (214) 101
Hingorani, V.N. and Y.-K. Ho, A structural model for the α-subunit of transducin: Implications of its role as a molecular switch in the visual signal transduction mechanism (220) 15

Hinterstoisser, B. and G.A. Peschek, Fluorimetric pH measurement in whole cells of dark aerobic and anaerobic cyanobacteria (217) 169
Hinuma, Y. see M. Fujii (223) 299
Hirano, H. see H. Kagawa (226) 145
Hirano, T. see T. Andus (221) 18
Hirasawa, M., J.M. Boyer, K.A. Gray, D.J. Davis and D.B. Knaff, The interaction of ferredoxin-linked sulfite reductase with ferredoxin (221) 343
Hirata, Y., H. Yoshimi, S. Takata, H. Matsubara, Y. Takagi, T. Iida, N. Chino, T.X. Watanabe, T. Kimura and S. Sakakibara, Binding of synthetic β-human atrial natriuretic peptide to cultured rat vascular smooth muscle cells (219) 369
Hird, S.M., T.A. Dyer and J.C. Gray, The gene for the 10 kDa phosphoprotein of photosystem II is located in chloroplast DNA (209) 181
Hirose, S. see K. Oda (214) 135
Hirose, T. see T. Kubo (209) 367
Hirsch, U. see C.N. Serhan (217) 242
Hirst, K. see P.W. Piper (220) 177
Hirst, T.R. see J. Sanchez (208) 194
Hishida, T. see M. Ueda (220) 31
Hishinuma, F. see T. Yamakuni (223) 117
Hitaka, T. see T. Mizutani (226) 227
Hiwasa, T., S. Yokoyama, J.-M. Ha, S. Noguchi and S. Sakiyama, c-Ha-*ras* gene products are potent inhibitors of cathepsins B and L (211) 23
Hiwatashi, A., N. Sakihama, M. Shin and Y. Ichikawa, Heterogeneity of adrenocortical ferredoxin (209) 311
Hiyama, T., T. Watanabe, M. Kobayashi and M. Nakazato, Interaction of chlorophyll *a'* with the 65 kDa subunit protein of photosystem I reaction center (214) 97
Ho, C.-L. and J.-L. Ko, Hornetin: the lethal protein of the hornet *(Vespa flavitarsus)* venom (209) 18
Ho, Y.-K. see V.N. Hingorani (220) 15
Ho, Y.-S. see J.R. Didsbury (211) 160
Hock, D. see M. Gagelmann (225) 251
Hodson, H.F. see J. Dawson (214) 171
Hodson, H.F. see R.W. Bonser (209) 134
Hoehn-Berlage, M. see R. Arkowitz (217) 21
Hofer, E. see E. Pöschl (226) 96
Hoffbrand, A.V. see B.S. Hall (223) 6
Hoffbrand, A.V. see R.G. Wickremasinghe (220) 52
Hoffman, B.M. see J. Telser (214) 117
Hoffman, E.J. see S.C. Huang (216) 128
Hoffmann, A. and P. Dimroth, Stereochemistry of the methylmalonyl-CoA decarboxylation reaction (220) 121
Hofmann, K.P. see K.M.P. Kamps (208) 241
Höfte, H., S. Buyssens, M. Vaeck and J. Leemans, Fusion proteins with both insecticidal and neomycin phosphotransferase II activity (226) 364
Hogenkamp, H.P.C., H. Follmann and R.K. Thauer, Ribonucleotide reductase in cell extracts of *Methanobacterium thermoautotrophicum* (219) 197
Höhne, B. see B. Stecher (223) 25
Hökfelt, T. see T. Miyamoto (216) 123
Holland, S.K. see K. Harlos (224) 97
Holloszy, J.O. see A. Klip (224) 224
Holmes, C.F.B., A new method for the selective isolation of phosphoserine-containing peptides (215) 21
Holmes, L. see D.A. Spandidos (218) 41
Holmgren, J. see J. Sanchez (208) 194
Holmquist, B. see T. Fairwell (222) 99
Holmsen, H. see O.-B. Tysnes (218) 68
Holst, J.J., C. Ørskov, O.V. Nielsen and T.W. Schwartz, Truncated glucagon-like peptide I, an insulin-releasing hormone from the distal gut (211) 169
Holzhöfer, A. see A.K. Keenan (217) 287
Homan, W., C. Sigon, W. van den Briel, R. Wagter, H. de Nobel, D. Mesland, A. Musgrave and H. van den Ende, Transport of membrane receptors and the mechanics of sexual cell fusion in *Chlamydomonas eugametos* (215) 323
Homburger, V. see M. Toutant (215) 339
Homburger, V. see M. Toutant (222) 51
Homma, M. see D.S. Samuels (209) 231
Homo, F. see A. Brossi (223) 77
Honek, J.F. see E.W. Szczepan (211) 239
Hönes, J., K.-D. Jany, G. Pfleiderer and A.F.V. Wagner, An integrated prediction of secondary, tertiary and quaternary structure of glucose dehydrogenase (212) 193
Hong, O. see D. Ukena (215) 203
Höög, J.-O. see J. Hempel (222) 95
Hoppe, E.M. see U. Hoppe (208) 26
Hoppe, J., L. Schumacher, W. Eichner and H.A. Weich, The long 3'-untranslated regions of the PDGF-A and -B mRNAs are only distantly related (223) 243
Hoppe, J. see H.A. Weich (213) 89
Hoppe, U., E.M. Hoppe, B.M. Peskar and B.A. Peskar, Radioimmunoassay for leukotriene E_4: Use for determination of total sulfidopeptide-leukotriene release from rat gastric mucosa (208) 26
Höppener, J.W.M., P.H. Steenbergh, R.J.C. Slebos, P. de Pagter-Holthuizen, B.A. Roos, M. Jansen, J.L. Van den Brande, J.S. Sussenbach, H.S. Jansz and C.J.M. Lips, Expression of insulin-like growth factor-I and -II genes in rat medullary thyroid carcinoma (215) 122
Höppener, J.W.M. see P.H. Steenbergh (209) 97
Hopple, S.L. see M. Bushfield (222) 299
Horii, A. see N. Tomita (225) 113
Horimoto, H. see Y. Shimohigashi (222) 251
Horn, R.S. see S.I. Walaas (220) 311
Hornby, E.J. see K.S. Authi (213) 95
Horniak, L., E. Kutejová and P. Balgavý, Effect of phase transitions in hydrated 1,2-dipalmitoylphosphatidylethanolamine bilayers on the spin probe order parameter (224) 283
Hornig, H. see B. Epe (213) 443
Horton, P. see K. Oxborough (221) 211
Horuk, R. see J.J. Huang (223) 294
Hoshi, M., E. Nishida, Y. Miyata, H. Sakai, T. Miyoshi, H. Ogawara and T. Akiyama, Protein kinase C phosphorylates tau and induces its functional alterations (217) 237
Hoshijima, M. see T. Tanimoto (226) 291
Hossenlopp, P., D. Seurin, B. Segovia-Quinson and M. Binoux, Identification of an insulin-like growth factor-binding protein in human cerebrospinal fluid with a selective affinity for IGF-II (208) 439
Hostikka, S.L. and K. Tryggvason, Extensive structural differences between genes for the α_1 and α_2 chains of type IV collagen despite conservation of coding sequences (224) 297
Hostikka, S.L., M. Kurkinen and K. Tryggvason, Nucleotide sequence coding for the human type IV collagen α_2 chain cDNA reveals extensive homology with the NC-1 domain of α_1(IV) but not with the collagenous domain or 3'-untranslated region (216) 281
Hottiger, T., T. Boller and A. Wiemken, Rapid changes of heat and desiccation tolerance correlated with changes of trehalose content in *Saccharomyces cerevisiae* cells subjected to temperature shifts (220) 113
Hough, D.W. see L.D. Smith (225) 277
Houslay, M.D. see D. Gawler (216) 94
Houslay, M.D. see F. Irvine (208) 455
Houslay, M.D. see R.M. O'Brien (212) 281
Houslay, M.D. see R.M. O'Brien (217) 253
Houston, K. see O.D. Rotstein (215) 223
Houston, M.E., M.D. Lingley, D.S. Stuart and R.W. Grange, Myosin light chain phosphorylation in intact human muscle (219) 469

Houwing, C.J. and E.M.J. Jaspars, Coat protein blocks the in vitro transcription of the virion RNAs of alfalfa mosaic virus (209) 284
Houwing, C.J. and E.M.J. Jaspars, In vitro evidence that the coat protein is the programming factor in alfalfa mosaic virus-induced RNA synthesis (221) 337
Howe, C.J. see P. Nicholson (221) 110
Howe, P.H. and A.A. Abdel-Latif, Phorbol ester-induced protein phosphorylation and contraction in sphincter smooth muscle of rabbit iris (215) 279
Howell, B.A. see D.P. Ringer (224) 59
Howell, S.L. see P.M. Jones (219) 139
Howells, R.D. see J. Furmaniak (215) 316
Howland, R.J. and A.D. Benning, Differential effects of noradrenaline and glucagon on lipolysis and fatty-acid utilization in brown adipose tissue (208) 128
Hozumi, N. see W.S. Trimble (219) 70
Hsi, K.L. see Y.X. Zhu (208) 253
Hsu, B.-D., W.-J. Lin, J.-Y. Lee and R.L. Pan, Evidence for structural distinction between PS II_α and PS II_β reaction centers (217) 53
Hsu, S.-Y., T. Noumi, M. Takeyama, M. Maeda, S. Ishibashi and M. Futai, β-Subunit of *Escherichia coli* F_1-ATPase: An amino acid replacement within a conserved sequence (G-X-X-X-X-G-K-T/S) of nucleotide-binding proteins (218) 222
Hsu, Y.C., D.P. Bloxham and I.G. Giles, Phosphorylation of type-L pyruvate kinase in intact hepatocytes: Localisation of the phosphorylation site in response to both glucagon and the Ca^{2+}-linked agonist phenylephrine (218) 1
Huang, J.J., R.C. Newton, R. Horuk, J.B. Matthew, M. Covington, K. Pezzella and Y. Lin, Muteins of human interleukin-1 that show enhanced bioactivities (223) 294
Huang, J.-K. see C.-S. Wang (222) 135
Huang, S.C., B.A. Williams, J.R. Barrio, J. Krivokapich, C. Nissenson, E.J. Hoffman and M.E. Phelps, Measurement of glucose and 2-deoxy-2-[^{18}F]fluoro-D-glucose transport and phosphorylation rates in myocardium using dual-tracer kinetic experiments (216) 128
Hubbard, J.A.M. see R.W. Mansfield (220) 74
Huber, M. and K. Lerch, The influence of copper on the induction of tyrosinase and laccase in *Neurospora crassa* (219) 335
Huber, S.C. see G.H. Walker (213) 375
Hucho, F., H. Krüger, I. Pribilla and U. Oberdieck, A 40 kDa inhibitor of protein kinase C purified from bovine brain (211) 207
Huck, S. and M.-P. Lefranc, Rearrangements to the JP1, JP and JP2 segments in the human T-cell rearranging gamma gene (TRGγ) locus (224) 291
Huck, S., G. Keyeux, N. Ghanem, M.-P. Lefranc and G. Lefranc, A gamma 3 hinge region probe: first specific human immunoglobulin subclass probe (208) 221
Hue, L. see M.I. Darville (224) 317
Hufford, C.D. see A. Brossi (223) 77
Huggins, J.P. and P.J. England, Evidence for a phosphorylation-induced conformational change in phospholamban from the effects of three proteases (217) 32
Hughes, G.J. see B. Roth (221) 172
Hughes, J. see W.W.Y. Lo (220) 155
Hughes, J. see W.W.Y. Lo (220) 327
Hughes, J. see W.W.Y. Lo (224) 1
Hughes, J. see W.W.Y. Lo (226) 67
Hughes, W.A. see S. Gilroy (212) 133
Hui, S.-W. see R.M. Epand (209) 257
Hunt, J.E. see T.J. Michalski (226) 72
Hunter, C.N. see M.K. Ashby (213) 245
Hunter, C.N. see P.J. Nixon (209) 83
Hunyady, L., B. Sarkadi, E.J. Cragoe, jr, A. Spät and G. Gárdos, Activation of sodium-proton exchange is not a prerequisite for Ca^{2+} mobilization and aggregation in human platelets (225) 72
Hunziker, W. see G. Sebastio (208) 460
Hurst, N.P. see J.K. French (212) 242
Huss, S. see P.F. Torrence (212) 267
Hussein, A. see B. Leighton (225) 93
Huston, G.E. see S. Patton (216) 151
Huszti, Z. and E. Tyihák, Formation of formaldehyde from *S*-adenosyl-L-[*methyl*-^{3}H]methionine during enzymic transmethylation of histamine (209) 362
Huynh-Dinh, T. see G. Bloch (219) 464

I

Iadarola, P. see M. Galliano (208) 364
Ibeanu, G.C. see L.L. Muldrow (213) 249
Ichihara, A. see T. Nakamura (224) 311
Ichihara, C. see M. Yamazaki (208) 147
Ichikawa, Y. see A. Hiwatashi (209) 3
Ichimura, T., T. Isobe, T. Okuyama, T. Yamauchi and H. Fujisawa, Brain 14-3-3 protein is an activator protein that activates tryptophan 5-monooxygenase and tyrosine 3-monooxygenase in the presence of Ca^{2+},calmodulin-dependent protein kinase II (219) 79
Ichiyama, A. see T. Kubo (209) 367
Ido, M., K. Sekiguchi, U. Kikkawa and Y. Nishizuka, Phosphorylation of the EGF receptor from A431 epidermoid carcinoma cells by three distinct types of protein kinase C (219) 215
Igarashi, K. see T. Kurokawa (213) 189
Igarashi, K. see U. Kikkawa (217) 227
Igarashi, K. see U. Kikkawa (223) 212
Igarashi, K. see Y. Ono (226) 125
Iglesias, A.A. see C.S. Andreo (213) 1
Iida, T. see Y. Hirata (219) 369
IJzerman, A.P. see P.J.M. van Galen (223) 197
Ikeda, F. see Y. Furukawa (208) 258
Ikeda, K. see S. Inoue (218) 17
Ikeda-Saito, M. and T. Inubushi, Proton magnetic resonance of the bovine spleen green heme-protein (214) 111
Ikegami, S. see N. Tsuchimori (218) 205
Ikehara, Y. see K. Oda (214) 135
Ikenaka, T. see S. Koyama (209) 265
Ikeuchi, M. and Y. Inoue, Specific ^{125}I labeling of D1 (herbicide-binding protein): An indication that D1 functions on both the donor and acceptor sides of photosystem II (210) 71
Ikeyama, S., Purification and characterization of a recombinant human IgE Fcε fragment lacking the C4 domain (224) 306
Ikezawa, H. see R. Taguchi (225) 273
Ikura, M., O. Minowa, M. Yazawa, K. Yagi and K. Hikichi, Sequence-specific assignments of downfield-shifted amide proton resonances of calmodulin: Use of two-dimensional NMR analysis of its tryptic fragments (219) 17
Imahori, K. see S. Imajoh (215) 274
Imaizumi, M. see M. Tanokura (209) 77

Imajoh, S., H. Kawasaki, Y. Emori, S. Ishiura, Y. Minami, H. Sugita, K. Imahori and K. Suzuki, A fragment of an endogenous inhibitor produced in *Escherichia coli* for calcium-activated neutral protease (CANP) retains an inhibitory activity (215) 274
Imajoh, S. see K. Ishidoh (223) 69
Imajoh, S. see K. Ishidoh (226) 33
Imajoh, S. see K. Suzuki (220) 271
Imamura, A. see H. Toh (219) 279
Imazu, S. see Y. Shimohigashi (222) 251
Imbach, J.-L. see P.F. Torrence (212) 267
Imura, H. see Y. Seino (223) 74
Inagaki, F., C. Kodama, M. Suzuki and A. Suzuki, Analysis of NMR spectra of sugar chains of glycolipids by 1D homonuclear Hartmann-Hahn and NOE experiments (219) 45
Inagaki, F., D. Kohda, C. Kodama and A. Suzuki, Analysis of NMR spectra of sugar chains of glycolipids by multiple relayed COSY and 2D homonuclear Hartman-Hahn spectroscopy (212) 91
Inagaki, N. see Y. Seino (223) 74
Inagami, T. see H. Kurose (219) 375
Inesi, G. see C. Sumbilla (210) 31
Inestrosa, N.C. see E. Brandan (213) 159
Infante, J.P., De novo CDP-choline-dependent glycerophosphorylcholine synthesis and its involvement as an intermediate in phosphatidylcholine synthesis (214) 149
Inge-Vechtomov, S.G. see V.V. Kushnirov (215) 257
Ingold, K.U. see K.H. Cheeseman (209) 191
Inoue, H. see Y. Kubota (212) 159
Inoue, H., Y. Hayase, S. Iwai and E. Ohtsuka, Sequence-dependent hydrolysis of RNA using modified oligonucleotide splints and RNase H (215) 327
Inoue, J., M. Seiki and M. Yoshida, The second pX product p27$^{x\text{-}III}$ of HTLV-1 is required for gag gene expression (209) 187
Inoue, M. see M. Doi (213) 265
Inoue, S., K. Ohkura, K. Ikeda and K. Hayashi, Amino acid sequence of a cytotoxin-like basic protein with low cytotoxic activity from the venom of the Thailand cobra *Naja naja siamensis* (218) 17
Inoue, T. see T. Gojobori (208) 231
Inoue, Y. see I. Vass (211) 215
Inoue, Y. see M. Ikeuchi (210) 71
Inouye, M. see J.-M. Bolla (224) 213
Inubushi, T. see M. Ikeda-Saito (214) 111
Inui, T. see A. Kawamori (217) 134
Ireland, C.M. and B.W. Stewart, DNA-mediated gene transfer as an indicator of DNA damage and its repair by recipient cells (212) 173
Irvine, F., N.J. Pyne and M.D. Houslay, The phorbol ester TPA inhibits cyclic AMP phosphodiesterase activity in intact hepatocytes (208) 455
Irvine, R.F. see L.G.J. Tertoolen (214) 365
Ischenko, K.A. see Yu.A. Ovchinnikov (223) 169
Ishibashi, S. see S.-Y. Hsu (218) 222
Ishibashi, T. see Y. Kuroda (224) 137
Ishida, T. see M. Doi (213) 265
Ishidoh, K., S. Imajoh, Y. Emori, S. Ohno, H. Kawasaki, Y. Minami, E. Kominami, N. Katunuma and K. Suzuki, Molecular cloning and sequencing of cDNA for rat cathepsin H: Homology in pro-peptide regions of cysteine proteinases (226) 33
Ishidoh, K., T. Towatari, S. Imajoh, H. Kawasaki, E. Kominami, N. Katunuma and K. Suzuki, Molecular cloning and sequencing of cDNA for rat cathepsin L (223) 69
Ishihara, H. and K. Shimura, Further evidence for the presence of a thiazoline ring in the isoleucylcysteine dipeptide intermediate in bacitracin biosynthesis(226) 319
Ishimatsu, Y. see N. Sakai (222) 341
Ishino, F. see M.D. Song (221) 167
Ishiura, S. see S. Imajoh (215) 274
Ishiura, S., T. Yoshimoto and C.A. Villee, ATP-dependent proteolysis, a fine system or a meaningless one? A reply (215) 195
Ishizaki, H. see H. Jhoti (219) 419
Isobe, T. see T. Ichimura (219) 79
Isobe, T., Y.-I. Fang, D. Muno, T. Okuyama, D. Ohmori and F. Yamakura, Amino acid sequence of iron-superoxide dismutase from *Pseudomonas ovalis* (223) 92
Isogai, Y., Yasusi Yamamoto, Yasuyuki Yamamoto and M. Nishimura, Isolation of photosystem II reaction center complex by affinity chromatography with the peripheral 33-kDa polypeptide as ligand (224) 71
Israelsson, M. see M. Forsgren (213) 254
Itami, M., T. Kuroki and K. Nose, Induction of c-*fos* proto-oncogene by a chemotactic peptide in human peripheral granulocytes (222) 289
Ito, E. see N. Murazumi (218) 131
Itoh, K., K. Ueno and S. Natori, Counteraction by 20-hydroxyecdysone of the effect of juvenile hormone on phosphorylation of ribosomal protein S6 (213) 85
Itoh, S. see Y. Kuwana (219) 360
Itoh, T., I. Yamashita and S. Fukui, Nucleotide sequence of the α-amylase gene (ALP1) in the yeast *Saccharomycopsis fibuligera* (219) 339
Ivanov, I., L. Gigova and E. Jay, Chemical synthesis and expression in *E. coli* of a human Val^{8}-calcitonin gene by fusion to a synthetic human interferon-γ gene (210) 56
Ivanov, P.L. see S.A. Limborska (212) 208
Ivanov, V.T. see A.S. Arseniev (213) 283
Ivanova, A.N. see B.N. Goldstein (217) 212
Ivanovienė, L. see B. Kholodenko (223) 247
Ivell, R. see S. Fehr (210) 45
Iwabuchi, M. see K. Mikami (223) 273
Iwai, M. and K. Jungermann, Possible involvement of eicosanoids in the actions of sympathetic hepatic nerves on carbohydrate metabolism and hemodynamics in perfused rat liver (221) 155
Iwai, S. see H. Inoue (215) 327
Iwamori, M. see N. Rodrig (221) 315
Iwanaga, S. see K. Akiyama (225) 168
Iwane, M. see T. Kurokawa (213) 189
Izaki, M. see T. Hibino (208) 273
Izaki, S. see T. Hibino (208) 273
Izard, J. see G. Landemore (209) 29
Izumi, T. see S. Kitamura (213) 169
Izumiya, N. see S. Ono (220) 332
Izumiya, N. see Y. Shimohigashi (222) 251

J

Jackson, A.P., M.P. Timmerman, C.R. Bagshaw and C.C. Ashley, The kinetics of calcium binding to fura-2 and indo-1 (216) 35
Jackson, J.B. see N.P.J. Cotton (219) 88
Jackson, J.B. see W. Crielaard (225) 6
Jackson, M.R. see R.B. Corser (213) 448

Jackson, W.J., P.J. Kiley, C.E. Haith, S. Kaplan and R.C. Prince, On the role of the light-harvesting B880 in the correct insertion of the reaction center of *Rhodobacter capsulatus* and *Rhodobacter sphaeroides* (215) 171
Jacobson, A.E. see B. DeCosta (223) 335
Jacobson, A.E. see D.J.J. Carr (224) 272
Jacobson, K.A., J. Zimmet, R. Schulick, S. Barone, J.W. Daly and K.L. Kirk, Adenosine analogs with covalently attached lipids have enhanced potency at A_1-adenosine receptors (225) 97
Jacobson, K.A. see D. Ukena (209) 122
Jacobsson, A., B. Cannon and J. Nedergaard, Physiological activation of brown adipose tissue destabilizes thermogenin mRNA (224) 353
Jacquemin, J.-M. see G. Lenzen (219) 254
Jacquot, J.-P. and P. Decottignies, Further evidence for a role of sulfhydryls in the thioredoxin dependent activation of corn NADP-malate dehydrogenase: Use of a cysteine free mutant of *E. coli* thioredoxin (209) 87
Jacquot, R. see P. Mayeux (211) 229
Jaenicke, L. see R. Günther (221) 293
Jahagirdar, A.P., G. Milton, T. Viswanatha and R.G.H. Downer, Calcium involvement in mediating the action of octopamine and hypertrehalosemic peptides on insect haemocytes (219) 83
Jähnig, F., R. Bülow, T. Baltz and P. Overath, Secondary structure of the variant surface glycoproteins of trypanosomes (221) 37
Jähnig, F. see I. Riede (215) 145
Jainazarov, A.B. see S.S. Kolesnikov (222) 37
Jakobs, K.H. see P. Gierschik (224) 219
Jalalpour, S. see I. Dodd (209) 13
Jallat, S. see L.-H. Tessier (208) 183
Jamall, I.S., Differential effects of cadmium on cytosolic and mitochondrial glutathione levels in the rat heart (214) 62
Jamall, I.S. see T.W. Simmons (218) 251
Jamaluddin, M.P. see L.K. Krishnan (212) 213
James, K. see J. Simpson (217) 62
Janković, V. see S. Suzić (216) 287
Jänne, J. see P. Leinonen (215) 68
Jänne, O.A. see P. Leinonen (215) 68
Jans, D.A. see B.A. Hemmings (209) 219
Jans, D.A. see S. Oehen (220) 47
Jansen, M. see J.W.M. Höppener (215) 122
Jansen, M. see P. de Pagter-Holthuizen (214) 259
Jansen, T., C. Rother, J. Steppuhn, H. Reinke, K. Beyreuther, C. Jansson, B. Andersson and R.G. Herrmann, Nucleotide sequence of cDNA clones encoding the complete '23 kDa' and '16 kDa' precursor proteins associated with the photosynthetic oxygen-evolving complex from spinach (216) 234
Jansson, C. see T. Jansen (216) 234
Jansz, H.S. see J.W.M. Höppener (215) 122
Jansz, H.S. see P.D. Baas (218) 119
Jansz, H.S. see P.H. Steenbergh (209) 97
Jantscheff, P. see R. Grosse (210) 123
Jany, K.-D. see J. Hönes (212) 193
Jarett, L. see E.H.A. Wong (213) 419
Jarett, L. see K.L. Kelly (209) 238
Jaspars, E.M.J. see C.J. Houwing (209) 284
Jaspars, E.M.J. see C.J. Houwing (221) 337
Jaubert, J. see F. Baklouti (223) 59
Jay, E. see I. Ivanov (210) 56
Jeffery, W.A. see E.J. Wawrzynczak (219) 51
Jeffrey, F.M.H. see C.R. Malloy (212) 58
Jenkins, G.I. see L.K. Barnett (224) 287
Jennings, P.R. see P.J. Wise (222) 17
Jensen, B. see J.R. Nielsen (218) 93
Jensen, L.H. see L.C. Sieker (208) 73
Jensen, L.H. see L.C. Sieker (209) 261
Jensen, R.A. see S. Ahmad (216) 133
Jeremy, J. see B. Pearce (211) 73
Jergil, B. see P.-M. Melin (223) 87
Ježek, P., Sulfhydryl groups are involved in H^+ translocation via the uncoupling protein of brown adipose tissue mitochondria (211) 89
Jhoti, H., A.N. McLeod, T.L. Blundell, H. Ishizaki, H. Nagasawa and A. Suzuki, Prothoracicotropic hormone has an insulin-like tertiary structure (219) 419
Ji, L.L., F.W. Stratman and H.A. Lardy, Chronic exercise training alters kinetic properties of rat skeletal muscle and myocardial lactate dehydrogenase (208) 297
Jiménez, M.A., J.L. Nieto, J. Herranz, M. Rico and J. Santoro, ^{1}H NMR and CD evidence of the folding of the isolated ribonuclease 50–61 fragment (221) 320
Jin, Y. and R.R. Crichton, Iron transfer from ferritin to transferrin: Effect of serum factors (215) 41
Jincharadze, A.G. see S.A. Limborska (212) 208
Joffre, M. see P. Duchatelle (217) 11
Johanningmeier, U., U. Bodner and G.F. Wildner, A new mutation in the gene coding for the herbicide-binding protein in *Chlamydomonas* (211) 221
Johansen, H.R. see T. Lund (208) 369
John, A. see G. .Theiss (218) 159
Johnson, J. see L.L. Muldrow (213) 249
Johnson, M.K., J.E. Morningstar, M. Oliver, and F.E. Frerman, Electron paramagnetic resonance and magnetic circular dichroism studies of electron-transfer flavoprotein-ubiquinone oxidoreductase from pig liver (226) 129
Jollès, P. see J. Berthou (218) 55
Jondal, M. see C. Nordstedt (220) 57
Jones, P.M., J.M. Fyles, S.J. Persaud and S.L. Howell, Catecholamine inhibition of Ca^{2+}-induced insulin secretion from electrically permeabilised islets of Langerhans (219) 139
Jongejan, J.A. see R.A. van der Meer (221) 299
Jordan, K.N. see R.S. Hale (224) 133
Jordan, L. see A.J. Aarsman (219) 176
Jordan, P.M. and M.J. Warren, Evidence for a dipyrromethane cofactor at the catalytic site of *E. coli* porphobilinogen deaminase (225) 87
Jordan, R.E. see S.O. Brennan (219) 431
Jørgensen, P.L. see D.A. Klærke (216) 211
Jörnvall, H. see J. Hempel (222) 95
Jörnvall, H. see L. Blomqvist (211) 127
Jörnvall, H. see L. Hederstedt (213) 385
Jörnvall, H. see O.U. Beg (216) 270
Jörnvall, H. see T. Fairwell (222) 99
Jörnvall, H. see Z. Chen (226) 43
Joseph, S.K., C.A. Hansen and J.R. Williamson, Inositol 1,3,4,5-tetrakisphosphate increases the duration of the inositol 1,4,5-trisphosphate-mediated Ca^{2+} transient (219) 125
Joseph, S.K. see C.V. Nicchitta (209) 243
Joshi, R.L., H.G. Faulhammer, A.L. Haenni and M. Sprinzl, Fluorescence labeling of an aminoacyl-tRNA at the 3′-end and its interaction with elongation factor Tu·GTP (208) 189
Joshi, R.L. see H.G. Faulhammer (217) 203
Joubert, R. see N.E. Fink de Cabutti (223) 330
Jovin, T.M. see D.M. Soumpasis (213) 341
Joyard, J. see J. Covès (208) 401
Joziasse, D.H., H.C.M. Damen, M. de Jong-Brink, H.T. Edzes and D.H. Van den Eijnden, Identification of a UDP-Gal:β-galactoside β1→3-galactosyltransferase in the albumen gland of the snail *Lymnaea stagnalis* (221) 139
Judah, J.D. see R.C. Foreman (219) 75
Julià, P. see T. Fairwell (222) 99
Jullienne, A. see S. Minvielle (223) 63

Junge, W. see S. Engelbrecht (219) 321
Jungermann, K. see B. Christ (221) 375
Jungermann, K. see G.P. Püschel (219) 145
Jungermann, K. see H. Bartels (221) 277
Jungermann, K. see M. Iwai (221) 155

K

Kabakov, A.E. see M.A. Glukhova (218) 292
Kader, J.C. see A. Oursel (219) 393
Kadiri, C. see J. Masliah (222) 11
Kador, P. see D. Carper (220) 209
Kagawa, H., F. Yamauchi and H. Hirano, Soybean basic 7 S globulin represents a protein widely distributed in legume species (226) 145
Kagawa, Y. see S. Ohta (226) 171
Kagramanova, V.K. see A.S. Mankin (219) 269
Kähäri, V.-M., P. Multimäki and E. Vuorio, Elevated proα2(I) collagen mRNA levels in cultured scleroderma fibroblasts result from an increased transcription rate of the corresponding gene (215) 331
Kahn, R. see J. Falloon (209) 352
Kai, H., H. Kanaide, T. Matsumoto and M. Nakamura, 8-Bromoguanosine 3′:5′-cyclic monophosphate decreases intracellular free calcium concentrations in cultured vascular smooth muscle cells from rat aorta (221) 284
Kai, H., H. Kanaide, T. Matsumoto, Y. Shogakiuchi and M. Nakamura, Adenosine decreases intracellular free calcium concentrations in cultured vascular smooth muscle cells from rat aorta (212) 119
Kaiho, S. see J. Abe (226) 58
Kaise, N. see T. Nakamura (224) 311
Kaiser, R. see T. Fairwell (222) 99
Kakei, M., R.P. Kelly, S.J.H. Ashcroft and F.M. Ashcroft, The ATP-sensitivity of K^+ channels in rat pancreatic B-cells is modulated by ADP (208) 63
Kakei, M. see F.M. Ashcroft (215) 9
Kakinuma, K. see A. Hata-Tanaka (214) 279
Kakinuma, K. see A. Tomoda (219) 472
Kąkol, I. see D. Szczęsna (210) 177
Kakol, I. see Yu.S. Borovikov (223) 409
Kakunaga, T. see H. Hayashi (223) 267
Kalderon, B., A. Gopher and A. Lapidot, A quantitative analysis of the metabolic pathways of hepatic glucose synthesis in vivo with ^{13}C-labeled substrates (213) 209
Kalderon, B., A. Gopher and A. Lapidot, Can two intramitochondrial oxaloacetate pools unravel the dispute? A reply to Katz (220) 259
Kalia, A. see K. Gopalakrishna (215) 95
Kaller, D. see E.W. Szczepan (211) 239
Kambouris, A.M. see P.D. Roach (222) 159
Kamerling, J.P. see J.A. van Kuik (221) 150
Kamihara, T. see Y. Uejima (214) 127
Kamoun, P.P., E. Schneider and M. Dy, Superoxide-induced deimination of arginine in hematopoietic cells (226) 285
Kampf, K. see W.B. Macklin (223) 417
Kamps, K.M.P. and K.P. Hofmann, ATP can promote activation and deactivation of the rod cGMP-phosphodiesterase: Kinetic light scattering on intact rod outer segments (208) 241
Kanai, R. see J.-I. Ohnishi (219) 347
Kanaide, H. see H. Kai (212) 119
Kanaide, H. see H. Kai (221) 284
Kanaide, H., Y. Shogakiuchi and M. Nakamura, The norepinephrine-sensitive Ca^{2+}-storage site differs from the caffeine-sensitive site in vascular smooth muscle of the rat aorta (214) 130
Kanaoka, Y. see H. Nakayama (208) 278
Kanda, K. see H. Toh (219) 279
Kang, M.S. see M. Obara (213) 261
Kangawa, K. see T. Kubo (209) 367
Kannagi, R. see E. Takano (208) 199
Kano-Murakami, Y. see M. Ohshima (225) 243
Kantor, C. see M. Patarroyo (210) 127
Kao, Y.S. see Y.X. Zhu (208) 253
Kaplan, S. see W.J. Jackson (215) 171
Kapral, M.K. see G.A. Woolley (224) 337
Kaptein, R. see J.A.W.H. Vermeulen (219) 426
Karasik, G.I. see N.M. Matveeva (217) 42
Kärgel, H.-J., J. Stahl, B. Gross, S. Knespel, H. Bielka and M. Saarma, Studies on interaction of 5 S RNA with ribosomal proteins (220) 126
Kariya, K. and Y. Takai, Distinct functions of down-regulation-sensitive and -resistant types of protein kinase C in rabbit aortic smooth muscle cells (219) 119
Kariya, K. see E. Lee (209) 49
Kariya, K., Y. Fukumoto, T. Tsuda, Y. Kawahara, H. Fukuzaki, T. Yamamoto and Y. Takai, Inhibition of DNA synthesis by phorbol esters through protein kinase C in cultured rabbit aortic smooth muscle cells (217) 69
Kärki, T., E. Hakkola, I.E. Hassinen and J.K. Hiltunen, β-oxidation of polyunsaturated fatty acids in peroxisomes: Subcellular distribution of Δ^3,Δ^2-enoyl-CoA isomerase activity in rat liver (215) 228
Karlsson, H., I. Carlstedt and G.C. Hansson, Rapid characterization of mucin oligosaccharides from rat small intestine with gas chromatography-mass spectrometry (226) 23
Karlsson, I.-M. see Å. Strid (224) 348
Karplus, M. see G.M. Clore (213) 269
Kasinsky, H.F. see M. Chiva (215) 237
Kass, G.E.N. see G.A. Moore (224) 331
Kasuga, M. see K. Tobe (215) 345
Katada, T., M. Oinuma, K. Kusakabe and M. Ui, A new GTP-binding protein in brain tissues serving as the specific substrate of islet-activating protein, pertussis toxin (213) 353
Katial, A. see T. van Veen (208) 133
Kato, A. see M. Ohshima (225) 243
Kato, R. see N. Sasakawa (223) 413
Kato, T. see S. Ono (220) 332
Kato, T. see Y. Shimohigashi (222) 251
Katoh, S. see I. Enami (226) 161
Katsaros, D., G. Tortora, P. Tagliaferri, T. Clair, S. Ally, L. Neckers, R.K. Robins and Y.S. Cho-Chung, Site-selective cyclic AMP analogs provide a new approach in the control of cancer cell growth (223) 97
Katsura, N. see S. Oshiro (220) 383
Kattelman, E.J., S.K. Arora, C.T. Lim, D.L. Venton and G.C. Le Breton, A photoaffinity label for the thromboxane A_2/prostaglandin H_2 receptor in human blood platelets (213) 179
Katunuma, N. see H. Kido (223) 223
Katunuma, N. see K. Ishidoh (223) 69
Katunuma, N. see K. Ishidoh (226) 33
Katz, J., 'A quantitative analysis of the metabolic pathways of hepatic glucose synthesis in vivo with ^{13}C-labeled substrates' by B. Kalderon, A. Gopher and A. Lapidot [(1987) FEBS Lett. 213, 209–214] (220) 257
Katz, J.J. see T.J. Michalski (226) 72

Katz, L.A., A.P. Koretsky and R.S. Balaban, Respiratory control in the glucose perfused heart: A ^{31}P NMR and NADH fluorescence study (221) 270
Kaul, R., D. Saluja and R.C. Sachar, Phosphorylation of small subunit plays a crucial role in the regulation of RuBPCase in moss and spinach (209) 63
Kaulen, A.D. see L.A. Drachev (209) 316
Kaulen, A.D. see L.A. Drachev (226) 139
Kawaguchi, H. and H. Yasuda, Effects of platelet-activating factor on conversion of angiotensin I to II (221) 305
Kawahara, Y. see K. Kariya (217) 69
Kawai, S., T. Umezawa and T. Higuchi, *p*-Benzoquinone monoketals, novel degradation products of *β-O*-4 lignin model compounds by *Coriolus versicolor* and lignin peroxidase of *Phanerochaete chrysosporium* (210) 61
Kawakatsu, H. see H. Hayashi (223) 267
Kawamori, A., J. Satoh, T. Inui and K. Satoh, EPR study of charge equilibrium at low temperatures in the S_2 state of oxygenevolving photosystem II particles (217) 134
Kawamura, M. see S. Noguchi (225) 27
Kawasaki, H. see K. Ishidoh (223) 69
Kawasaki, H. see K. Ishidoh (226) 33
Kawasaki, H. see K. Suzuki (220) 271
Kawasaki, H. see S. Imajoh (215) 274
Kawasaki, K. see A. Tomoda (219) 472
Kawasaki, T. see A. Kurosaka (215) 137
Kawase, M. see R. Taguchi (225) 273
Kawata, M. see T. Tanimoto (226) 291
Kawata, T. see K. Mikami (223) 273
Kay, C.M. see I.K.M. Leung (214) 35
Kayano, T. see Y. Seino (223) 74
Kebabian, J.W. see T. Yamamoto (219) 326
Keen, J.N. see A. Cavaggioni (212) 225
Keenan, A.K., D. Cooney, A. Holzhöfer, C. Dees and M. Hekman, Unimpaired coupling of phosphorylated, desensitized *β*-adrenoceptor to G_s in a reconstitution system (217) 287
Keleti, T., Two rules of enzyme kinetics for reversible Michaelis-Menten mechanisms (208) 109
Keller, B.U. see M. Criado (224) 172
Kellie, S. and N.M. Wigglesworth, The cytoskeletal protein vinculin is acylated by myristic acid (213) 428
Kelly, K., B.N. Rospendowski, W.E. Smith and C.R. Wolf, Surface enhanced resonance Raman scattering as a probe of the spin state of structurally related cytochromes P-450 from rat liver (222) 120
Kelly, K.L., J.M. Mato and L. Jarett, The polar head group of a novel insulin-sensitive glycophospholipid mimics insulin action on phospholipid methyltransferase (209) 238
Kelly, R.P. see F.M. Ashcroft (215) 9
Kelly, R.P. see M. Kakei (208) 63
Kemp, B.E. see W. Kudlicki (215) 16
Kennedy, W.P.K. see P.G. Penketh (221) 427
Kerlavage, A.R. see F.-Z. Chung (211) 200
Keso, L., M. Lukka, C. Ehnholm, M. Baumann, P. Vihko and M. Olkinuora, Apolipoprotein A-I-binding protein from human term placenta: Purification and partial characterization (215) 105
Kesvatera, T. see V. Tõugu (225) 77
Keyeux, G. see S. Huck (208) 221
Khamis, M.I., J.R. Casas-Finet, A.H. Maki, J.B. Murphy and J.W. Chase, Role of tryptophan 54 in the binding of *E. coli* single-stranded DNA-binding protein to single-stranded polynucleotides (211) 155
Khamis, M.I. see J.R. Casas-Finet (220) 347
Khlebodarova, T.M. see N.M. Matveeva (217) 42
Khodjaev, E.Yu. see B.V. Chernyak (215) 300
Khoja, S.M., Effects of female sex hormones on the activity of serum hyaluronidase (226) 220
Kholodenko, B., V. Žilinskienė, V. Borutaitė, L. Ivanovienė, A. Toleikis and A. Praškevičius, The role of adenine nucleotide translocators in regulation of oxidative phosphorylation in heart mitochondria (223) 247
Khramtsov, N.V. see Yu.A. Ovchinnikov (223) 169
Kido, H., N. Fukusen and N. Katunuma, Epidermal growth factor as a new regulator of induction of tyrosine aminotransferase and tryptophan oxygenase by glucocorticoids (223) 223
Kiesel, L., G.L. Lukács, I. Eberhardt, B. Runnebaum and A. Spät, Effect of inositol 1,4,5-trisphosphate and GTP on calcium release from pituitary microsomes (217) 85
Kijatkin, N.I. see E.D. Sverdlov (217) 275
Kikkawa, U., K. Ogita, Y. Ono, Y. Asaoka, M.S. Shearman, T. Fujii, K. Ase, K. Sekiguchi, K. Igarashi and Y. Nishizuka, The common structure and activities of four subspecies of rat brain protein kinase C family (223) 212
Kikkawa, U. see M. Ido (219) 215
Kikkawa, U. see Y. Ono (226) 125
Kikkawa, U., Y. Ono, K. Ogita, T. Fujii, Y. Asaoka, K. Sekiguchi, Y. Kosaka, K. Igarashi and Y. Nishizuka, Identification of the structures of multiple subspecies of protein kinase C expressed in rat brain (217) 227
Kikuchi, S. see F. Takaiwa (221) 43
Kilburn, D.G. see M.L. Langsford (225) 163
Kiley, P.J. see W.J. Jackson (215) 171
Kim, J.M., S. Shimizu and H. Yamada, Evidence for the presence of a cytosine deaminase that does not catalyze the deimination of creatine (210) 77
Kim, T.W. see C. Yang (224) 261
Kimura, H. see Y. Shimohigashi (222) 71
Kimura, J. and M. Kimura, The complete amino acid sequences of the 5 S rRNA binding proteins L5 and L18 from the moderate thermophile *Bacillus stearothermophilus* ribosome (210) 85
Kimura, J., E. Arndt and M. Kimura, Primary structures of three highly acidic ribosomal proteins S6, S12 and S15 from the archaebacterium *Halobacterium marismortui* (224) 65
Kimura, J. see M. Kimura (210) 91
Kimura, M. and J. Kimura, The complete amino acid sequence of ribosomal protein S12 from *Bacillus stearothermophilus* (210) 91
Kimura, M. see J. Kimura (210) 85
Kimura, M. see J. Kimura (224) 65
Kimura, T. see Y. Hirata (219) 369
King, J.A. see C.E. Smith (225) 247
King, K.D. see W.B. Macklin (223) 417
Kinnally, K.W., H. Tedeschi and C.A. Mannella, Evidence for a novel voltage-activated channel in the outer mitochondrial membrane (226) 83
Kinoshita, J.H. see D. Carper (220) 209
Kinosian, H.J. see L.A. Selden (217) 89
Kirillova, M.A. see L.P. Sashchenko (226) 261
Kirk, K.L. see D. Ukena (209) 122
Kirk, K.L. see K.A. Jacobson (225) 97
Kirkness, E.F. see K.B. Widdows (222) 125
Kirschke, H. see D. Brömme (219) 441
Kishimoto, T. see T. Andus (221) 18
Kiss, Z., E. Deli and J.F. Kuo, Cyclic AMP-like effects of polyamines on phosphatidylcholine synthesis and protein phosphorylation in human promyelocytic leukemia HL60 cells: Comparison with the effects of phorbol ester (213) 365Kitamura, S., T. Shimizu, T. Izumi and Y. Seyama, Synthesis of 11,12-leukotriene A_4,11*S*,12*S* - oxido - 5*Z*,7*E*,9*E*,14*Z* - eicosatetra-enoic acid, a novel leukotriene of the 12-lipoxygenase pathway (213) 169
Kiss, Z. see E. Deli (221) 365
Kitabgi, P. see J.-C. Bozou (211) 151
Kitada, K. see S. Oshiro (220) 383

Kitagawa, H. see A. Kurosaka (215) 137
Kitagawa, H. see A. Yamaguchi (225) 228
Kitagawa, T. see S. Hashimoto (208) 305
Kitagawa, Y. see Y. Aratani (218) 47
Kitasato, H. see K. Tobe (215) 345
Kiyatkin, N.I. see Yu.A. Ovchinnikov (213) 73
Kizer, D.E. see D.P. Ringer (224) 59
Klærke, D.A., J. Petersen and P.L. Jørgensen, Purification of Ca^{2+}-activated K^+ channel protein on calmodulin affinity columns after detergent solubilization of luminal membranes from outer renal medulla (216) 211
Klee, A. see B. Tocqué (222) 327
Kleefeld, S. see H. Strotmann (221) 265
Klein, R. see R. Thiery (223) 381
Kleppe, K. see D.E. Helland (213) 215
Kliger, D.S. see K.R. Parker (211) 35
Klimov, V.V. see S.I. Allakhverdiev (226) 186
Klingenberg, M. see C. Eckerskorn (226) 166
Klingenberg, M. see H. Aquila (212) 1
Klinz, F.-J., V.C. Yu, W. Sadée and T. Costa, Differential expression of α-subunits of G-proteins in human neuroblastoma-derived cell clones (224) 43
Klip, A., T. Ramlal, D.A. Young and J.O. Holloszy, Insulin-induced translocation of glucose transporters in rat hindlimb muscles (224) 224
Knaff, D.B. see M. Hirasawa (221) 343
Knaus, H.-G. see J. Striessnig (212) 247
Knecht, E. see E. Roche (221) 231
Knecht, E. see J.L. Vargas (224) 182
Knecht, E. see R. Wallace (208) 427
Knespel, S. see H.-J. Kärgel (220) 126
Knibiehler, M. and C. Lazdunski, Conformation of colicin A: apparent difference between cytoplasmic and extracellular polypeptide chain (216) 183
Knight, D.E. and M.C. Scrutton, Secretion of 5-hydroxytryptamine from electropermeabilised human platelets: Effects of GTP and cyclic 3′,5′-AMP (223) 47
Knowler, J.T. see M.T. Travers (211) 27
Knox, P.P. see V.Z. Paschenko (214) 28
Ko, J.-L. see C.-L. Ho (209) 18
Kobayashi, M. see T. Hiyama (214) 97
Kobayashi, Y. see S. Koyama (209) 265
Kobelt, A., A. Pfaltz, D. Ankel-Fuchs and R.K. Thauer, The L-form of *N*-7-mercaptoheptanoyl-*O*-phosphothreonine is the enantiomer active as component B in methyl-CoM reduction to methane (214) 265
Kobelt, A. see J. Ellermann (220) 358
Kodama, C. see F. Inagaki (212) 91
Kodama, C. see F. Inagaki (219) 45
Kodama, H. see Y. Shimohigashi (222) 251
Kodera, Y. see S. Ono (220) 332
Koenderman, A.H.L., P.W. Wijermans and D.H. van den Eijnden, Changes in the expression of *N*-acetylglucosaminyltransferase III, IV, V associated with the differentiation of HL-60 cells (222) 42
Koga, T. see Y. Uejima (214) 127
Koh, G. see Y. Seino (223) 74
Kohchi, T. see H. Fukuzawa (220) 61
Kohda, D. see F. Inagaki (212) 91
Köhler, K. see M. Thaler (219) 351
Kohring, G.-W. and F. Mayer, In situ distribution of *Eco*RI methylase and restriction endonuclease in cells of *Escherichia coli Bs 5* (216) 207
Koide, T. and S. Odani, Histidine-rich glycoprotein is evolutionarily related to the cystatin superfamily: Presence of two cystatin domains in the N-terminal region (216) 17
Koide, T. see R. Yamagishi (225) 109
Koivu, J., Disulfide bonding as a determinant of the molecular composition of types I, II and III procollagen (217) 216
Koivu, J., Identification of disulfide bonds in carboxy-terminal propeptides of human type I procollagen (212) 229
Koizumi, M. see Y. Shimonishi (215) 165
Kojro, E., I. Willhardt, A. Römbach, Z. Grzonka and P. Hermann, Tetrazole analogs of amino acids as constituents of modifiers of carboxypeptidase A catalysis (212) 83
Kolchanov, N.A., V.V. Solovyov and I.B. Rogozin, Pecularities of immunoglobulin gene structures as a basis for somatic mutation emergence (214) 87
Kolesnikov, S.S., A.B. Jainazarov and E.E. Fesenko, Time-dependent cGMP-activated conductance of detached patches of ROS plasma membrane (222) 37
Koll, R. see S. Fehr (210) 45
Kolsteeg, C.E.M. see J.A. van Kuik (221) 150
Komano, O. see B. Cantournet (220) 143
Kominami, E. see K. Ishidoh (223) 69
Kominami, E. see K. Ishidoh (226) 33
Kondo, M. see Y. Shimohigashi (222) 251
Konings, F., Comment on the letter of Drs. De Block and De Potter (222) 359
Konings, W.N. see M. Opekarovà (213) 45
Konings, W.N. see W. Crielaard (225) 6
Konishi, T. and N. Murakami, $\Delta\psi$-dependent gating of Na^+/H^+ exchange in *Halobacterium halobium:* a $\Delta\tilde{\mu}H^+$-driven Na^+ pump (226) 270
Konno, T. see T. Kurosaki (214) 253
Konno, T. see T. Tobimatsu (222) 56
Kono, N. see H. Nakajima (223) 113
Kononenko, A.A. see V.Z. Paschenko (214) 28
Konopka, A. see K.P. Samuel (218) 81
Konstantinov, A.A. see T. Vygodina (219) 387
Kopchick, J.J. see R.C. Gorewit (225) 238
Kopecký, P. see L. Morávek (208) 435
Koprowski, H. see G. Larson (214) 41
Koprowski, H. see J. Thurin (208) 17
Koretsky, A.P. see L.A. Katz (221) 270
Koretz, J.F. see R.C. Augusteyn (222) 1
Korn, E.D. see M. Coué (213) 316
Kornblihtt, A. see M. Obara (213) 261
Korneev, S.A. see S.A. Limborska (212) 208
Kornilova, E.S. see L.V. Teslenko (221) 105
Korobkova, E.N. see G.I. Muzya (220) 371
Korth, R. see J. Benveniste (226) 371
Korvatovsky, B.N. see V.Z. Paschenko (214) 28
Kosaka, Y. see U. Kikkawa (217) 227
Koschel, K. see H.G. Kress (221) 28
Koster, J.F. see A. Montfoort (226) 101
Kostič, M.M., D. Maretzki, R. Živković, E.G. Krause and S.M. Rapoport, Activation and desensitization of glycolysis by stimulation of adenylate cyclase in rat reticulocytes (217) 163
Kostina, M.B. see E.D. Sverdlov (217) 275
Kostina, M.B. see Yu.A. Ovchinnikov (213) 73
Kostyrko, V.A. see I.A. Smirnova (214) 343
Koteliansky, V.E. see A.M. Belkin (220) 291
Koteliansky, V.E. see M.A. Chernousov (217) 124
Koteliansky, V.E. see M.A. Glukhova (218) 292
Kothekar, V. see B. Chandrasekhar (225) 151
Kothekar, V. see K. Gopalakrishna (215) 95
Koyama, J. see M. Sato (224) 29
Koyama, S., H. Daiyasu, S. Hase, Y. Kobayashi, Y. Kyogoku and T. Ikenaka, ^{1}H-NMR analysis of the sugar structures of glycoproteins as their pyridylamino derivatives (209) 265
Kraayenhof, R. see H.S. van Walraven (208) 138
Krab, K. see H.S. van Walraven (208) 138
Krahmer, M. and R. Prohaska, Characterization of human red cell Rh (Rhesus-)specific polypeptides by limited proteolysis (226) 105
Kramer, G. see W. Kudlicki (215) 16
Kramer, J.H. see C.M. Arroyo (221) 101

Kramer, M.D. see H.-G. Simon (223) 352
Krasnovsky, A.A. and V.V. Nikandrov, The photobiocatalytic system: inorganic semiconductors coupled to bacterial cells (219) 93
Krause, E.G. see M.M. Kostić (217) 163
Krayevsky, A. see N. Dyatkina (219) 151
Kreisel, M. see R.B. Frydman (219) 380
Krenn, B.E. see R. Wever (216) 1
Kress, H.G., H. Eckhardt-Wallasch, P.W.L. Tas and K. Koschel, Volatile anesthetics depress the depolarization-induced cytoplasmic calcium rise in PC 12 cells (221) 28
Kretzmer, K.K. see T.-C. Wun (210) 11
Kriebardis, A. and W. Guschlbauer, *dam* methylase from *E. coli*: Circular dichroism investigations of the secondary structure and influence of *S*-adenosylmethionine (213) 297
Kriebardis, A. see R. Lauster (220) 167
Krikun, G. and A.I. Cederbaum, Effect of chronic ethanol consumption on microsomal lipid peroxidation: Role of iron and comparison between controls (208) 292
Krishnan, L.K. and M.P. Jamaluddin, Conformational responses of an arachidonate- and U46619-binding haemoprotein in relation to platelet activation (212) 213
Krivokapich, J. see S.C. Huang (216) 128
Krizek, J., L.M. Coluccio and A. Bretscher, ATPase activity of the microvillar 110 kDa polypeptide-calmodulin complex is activated in Mg^{2+} and inhibited in K^+-EDTA by F-actin (225) 269
Kröger, B. see R.K. Hartmann (218) 215
Krömer, S., M. Stitt and H.W. Heldt, Mitochondrial oxidative phosphorylation participating in photosynthetic metabolism of a leaf cell (226) 352
Krstenansky, J.L. and S.J.T. Mao, Antithrombin properties of C-terminus of hirudin using synthetic unsulfated N^{α}-acetyl-hirudin$_{45-65}$ (211) 10
Krüger, H. see F. Hucho (211) 207
Krupp, G. and D. Söll, Simplified in vitro synthesis of mutated RNA molecules: An oligonucleotide promoter determines the initiation site of T_7 RNA polymerase on ss M13 phage DNA (212) 271
Kryceve-Martinerie, C., J. Soret, J. Crochet, M. Baluda and B. Perbal, Expression of a truncated *v-myb* product in transformed chicken embryo fibroblasts (214) 81
Krykbaev, R.A. see E.D. Sverdlov (212) 233
Krynetskaya, N.F. see V.G. Metelev (226) 232
Kubo, K., S. Ohno and K. Suzuki, Primary structures of human protein kinase C βI and βII differ only in their C-terminal sequences (223) 138
Kubo, S. see I. Matsuoka (216) 295
Kubo, T., A. Maeda, K. Sugimoto, I. Akiba, A. Mikami, H. Takahashi, T. Haga, K. Haga, A. Ichiyama, K. Kangawa, H. Matsuo, T. Hirose and S. Numa, Primary structure of porcine cardiac muscarinic acetylcholine receptor deduced from the cDNA sequence (209) 367
Kubota, Y., T. Shuin, M. Yao, H. Inoue and T. Yoshioka, The enhanced ^{32}P labeling of CDP-diacylglycerol in c-*myc* gene expressed human kidney cancer cells (212) 159
Kuchel, P.W., B.E. Chapman and J.R. Potts, Glucose transport in human erythrocytes measured using ^{13}C NMR spin transfer (219) 5
Kudlicki, W., R.E.H. Wettenhall, B.E. Kemp, R. Szyszka, G. Kramer and B. Hardesty, Evidence for a second phosphorylation site on eIF-2α from rabbit reticulocytes (215) 16
Kühlbrandt, W., A. Becker and W. Mäntele, Chlorophyll dichroism of three-dimensional crystals of the light-harvesting chlorophyll *a*/*b*-protein complex (226) 275
Kühn, H., R. Wiesner, L. Alder, T. Schewe and H. Stender, Formation of lipoxin B by the pure reticulocyte lipoxygenase (208) 248
Kühn, K. see U. Schwarz-Magdolen (208) 203
Kühnemund, O. see L. Morávek (208) 435
Kukhanova, M. see N. Dyatkina (219) 151
Kulikov, V.A. see S.V. Shlyapnikov (209) 335
Kulinski, T. see A.J.W.G. Visser (224) 406
Kumar, S., J. de Vellis, N.J. Lowe and D.P. Weingarten, all-*trans*-Retinoic acid inhibits the appearance of two phorbol ester-induced ornithine decarboxylase mRNAs in mouse epidermis (208) 151
Kumar, S. see G.J. Anderson (220) 323
Kun, E. see A. Hakam (212) 73
Kunze, H. see B.-M. Löffler (216) 51
Kuo, J.F. see E. Deli (221) 365
Kuo, J.F. see Z. Kiss (213) 365
Kupriyanov, V.V., N.V. Lyulina, A.Y. Steinschneider, M.Yu. Zueva and V.A. Saks, Creatine kinase-catalyzed ATP-phosphocreatine exchange: Comparison of ^{31}P-NMR saturation transfer technique and radioisotope tracer methods (208) 89
Kurihara, K. see I. Matsuoka (216) 295
Kuriki, Y., Heat shock inactivates a supernatant factor(s) specifically required for efficient expression of the *amp* gene in *Escherichia coli* (223) 127
Kurkinen, M. see S.L. Hostikka (216) 281
Kuroda, Y., H. Nakayama, T. Ishibashi, S. Aoki, S. Tushima and S. Nakagawa, A significant increase of lysophosphatidylinositol 4-phosphate with insulin in isolated rat fat cells (224) 137
Kurokawa, T., R. Sasada, M. Iwane and K. Igarashi, Cloning and expression of cDNA encoding human basic fibroblast growth factor (213) 189
Kuroki, T. see M. Itami (222) 289
Kurono, M. see Y. Seino (223) 74
Kurosaka, A., S. Fukui, H. Kitagawa, H. Nakada, Y. Numata, I. Funakoshi, T. Kawasaki and I. Yamashina, Mucin-carbohydrate directed monoclonal antibody (215) 137
Kurosaki, T., K. Fukuda, T. Konno, Y. Mori, K.-I. Tanaka, M. Mishina and S. Numa, Functional properties of nicotinic acetylcholine receptor subunits expressed in various combinations (214) 253
Kurosawa, Y. see Y. Kuwana (219) 360
Kurose, H., T. Inagami and M. Ui, Participation of adenosine 5′-triphosphate in the activation of membrane-bound guanylate cyclase by the atrial natriuretic factor (219) 375
Kuryatov, A.B. see A.S. Arseniev (213) 283
Kusakabe, K. see T. Katada (213) 353
Kushnirov, V.V., M.D. Ter-Avanesyan, A.P. Surguchov, V.N. Smirnov and S.G. Inge-Vechtomov, Localization of possible functional domains in *sup*2 gene product of the yeast *Saccharomyces cerevisiae* (215) 257
Kutejová, E. see L. Horniak (224) 283
Kutyshenko, V.P. see G.V. Semisotnov (224) 9
Kuwajima, K., H. Yamaya, S. Miwa, S. Sugai and T. Nagamura, Rapid formation of secondary structure framework in protein folding studied by stopped-flow circular dichroism (221) 115
Kuwana, Y., S. Itoh, F. Nagase, I. Nakashima and Y. Kurosawa, Production of the constant domain of murine T-cell receptor β-chain in *Escherichia coli* (219) 360
Kuwano, R. see T. Yamakuni (223) 117
Kuznetsov, A.M., V.A. Bogdanovskaya, M.R. Tarasevich and E.F. Gavrilova, The mechanism of cathode reduction of oxygen in a carbon carrier-laccase system (215) 219
Kuznetsov, S.A. and V.I. Gelfand, 18 kDa microtubule-associated protein: identification as a new light chain (LC-3) of microtubule-associated protein 1 (MAP-1) (212) 145

Kwan, A.P.L. see S. Ayad (220) 181
Kyogoku, Y. see S. Koyama (209) 265
Kyogoku, Y. see S. Nishimoto (213) 293

L

La Mantia, G. see A. Pascucci (226) 297
La Mantia, G. see L. Lania (219) 400
Labouesse, J. see A.M. Garrigues (224) 267
Labouesse, J. see H. Valeins (226) 331
Labourdette, G. see C. Gensburger (217) 1
Laburthe, M. see J.-C. Bozou (211) 151
Laduron, P.M. see W. Wouters (213) 359
Lafaye, P. and C. Lapresle, Location of penicilloyl groups on CNBr fragments of the albumin from penicillin-treated patients (220) 206
Laffi, G., F. Cominelli, M. Ruggiero, S. Fedi, V. Chiarugi and P. Gentilini, Molecular mechanism underlying impaired platelet responsiveness in liver cirrhosis (220) 217
Lagenfelt, G. see R. Aasa (221) 245
Lahlou, B. see M.E. Guibbolini (220) 98
Laidman, D.L. see B.N. Zaba (213) 49
Laine, R. see P. Leinonen (215) 68
Laitinen, J., R. Löppönen, J. Merenmies and H. Rauvala, Binding of laminin to brain gangliosides and inhibition of lamininneuron interaction by the gangliosides (217) 94
Lakey, J.H., Voltage gating in porin channels (211) 1
Lala, A.K. see R.M. Mogre (221) 408
Lambeth, J.D. see E. Deli (221) 365
Lambolez, B. and J. Rossier, Benzodiazepine agonists protect a histidine residue from modification by diethyl pyrocarbonate whereas propyl β-carboline does not (219) 301
Lamerichs, R.M.J.N. see J.A.W.H. Vermeulen (219) 426
Lammi, M., C.L. Pon and C.O. Gualerzi, The NH_2-terminal cleavage of *Escherichia coli* translational initiation factor IF3: A mechanism to control the intracellular level of the factor? (215) 115
Lampen, J.O. see N.J. Nicholls (221) 179
Lamponi, S., C. Galassi, P. Tortora and A. Guerritore, Glucose-induced degradation of yeast fructose-1,6-bisphosphatase requires additional triggering events besides protein phosphorylation (216) 265
Landau, I. see A. Brossi (214) 291
Landau, I. see A. Brossi (223) 77
Landemore, G., S.-E. Letaïef, J. Bocquet and J. Izard, Kurloff cell proteoglycans: Evidence for de novo synthesis of chondroitin sulphate proteoglycans by purified Kurloff cells (209) 299
Lang, B.F. see R. Cedergren (226) 63
Langley, O.K., M.C. Aletsee and M. Gratzl, Endocrine cells share expression of N-CAM with neurones (220) 108
Langsford, M.L., N.R. Gilkes, B. Singh, B. Moser, R.C. Miller, jr, R.A.J. Warren and D.G. Kilburn, Glycosylation of bacterial cellulases prevents proteolytic cleavage between functional domains (225) 163
Lania, L., A. Pannuti, G.La Mantia and C. Basilico, The transcription of B2 repeated sequences is regulated during the transition from quiescent to proliferative state in cultured rodent cells (219) 400
Lania, L. see A. Pascucci (226) 297
Lanzara, V. and E. Grazi, On an Mg^{2+}-dependent interaction of actin with glyceraldehyde-phosphate dehydrogenase: The fundamental role of KCl in the organization of F-actin (221) 387
Lapidot, A. see B. Kalderon (213) 209
Lapidot, A. see B. Kalderon (220) 259
Lapresle, C. see P. Lafaye (220) 206
Lardy, H.A. see L.L. Ji (208) 297
Larkins, R.G. see M. Dunlop (220) 84
Larson, G., P. Watsfeldt, P. Falk, H. Leffler and H. Koprowski, Fecal excretion of intestinal glycosphingolipids by newborns and young children (214) 41
Larsson, K. see M. Forsgren (213) 254
Larsson, R. see P. Nygren (213) 195
Larsson, U.K. see M. Spangfort (224) 343
Lasch, J., G. Niedermann, A.A. Bogdanov and V.P. Torchilin, Thiolation of preformed liposomes with iminothiolane (214) 13
Lasmoles, F. see S. Minvielle (223) 63
Laurent, A., M. Basset, M. Dorée and C.J. Le Peuch, Involvement of a calcium-phospholipid-dependent protein kinase in the maturation of *Xenopus laevis* oocytes (226) 324
Laurent, E. see I. Graff (210) 204
Lauster, R., A. Kriebardis and W. Guschlbauer, The GATATC-modification enzyme *Eco*RV is closely related to the GATC-recognizing methyltransferases *Dpn*II and *dam* from *E. coli* and phage T_4 (220) 167
Lautenberger, J.A. see K.P. Samuel (218) 81
Laverdure, G.R., D. Banerjee, I. Chackalaparampil and B.B. Mukherjee, Epidermal and transforming growth factors modulate secretion of a 69 kDa phosphoprotein in normal rat kidney fibroblasts (222) 261
Lavergne, J.-P., F. Conquet, J.-P. Reboud and A.-M. Reboud, Role of acidic phosphoproteins in the partial reconstitution of the active 60 S ribosomal subunit (216) 83
Lavrik, O.I., A.S. Levina, G.A. Nevinsky and V.N. Podust, Role of nucleoside components and internucleotide phosphate groups of oligodeoxyribonucleotide template in its binding to human DNA polymerase α (216) 225
Lavrik, O.I. see S.V. Doronin (216) 221
Law, S.W. see S.S. Fojo (213) 221
Lawrence, M. see R.J.P. Williams (209) 111
Laychock, S.G. and S. Bilgin, α_2-Adrenergic inhibition of pancreatic islet glucose utilization is mediated by an inhibitory guanine nucleotide regulatory protein (218) 7
Lazard, M., M. Mirande and J.-P. Waller, Overexpression of mammalian phenylalanyl-tRNA synthetase upon phenylalanine restriction (216) 27
Lazdunski, C. see J.-M. Bolla (224) 213
Lazdunski, C. see M. Knibiehler (216) 183
Lazdunski, M. see A. Lombet (219) 355
Lazer, G. see M.K. Agarwal (217) 221
Lazure, C. see N.G. Seidah (211) 144
Lazure, C. see S. Benjannet (224) 142
Le Bouc, Y., P. Noguiez, P. Sondermeijer, D. Dreyer, F. Girard and M. Binoux, A new 5'-non-coding region for human placental insulin-like growth factor II mRNA expression (222) 181
Le Breton, G.C. see E.J. Kattelman (213) 179
Le Call, J. see M.-C. Liu (218) 227
Le Cam, A. see G. Le Cam (210) 1
Le Cam, G. and A. Le Cam, Synthesis of the growth hormone-regulated rat liver anti-protease GHR-P^{63} is inhibited by acute inflammation (210) 1
Le Maire, M. see A. Viel (223) 232
Le Peuch, C.J. see A. Laurent (226) 324
Leadlay, P.F. see R.S. Hale (224) 133
Lebedeva, N.N. see Yu.S. Borovikov (223) 409
Leber, B. see J.D. Norton (215) 127

Leblond-Francillard, M. see M. Ekker (222) 337
LeBoff, M. see E. Brown (218) 113
LeBoff, M. see M. Oetting (208) 99
Lecomte, P.J. and J. Ninio, Variations with position of replication errors due to exonuclease warm-up (221) 194
Leduc, R. see S. Benjannet (224) 142
Lee, E. and K. Kariya, Propylthiouracil, a selective inhibitor of NADH-cytochrome b_5 reductase (209) 49
Lee, J.S. see B.L. Haug (215) 252
Lee, J.-Y. see B.-D. Hsu (217) 53
Lee, K.M. and J.-F. Biellmann, Enzyme and organic solvents: horse liver alcohol dehydrogenase in non-ionic microemulsion: Stability and activity (223) 33
Lee, N.I. see L.L. Muldrow (213) 249
Lee, P. see K. Oxborough (221) 211
Lee, S. see S. Ono (220) 332
Lee, Y.C., J.A. Ball, D. Reece and S.R. Bloom, Neuromedin N: presence and chromatographic characterization in the rat (220) 243
Leemans, J. see H. Höfte (226) 364
Lefèvre, J.F. see E. Bertrand-Burggraf (215) 83
Leffak, M. see P.S. Fink (214) 75
Leffak, M. see P.S. Fink (220) 263
Leffler, H. see G. Larson (214) 41
Lefranc, G. see M.-P. Lefranc (213) 231
Lefranc, G. see S. Huck (208) 221
Lefranc, M.-P. and G. Lefranc, Human immunoglobulin heavy-chain multigene deletions in healthy individuals (213) 231
Lefranc, M.-P. see S. Huck (208) 221
Lefranc, M.-P. see S. Huck (224) 291
LeGall, J. see A.J.W.G. Visser (224) 406
LeGall, J. see E. Samain (216) 140
LeGall, J. see G.D. Fauque (215) 63
LeGall, J. see L.C. Sieker (208) 73
LeGall, J. see L.C. Sieker (209) 261
LeGall, J. see Y.M. Berlier (221) 241
Lehto, V.P. see V.M. Wasenius (221) 73
Leighton, B., R. Curi, A. Hussein and E.A. Newsholme, Maximum activities of some key enzymes of glycolysis, glutaminolysis, Krebs cycle and fatty acid utilization in bovine pulmonary endothelial cells (225) 93
Leinonen, P., L. Alhonen-Hongisto, R. Laine, O.A. Jänne and J. Jänne, Chronic exposure to dexamethasone induces hypomethylation of ornithine decarboxylase genes in a human myeloma cell line (215) 68
Leisola, M.S.A. see S.D. Haemmerli (220) 149
Lekka, M., A.D. Tselepis and D. Tsoukatos, 1-*O*-Alkyl-2-acetyl-*sn*-glyceryl-3-phosphorylcholine (PAF) is a minor lipid component in *Tetrahymena pyriformis* cells (208) 52
Lelkes, P.I., J.E. Friedman, K. Rosenheck and A. Oplatka, Destabilization of actin filaments as a requirement for the secretion of catecholamines from permeabilized chromaffin cells (208) 357
LeMaster, D.M., Chiral β and random fractional deuteration for the determination of protein sidechain conformation by NMR (223) 191
Lenaz, G. see M. Rugolo (212) 313
Leneveu, E. and M. Simonneau, Scorpion venom inhibits selectively Ca^{2+}-activated K^+ channels in situ (209) 165
Leng, M. see B. Hartmann (225) 11
Lentes, K.-U. see F.-Z. Chung (211) 200
Lenzen, G., P. Masson, J.-M. Jacquemin and A. Danchin, A TY1 element is inserted in the CYR1 control region of *Saccharomyces cerevisiae* strain AB320 (219) 254
Lerch, K. see M. Huber (219) 335
Lespinat, P.A. see Y.M. Berlier (221) 241
Lessire, R. see J.-J. Bessoule (214) 158
Letaïef, S.E. see G. Landemore (209) 299
Leung, I.K.M., R.S. Mani and C.M. Kay, Fluorescence studies on the Ca^{2+} and Zn^{2+} binding properties of the α-subunit of bovine brain S-100a protein (214) 35
Leung, P.C.K. see T. Minegishi (214) 139
Leunissen-Bijvelt, J. see A.M. Batenburg (223) 148
Level, M. see J.-P. Tenu (220) 93
Leverve, X.M., L.H.P. Caro, P.J.A.M. Plomp and A.J. Meijer, Control of proteolysis in perifused rat hepatocytes (219) 455
Levin, D.B. and M.S. DuBow, Cloning and localization of the repressor gene (*c*) of the Mu-like transposable phage D108 (222) 199
Levina, A.S. see O.I. Lavrik (216) 225
Levina, N.B. see Yu.A. Ovchinnikov (226) 91
Levine, B.A. see J.S. Evans (208) 211
Levinger, L.F. and G.S. Nass, A unique nucleoprotein structure associated with the *Drosophila melanogaster* 18–28 S rDNA nontranscribed spacer (209) 340
Levitsky, D.I., L.A. Shuvalova and B.F. Poglazov, The effect of myosin light chain phosphorylation on the actin-stimulated ATPase activity of myosin minifilaments (221) 77
Levitzki, A., Regulation of adenylate cyclase by hormones and G-proteins (211) 113
Levy, H.R. see M.M. Bhadbhade (211) 243
Lewis, G.P. see L.A.J. O'Neill (212) 35
Lewis, J.A. and A. Bendicenti di Girolamo, Activation of metallothionein expression is potentiated by DNA sequences present in the herpes simplex virus thymidine kinase gene (217) 292
Lewis, J.W. see K.R. Parker (211) 35
Lewis, T.J. see B.N. Zaba (213) 49
Leymarie, P. see A. Benhaim (223) 321
Lezza, A.M.S. see P. Cantatore (213) 144
Lian, L.-Y. see C.M. Dobson (225) 183
Lidholm, D.-A., G.H. Gudmundsson, K.G. Xanthopoulos and H.G. Boman, Insect immunity: cDNA clones coding for the precursor forms of cecropins A and D, antibacterial proteins from *Hyalophora cecropia* (226) 8
Lietz, H. see J. Striessnig (212) 247
Liewerink, H. see P.D. Baas (218) 119
Lifchitz, A. see J. Berthou (218) 55
Lilja, H. see Å. Lundwall (214) 317
Liljas, A. see W.G. Garland (220) 43
Liljenström, H., Maintenance of accuracy during amino acid starvation (223) 1
Lim, C.T. see E.J. Kattelman (213) 179
Limborska, S.A., S.A. Korneev, N.E. Maleeva, P.A. Slominsky, A.G. Jincharadze, P.L. Ivanov and A.P. Ryskov, Cloning of Alu-containing cDNAs from human fibroblasts and identification of small Alu^+ $poly(A)^+$ RNAs in a variety of human normal and tumor cells (212) 208
Lin, T. and W.A. Bridger, Phosphorylation by cyclic AMP-dependent protein kinase does not affect the association of ATP citrate-lyase with isolated mitochondria (224) 322
Lin, W.-J. see B.-D. Hsu (217) 53
Lin, Y. see J.J. Huang (223) 294
Lin-Chao, S. see C.C.-K. Chao (225) 133
Lindau, M. and O. Nüße, Pertussis toxin does not affect the time course of exocytosis in mast cells stimulated by intracellular application of GTP-γ-S (222) 317
Lindau, M. see J.M. Fernandez (216) 89
Lindgren, J.Å. see T. Miyamoto (216) 123
Lindh, E. see P. Nygren (213) 195
Lindley, I. see E. Pöschl (226) 96
Lindorfer, M.A. see S. Takamiya (218) 277
Lindstrom, J. see P. Whiting (213) 55
Lindstrom, J. see P. Whiting (219) 459
Lindstrom, J. see R. Schoepfer (226) 235

Lindy, S., T. Sorsa, K. Suomalainen and H. Turto, Gold sodium thiomalate activates latent human leukocyte collagenase (208) 23
Ling, C. see E. Bertrand-Burggraf (215) 83
Lingens, F. see W. Wiesner (209) 321
Lingley, M.D. see M.E. Houston (219) 469
Link, T.A. see H. Aquila (212) 1
Link, T.A. see H. Schägger (219) 161
Linnemann, D. see E. Bock (225) 33
Lipkin, V.M. see Yu.A. Ovchinnikov (223) 169
Lipkin, V.M. see Yu.A. Ovchinnikov (226) 91
Lips, C.J.M. see J.W.M. Höppener (215) 122
Lips, C.J.M. see P.H. Steenbergh (209) 97
Litchfield, D.W. see G.A. Cates (218) 195
Littlechild, J.A., G.J. Davies, S.J. Gamblin and H.C. Watson, Phosphoglycerate kinase from the extreme thermophile *Thermus thermophilus*: Crystallization and preliminary X-ray data (225) 123
Liu, M.-C., M.-Y. Liu, W.J. Payne, H.D. Peck jr, J. Le Gall and D.V. DerVartanian, Comparative EPR studies on the nitrite reductases from *Escherichia coli* and *Wolinella succinogenes* (218) 227
Liu, M.-Y. see M.-C. Liu (218) 227
Ljunghall, S. see P. Nygren (213) 195
Llinás, M. see J.A.W.H. Vermeulen (219) 426
Lloyd, C.J. see R.J. Simpson (224) 128
Lo, C.-H. see S.-H. Chiou (221) 134
Lo, T.-B. see S.-H. Chiou (209) 107
Lo, W.W.Y. and J. Hughes, A novel cholera toxin-sensitive G-protein (G_c) regulating receptor-mediated phosphoinositide signalling in human pituitary clonal cells (220) 327
Lo, W.W.Y. and J. Hughes, Pertussis toxin distinguishes between muscarinic receptor-mediated inhibition of adenylate cyclase and stimulation of phosphoinositide hydrolysis in Flow 9000 cells (220) 155
Lo, W.W.Y. and J. Hughes, Receptor-phosphoinositidase C coupling: Multiple G-proteins? (224) 1
Lo, W.W.Y. and J. Hughes, Specific and nonspecific effects of nucleotides on hormone-induced phosphoinositide turnover in permeabilized human pituitary tumour cells (Flow 9000) (226) 67
Lo Presti, E. see C. Delfini (210) 17
LoBrutto, R. see J. Telser (214) 117
Lockard, R.E., Identification of the proteins in direct contact with duck globin mRNA (219) 410
Lockau, W. see K. Ziegler (217) 16
Lockau, W. see W. Nitschke (218) 283
Lockheart, A. see P.W. Piper (214) 143
Lockshin, R.A. see T.W. Simmons (218) 251
Loeb, J. see B. Cantournet (220) 143
Löffelhardt, W. see J. Berenguer (224) 401
Löffler, B.-M. and H. Kunze, Fractionation, biochemical characterization and lysosomal phospholipases of human liver (216) 51
Logunov, S.L. see V.Z. Paschenko (214) 28
Lohse, D. see H. Strotmann (221) 265
Lohse, P. see J.R. Havinga (216) 275
Loidl, P. see G. Golderer (222) 322
Lombet, A., J.-N. Bidard and M. Lazdunski, Ciguatoxin and brevetoxins share a common receptor site on the neuronal voltage-dependent Na^+ channel (219) 355
Lomri, A., P.J. Marie, M. Escurat and M.-M. Portier, Cytoskeletal protein synthesis and organization in cultured mouse osteoblastic cells: Effects of cell density (222) 311
Londesborough, J. and M. Nuutinen, Ca^{2+}/calmodulin-dependent protein kinase in *Saccharomyces cerevisiae* (219) 249
Long, K. see N.J. Philp (225) 127
Löppönen, R. see J. Laitinen (217) 94
Lord, J.M. see M. O'Hare (216) 73
Lorenzo, J.L., M. Allorio, F. Bernini, A. Corsini and R. Fumagalli, Regulation of low density lipoprotein metabolism by 26-hydroxycholesterol in human fibroblasts (218) 77
Lottspeich, F. see R. Grimm (225) 215
Lottspeich, F. see R. Schlipfenbacher (209) 57
Louie, K.A. see W.G. Garland (220) 43
Lowe, N.J. see S. Kumar (208) 151
Loyter, A. see O. Nussbaum (221) 61
Lucantoni, A. and G.M. Polya, Activation of wheat embryo calcium-regulated protein kinase by unsaturated fatty acids in the presence and absence of calcium (221) 33
Lucas, C.A. see C.J. Rhodes (215) 179
Lucas, M. see C. Osuna (211) 41
Lücken, U. see E.P. Gogol (219) 274
Luduena, R.F. see A. Banerjee (219) 103
Lührmann, R. see P. Bringmann (213) 309
Lukács, G.L. see L. Kiesel (217) 85
Lukanidin, E.M. see L.P. Sashchenko (226) 261
Lukasheva, L.I. see S.Yu. Morozov (213) 438
Lukjanova, T.I. see L.P. Sashchenko (226) 261
Lukka, M. see L. Keso (215) 105
Lund, T., R. Bravo, H.R. Johansen, J. Zeuthen and J. Vuust, Synthesis, processing, and secretion of rat immunoglobulin E made in *Xenopus* oocytes (208) 369
Lundell, D.J. see F. Suter (217) 279
Lundwall, Å. and H. Lilja, Molecular cloning of human prostate specific antigen cDNA (214) 317
Lundwall, Å. see M. Abrahamson (216) 229
Lundwall, Å. see S. Gershagen (220) 129
Lunt, G.G. see C. Rapier (212) 292
Lunt, G.G. see D.R.E. Macallan (226) 357
Lunt, G. see Y. Aracava (222) 63
Lupker, J. see P. Ferrara (226) 47
Luther, M. see R. Schoepfer (226) 235
Lutsenko, S.V. see Yu.A. Ovchinnikov (217) 111
Luu, B. see J. Borg (213) 406
Luzzato, L. see E.A. Usanga (209) 23
Lyakhovich, V.V. see Ya.Yu. Woldman (212) 53
Lyles, J.M. see E. Bock (225) 33
Lyulina, N.V. see V.V. Kupriyanov (208) 89

M

Maassen, J.A. see G.D.F. Maessen (208) 77
Macallan, D.R.E., G.G. Lunt, S. Wonnacott, K.L. Swanson, H. Rapoport and E.X. Albuquerque, Methyllycaconitine and (+)-anatoxin-a differentiate between nicotinic receptors in vertebrate and invertebrate nervous systems (226) 357
MacDonald, H.R., P. Wingfield, U. Schmeissner, A. Shaw, G.M. Clore and A.M. Gronenborn, Point mutations of human interleukin-1 with decreased receptor binding affinity (209) 295
MacDonald, H.R. see C. Bron (219) 365
MacDonald, H.R. see P. Wingfield (215) 160
Machado, A.J. see L.E. Mallea (218) 143
Machleidt, W. see H. Schägger (219) 161
MacIntyre, D.E. see M. Bushfield (222) 299

Macklin, W.B., M.V. Gardinier, K.D. King and K. Kampf, An AG→GG transition at a splice site in the myelin proteolipid protein gene in jimpy mice results in the removal of an exon (223) 417
MacLennan, P.A., R.A. Brown and M.J. Rennie, A positive relationship between protein synthetic rate and intracellular glutamine concentration in perfused rat skeletal muscle (215) 187
Macleod, A.F. see J.D. Bell (219) 239
MacNicol, A.M., G.R. Banks and R.A. Cox, Biosynthesis and activity of DNA polymerase throughout the mitotic cycle of *Physarum polycephalum* (221) 48
Macrì, F. and A. Vianello, ADP- or pyrophosphate-dependent proton pumping of pea stem tonoplast-enriched vesicles (215) 47
Maddix, S. see K.H. Cheeseman (209) 191
Madsen, P. see J.E. Celis (209) 277
Madsen, P. see J.E. Celis (220) 1
Maeda, A. see T. Kubo (209) 367
Maeda, M. see S.-Y. Hsu (218) 222
Maeda, M. see T. Noumi (213) 381
Maekawa, S. and H. Sakai, A novel 60-kDa smooth muscle protein that binds filamin-actin filament complex (221) 68
Maeno, T. see K. Enomoto (223) 82
Mäenpää, P., B. Andersson and C. Sundby, Difference in sensitivity to photoinhibition between photosystem II in the appressed and non-appressed thylakoid regions (215) 31
Maeshima, M., A. Takeuchi and T. Asahi, Cell-free synthesis of alkaline lipase, a glyoxysomal membrane protein, from castor bean endosperm (220) 23
Maessen, G.D.F., R. Amons, J.A. Maassen and W. Möller, Primary structure of elongation factor 1β from *Artemia* (208) 77
Maessen, G.D.F., R. Amons, J.P. Zeelen and W. Möller, Primary structure of elongation factor 1γ from *Artemia* (223) 181
Maffey, L. see M.E. Schininà (221) 87
Magnaldo, I., H. Talwar, W.B. Anderson and J. Pouysségur, Evidence for a GTP-binding protein coupling thrombin receptor to PIP_2-phospholipase C in membranes of hamster fibroblasts (210) 6
Magnusson, R.P., J. Gestautas, P. Seto, A. Taurog and B. Rapoport, Isolation and characterization of a cDNA clone for porcine thyroid peroxidase (208) 391
Maity, S.N. and B. Bhattacharyya, Properties of B-ring analogues of colchicine (218) 102
Major, G.N. see P.R. Tempest (209) 357
Makarov, V.L., I. Smirnov and S.I. Dimitrov, Higher order folding of chromatin is induced in different ways by monovalent and by bivalent cations (212) 263
Maki, A.H. see J.R. Casas-Finet (220) 347
Maki, A.H. see M.I. Khamis (211) 155
Maki, A.H. see S.-Y. Mao (211) 83
Maki, M., E. Takano, H. Mori, A. Sato, T. Murachi and M. Hatanaka, All four internally repetitive domains of pig calpastatin possess inhibitory activities against calpains I and II (223) 174
Maki, M. see E. Takano (208) 199
Makita, A. see Y. Uehara (208) 352
Male, R. see D.E. Helland (213) 215
Malech, H.L. see P.M. Murphy (221) 81
Malech, H. see J. Falloon (209) 352
Maleeva, N.E. see S.A. Limborska (212) 208
Malgaroli, A. see A. Pandiella-Alonso (208) 48
Malkin, R., Interaction of stigmatellin and DNP-INT with the Rieske iron-sulfur center of the chloroplast cytochrome b_6-f complex (208) 317
Malkin, R., On the function of two vitamin K_1 molecules in the PS I electron acceptor complex (208) 343
Malkin, R. see Z. Adam (225) 67
Mallea, L.E., A.J. Machado, F. Navaroli and F.F.G. Rommerts, Modulation of stimulatory action of follicle stimulating hormone (FSH) and inhibitory action of epidermal growth factor (EGF) on aromatase activity in Sertoli cells by calcium (218) 143
Malloy, C.R., A.D. Sherry and F.M.H. Jeffrey, Carbon flux through citric acid cycle pathways in perfused heart by ^{13}C NMR spectroscopy (212) 58
Malmström, B.G. see M. Fabian (213) 396
Malmström, K. and H. Murer, Parathyroid hormone regulates phosphate transport in OK cells via an irreversible inactivation of a membrane protein (216) 257
Malo, C. and A. Berteloot, Proximo-distal gradient of Na^+-dependent D-glucose transport activity in the brush border membrane vesicles from the human fetal small intestine (220) 201
Malvoisin, E., F. Wild and G. Zwingelstein, 12-*O*-Tetradecanoyl phorbol 13-acetate stimulates the myristylation of an ~82 kDa protein in HL-60 cells (215) 175
Malvy, C. and J.R. Bertrand, Differential reactivity of 9-NH_2-ellipticine on apurinic and apyrimidinic sites in circular DNA (208) 155
Malyshev, I.V. see E.D. Sverdlov (217) 275
Malyshev, I.V. see Yu.A. Ovchinnikov (213) 73
Mamedov, M.D. see L.A. Drachev (213) 128
Manachini, P.L., C. Parini, M.G. Fortina and L. Benazzi, *Bli*I, a restriction endonuclease from *Bacillus licheniformis* (214) 305
Mancini, M. see G.J. Arlaud (222) 129
Mandl, J. see L. Buday (223) 15
Mangold, H.K. see N. Weber (211) 225
Mangold, H.K., T. Muramatsu, B.S. Roughley and N. Weber, Biologically active lipids: Semi-synthesis of ^3H-labeled ether glycerophospholipids and ether glyceroglycolipids from ratfish liver oil (220) 220
Mani, R.S. see I.K.M. Leung (214) 35
Mankin, A.S., E.A. Skripkin and V.K. Kagramanova, A putative internal promoter in the 16 S/23 S intergenic spacer of the rRNA operon of archaebacteria and eubacteria (219) 269
Mann, K., R. Deutzmann, M. Paulsson and R. Timpl, Solubilization of protein BM-40 from a basement membrane tumor with chelating agents and evidence for its identity with osteonectin and SPARC (218) 167
Mannella, C.A. see K.W. Kinnally (226) 83
Mansfield, R.W., J.A.M. Hubbard, J.H.A. Nugent and M.C.W. Evans, Extraction of electron acceptor A_1 from pea photosystem I (220) 74
Mäntele, W. see W. Kühlbrandt (226) 275
Mao, S.J.T. see J.L. Krstenansky (211) 10
Mao, S.-Y., A.H. Maki and G.H. de Haas, Comparative optically detected magnetic resonance studies of mammalian phospholipase A_2-lipid interactions (211) 83
Marchand, M.J. see M.I. Darville (224) 317
Marchese, E. see P. Ferrara (226) 47
Marcinkiewicz, M. see S. Benjannet (224) 142
Maretzki, D. see M.M. Kostić (217) 163
Margelin, D. see J. Gardette (225) 178
Margolis, L.B., I.A. Rozovskaja and V.P. Skulachev, Acidification of the interior of Ehrlich ascites tumor cells by nigericin inhibits DNA synthesis (220) 288
Marie, P.J. see A. Lomri (222) 311
Marinetti, T., Counterion collapse and the effect of diamines on bacteriorhodopsin (216) 155
Marinov, B.S. see N.V. Gulyaeva (211) 211
Markovits, A. see G. Premecz (226) 13
Marmé, D. see C. Bruns (212) 40
Marmé, D. see C. Bruns (221) 23

Marqués, S. see F.J. Florencio (223) 37
Marrec, N. see F. Heymans (218) 35
Marrero, H. and K.J. Rothschild, Bacteriorhodopsin's M_{412} and BR_{605} protein conformations are similar: Significance for proton transport (223) 289
Marsal, J. see X. Guitart (219) 219
Marsal, J. see X. Rabasseda (213) 337
Marschall, H. -U., B. Egestad, H. Matern, S. Matern and J. Sjövall, Evidence for bile acid glucosides as normal constituents in human urine (213) 411
Martin, B.M. see H.H. Valdivia (226) 280
Martin, B.M. see M.R. Ruff (211) 17
Martin, J. -L. see J. Breton (209) 37
Martin, T.J. see A.E. Fletcher (208) 263
Martín-Sanz, P., M. Cascales and L. Boscá, Fructose 2,6-bisphosphate in isolated foetal hepatocytes (225) 37
Marti, T., K. Takio, K.A. Walsh, G. Terzi and J.W. Truman, Microanalysis of the amino acid sequence of the eclosion hormone from the tobacco hornworm *Manduca sexta* (219) 415
Marti, T. see E. Takano (208) 199
Maruyama, I., H. Oyamada, T. Hasegawa, K. Ohtsuka and K. Momose, Effect of DFP on loading of fura 2/AM and quin 2/AM into single smooth muscle cells prepared from guinea pig taenia coli (220) 89
Maruyama, K. see M. Oosawa (213) 433
März, L. see F. Altmann (221) 145
Masliah, J., C. Kadiri, D. Pepin, T. Rybkine, J. Etienne, J. Chambaz and G. Bereziat, Antigenic relatedness between phospholipases A_2 from *Naja naja* venom and from mammalian cells (222) 11
Mason, R.P., J.K.M. Sanders and A. Cornish, In vivo enzymology: ^{13}C NMR measurement of a kinetic isotope effect for methanol oxidation in *Methylosinus trichosporium* OB3b (216) 4
Mason, R.P. see K.M. Morehouse (222) 246
Massare, M.J. and H.A. Blough, Inhibition of herpesvirus-induced thymidine kinase and DNA polymerase by β-hydroxynorvaline (223) 122
Masson, D., M. Zamai and J. Tschopp, Identification of granzyme A isolated from cytotoxic T-lymphocyte-granules as one of the proteases encoded by CTL-specific genes (208) 84
Masson, P. see G. Lenzen (219) 254
Matallana, E. see J.E. Pérez-Ortín (208) 31
Matern, H. see H. -U. Marschall (213) 411
Matern, S. see H. -U. Marschall (213) 411
Matfield, M. see W.J. Brammar (215) 291
Matheson, A.T. see W.G. Garland (220) 43
Mato, J.M. see I. Varela (211) 64
Mato, J.M. see K.L. Kelly (209) 238
Matozaki, T. see M. Nagao (214) 107
Matschinsky, F.M. see M. Prentki (220) 103
Matsubara, H. see H. Oh-oka (218) 52
Matsubara, H. see Y. Hirata (219) 369
Matsubara, K. see N. Tomita (225) 113
Matsuhashi, M. see M.D. Song (221) 167
Matsumiya, H. see Y. Tsunoda (222) 149
Matsumoto, S. see S. Ohno (222) 279
Matsumoto, T. see H. Kai (212) 119
Matsumoto, T. see H. Kai (221) 284
Matsuno, T. see E.F. Sato (214) 181
Matsuo, H. see T. Kubo (209) 367
Matsuoka, I., B. Syoto, K. Kurihara and S. Kubo, ADP-ribosylation of specific membrane proteins in pheochromocytoma and primary-cultured brain cells by botulinum neurotoxins type C and D (216) 295
Matsuoka, M. see M. Ohshima (225) 243
Matsusaka, H. see T. Shinozawa (219) 293
Matsuzawa, H. see H. Adachi (226) 150
Mattera, R. see W.N. Suki (220) 187
Matthew, J.B. see J.J. Huang (223) 294
Matthews, E.K. see G. Yonuschot (213) 401
Mattingly, R.R., M.L. Dreher and M.R. Hanley, Down-regulation of phorbol diester binding to NG115-401L neuronal cells is dependent on structure, concentration and time (223) 11
Matveetz, Ya.A. see S.V. Chekalin (216) 245
Matveeva, N.M., T.M. Khlebodarova, G.I. Karasik, N.B. Rubtsov, O.L. Serov, E.D. Sverdlov, N.E. Broude, N.N. Modyanov, G.S. Monastyrskaya and Yu.A. Ovchinnikov, Chromosomal localization of the gene coding for α-subunit of Na^+,K^+-ATPase in the American mink (*Mustela vison*) (217) 42
Matveyev, A.V. see M. Szekeres (222) 89
Mauk, A.G. see R.T. Hartshorn (213) 391
Mauk, M.R. see R.T. Hartshorn (213) 391
Maurer, A., A.T. Tu and P. Volpe, Crystallization of the Ca^{2+}-ATPase of skeletal muscle sarcoplasmic reticulum: Inhibition by myotoxin *a* (224) 89
Maurer, B. and R.M. Flügel, The 3′-orf protein of human immunodeficiency virus 2 shows sequence homology with the *bel*3 gene of the human spumaretrovirus (222) 286
Mauring, K., I. Renge and R. Avarmaa, A spectral hole-burning study of long-wavelength chlorophyll *a* forms in greening leaves at 5 K (223) 165
Mawer, E.B. see M.E. Hayes (220) 307
Mawhinney, T.P. see H.C. Riethman (215) 209
Mayer, F. see G.-W. Kohring (216) 207
Mayer, R.J. see D.G. Fernig (210) 165
Mayeux, P., C. Billat and R. Jacquot, Murine erythroleukaemia cells (Friend cells) possess high-affinity binding sites for erythropoietin (211) 229
Mayfield, M.B. see K. Miki (210) 199
Mayor, F. see M.C. Aragón (212) 87
Mayr, G.W. and W. Dietrich, The only inositol tetrakisphosphate detectable in avian erythrocytes is the isomer lacking phosphate at position 3: a NMR study (213) 278
Mazabraud, A. see A. Viel (223) 232
Mazat, J.-P. see P. Dabadie (226) 77
Mazière, C., S. Goldstein, M. Moreau, J.C. Mazière and J. Polonovski, Aspirin induces alterations in low-density lipoprotein and decreases its catabolism by cultured human fibroblasts (218) 243
Mazière, J.C. see C. Mazière (218) 243
Maziere, J.C. see J. Gardette (225) 178
M'Bina, I. see J. Olive (208) 308
Mbikay, M. see S. Benjannet (224) 142
McConkey, D.J. see G.A. Moore (224) 331
McCracken, J. see D.M. Dooley (214) 274
McDougall, J. and R.N. Nazar, Accessibility of phosphodiester bonds in the yeast ribosomal 5 S RNA protein complex (209) 52
McGee Tucker, M. see M.L. Ernst-Fonberg (215) 261
McGinn, M.T. see J.M. Mullin (221) 359
McGuirl, M.A. see D.M. Dooley (214) 274
McLaughlin, C.S. see M.F. Tuite (225) 205
McLeod, A.N. see H. Jhoti (219) 419
McLick, J. see A. Hakam (212) 73
McNaughton, G.A.L. see G.A. Nimmo (213) 18
Meggio, F., A.M. Brunati and L.A. Pinna, Polycation-dependent, Ca^{2+}-antagonized phosphorylation of calmodulin by casein kinase-2 and a spleen tyrosine protein kinase (215) 241
Meggio, F. see P. Agostinis (224) 385
Megyeri, K. see I. Rosztóczy (208) 56
Mehta, P.P. see G.M. Olins (224) 325

Meier, P.J. see J.C. Griffiths (213) 34
Melandri, B.A. see G. Venturoli (219) 477
Meldolesi, J. see A. Pandiella-Alonso (208) 48
Melin, P.-M., M. Sommarin, A.S. Sandelius and B. Jergil, Identification of Ca^{2+}-stimulated polyphosphoinositide phospholipase C in isolated plant plasma membranes (223) 87
Melis, A. see S. Demeter (214) 370
Melkov, A.M. see E.D. Sverdlov (217) 275
Melkov, A.M. see Yu.A. Ovchinnikov (213) 73
Ménez, A. see O. Trémeau (208) 236
Merati, G. and G. Zanetti, Chemical cross-linking of ferredoxin to spinach thylakoids: Evidence for two independent binding sites of ferredoxin to the membrane (215) 37
Mercer, E. see T.E. Cawston (209) 9
Merenmies, J. see J. Laitinen (217) 94
Merlevede, W. see J.R. Vandenheede (211) 190
Merlevede, W. see J.R. Vandenheede (216) 291
Merlevede, W. see P. Agostinis (224) 385
Mertens, E., E. Van Schaftingen and H.-G. Hers, Fructose 2,6-bisphosphate and the control of the energy charge in higher plants (221) 124
Meshi, T. see Y. Watanabe (219) 65
Mesland, D. see W. Homan (215) 323
Mészáros, G. see L. Buday (223) 15
Metcalfe, J.C. see G. Yonuschot (213) 401
Metelev, V.G., G.V. Zayakina, I.L. Ryabushenko, N.F. Krynetskaya, E.A. Romanova, T.S. Oretskaya and Z.A. Shabarova, Influence of probe structure on unique (regio-specific) cleavage of RNA by RNase H (226) 232
Metrione, R.M., H. Schweitz and K.A. Walsh, The amino acid sequence of toxin Rp_{III} from the sea anemone, *Radianthus paumotensis* (218) 59
Metters, K.M. see N.G. Seidah (211) 144
Meijer, A.J. see X.M. Leverve (219) 455
Meyer, M.S., R. Alon and S. Shibolet, Increased 25-hydroxyvitamin D levels in portal blood following cholecystokinin injection in the dog (212) 138
Michalak, K., J. Gutowicz and T. Modrzycka, Fluorescent probe studies on binding of glyceraldehyde-3-phosphate dehydrogenase to phosphatidylinositol liposomes: Further evidence for conformational changes (219) 233
Michalski, T.J., C. Bradshaw, J.E. Hunt, J.R. Norris and J.J. Katz, Triton X-100 reacts with chlorophyll in the presence of chlorophyllase (226) 72
Michel, H.P. and J. Bennett, Identification of the phosphorylation site of an 8.3 kDa protein from photosystem II of spinach (212) 103
Michel, H. see H.P. Braun (221) 221
Michel-Beyerle, M.E. see H.P. Braun (221) 221
Michelson, A.M. see M.E. Schininà (221) 87
Michoudet, C. and G. Baverel, Metabolism of acetaldehyde in human and baboon renal cortex: Ethanol synthesis by isolated baboon kidney-cortex tubules (216) 113
Migala, A. see W. Hasselbach (221) 119
Migliore-Samour, D. see J. Berthou (218) 55
Migus, A. see J. Breton (209) 37
Mikami, A. see T. Kubo (209) 367
Mikami, K., T. Tabata, T. Kawata, T. Nakayama and M. Iwabuchi, Nuclear protein(s) binding to the conserved DNA hexameric sequence postulated to regulate transcription of wheat histone genes (223) 273
Miki, K., V. Renganathan, M.B. Mayfield and M.H. Gold, Aromatic ring cleavage of a β-biphenyl ether dimer catalyzed by lignin peroxidase of *Phanerochaete chrysosporium* (210) 199
Mildner, P. see V. Mrša (217) 174
Miles, H.T. see V. Sklenář (208) 94
Milgrom, Ya.M. and M.B. Murataliev, Characterization of the nucleotide tight-binding sites of the isolated mitochondrial F_1-AT-Pase (219) 156
Milgrom, Ya.M. and M.B. Murataliev, On the rate of F_1-ATPase turnover during ATP hydrolysis by the single catalytic site: Evidence that hydrolysis with a slow rate of product release does not occur at the alternating active site (222) 32
Milgrom, Ya.M. and M.B. Murataliev, Steady-state rate of F_1-ATPase turnover during ATP hydrolysis by the single catalytic site (212) 63
Milhaud, G. see S. Minvielle (223) 63
Milia, A. see K.H. Cheeseman (209) 191
Millar, R.P. see C.E. Smith (225) 247
Miller, C.G. see Z. Xue (223) 391
Miller, jr, R.C. see M.L. Langsford (225) 163
Millet, P. see A. Brossi (214) 291
Millet, P. see A. Brossi (223) 77
Milligan, G. see J. Falloon (209) 352
Milligan, G. see R.M. O'Brien (212) 281
Millner, P.A., Are guanine nucleotide-binding proteins involved in regulation of thylakoid protein kinase activity? (226) 155
Milne, I.H. see J. Simpson (217) 62
Milton, G. see A.P. Jahagirdar (219) 83
Mimuro, M., N. Tamai, T. Yamazaki and I. Yamazaki, Excitation energy transfer in spinach chloroplasts: Analysis by the time-resolved fluorescence spectrum at $-196°C$ in the picosecond time range (213) 119
Minami, Y. see K. Ishidoh (226) 33
Minami, Y. see K. Suzuki (220) 271
Minami, Y. see S. Imajoh (215) 274
Minassian, G. see B. Rothhut (219) 169
Minassian, S. see N. Dyatkina (219) 151
Minchiotti, L. see M. Galliano (208) 364
Minegishi, T., J. Wang and P.C.K. Leung, Luteinizing hormone-releasing hormone (LHRH)-induced arachidonic acid release in rat granulosa cells: Role of calcium and protein kinase C (214) 139
Mineyev, A.P. and A.P. Razjivin, Statistics of the photon distribution in the set of photosynthetic antenna domains (223) 187
Minotti, G. see S. Borrello (209) 305
Minowa, O. see M. Ikura (219) 17
Minvielle, S., M. Cressent, M.C. Delehaye, N. Segond, G. Milhaud, A. Jullienne, M.S. Moukhtar and F. Lasmoles, Sequence and expression of the chicken calcitonin gene (223) 63
Mirande, M. see M. Lazard (216) 27
Mire-Sluis, A.R. see R.G. Wickremasinghe (220) 52
Mishina, M. see S. Noguchi (225) 27
Mishina, M. see T. Kurosaki (214) 253
Mishina, M. see T. Nukada (211) 5
Mishina, M. see T. Tobimatsu (222) 56
Misumi, Y. see K. Oda (214) 135
Mitani, T. see Y. Seino (223) 74
Mitchell, P., A new redox loop formality involving metal-catalysed hydroxide-ion translocation: A hypothetical Cu loop mechanism for cytochrome oxidase (222) 235
Mittre, H. see A. Benhaim (223) 321
Miwa, M. see H. Shima (209) 289
Miwa, S. see K. Kuwajima (221) 115
Miwatani, T. see Y. Shimonishi (215) 165
Miyahara, M. see E.F. Sato (214) 181
Miyamoto, K. see J. Abe (226) 58
Miyamoto, T., J.Å. Lindgren, T. Hökfelt and B. Samuelsson, Regional distribution of leukotriene and mono-hydroxyeicosatetraenoic acid production in the rat brain: Highest leukotriene C_4 formation in the hypothalamus (216) 123

Miyashiro, H. see V. Sklenář (208) 94
Miyashiro, S. see N. Tsuchimori (218) 205
Miyata, Y. see M. Hoshi (217) 237
Miyaura, C. see J. Abe (226) 58
Miyaura, C. see T. Hayashi (218) 200
Miyazaki, T. see Y. Uehara (208) 352
Miyoshi, T. see M. Hoshi (217) 237
Mizutani, T. and T. Hitaka, Stronger affinity of reticulocyte release factor than natural suppressor $tRNA^{Ser}$ for the opal termination codon (226) 227
Mockel, J. see I. Graff (210) 204
Modak, M.J. see V.N. Pandey (213) 204
Modrzycka, T. see K. Michalak (219) 233
Modyanov, N.N. see E.D. Sverdlov (217) 275
Modyanov, N.N. see E.D. Sverdlov (221) 129
Modyanov, N.N. see N.M. Matveeva (217) 42
Modyanov, N.N. see Yu.A. Ovchinnikov (213) 73
Modyanov, N.N. see Yu.A. Ovchinnikov (217) 111
Modyanov, N.N. see Yu.A. Ovchinnikov (217) 269
Mogre, R.M., H.F. Batliwala, P.S.R. Anjaneyulu and A.K. Lala, A new carbene based heterobifunctional reagent: Photochemical crosslinking of aldolase (221) 408
Mojsilovic, L. see D.R. Cooper (214) 122
Mokhova, E.N. see A.Yu. Andreyev (226) 265
Molle, G., H. Duclohier and G. Spach, Voltage-dependent and multi-state ionic channels induced by trichorzianines, anti-fungal peptides related to alamethicin (224) 208
Möller, W. see G.D.F. Maessen (208) 77
Möller, W. see G.D.F. Maessen (223) 181
Mollinedo, F. and D.L. Schneider, Intracellular organelle motility and membrane fusion processes in human neutrophils upon cell activation (217) 158
Momayezi, M. see B. Stecher (223) 25
Mommsen, T.P., P.C. Andrews and E.M. Plisetskaya, Glucagon-like peptides activate hepatic gluconeogenesis (219) 227
Momose, K. see I. Maruyama (220) 89
Monastyrskaya, G.S. see E.D. Sverdlov (217) 275
Monastyrskaya, G.S. see E.D. Sverdlov (221) 129
Monastyrskaya, G.S. see N.M. Matveeva (217) 42
Monastyrskaya, G.S. see Yu.A. Ovchinnikov (213) 73
Montecucco, C. see E. Papini (215) 73
Monteiro, M.J. and R.A. Cox, Differential expression of an α-tubulin gene during the development of *Physarum polycephalum* (217) 260
Montenarh, M. and D. Müller, The phosphorylation at Thr 124 of simian virus 40 large T antigen is crucial for its oligomerization (221) 199
Montesinos, C. see R. Serrano (208) 143
Montfoort, A., K. Bezstarosti, M.M.J. Groh and J.F. Koster, The influence of the chain length of aldehydes on the fluorescence of chromolipids (226) 101
Montibeller, J. see J.P. Rieker (212) 154
Moody, A.J., F. Norris, K. Norris, M.T. Hansen and L. Thim, The secretion of glucagon by transformed yeast strains (212) 302
Moolenaar, W.H. see L.G.J. Tertoolen (214) 365
Moonen, P., R. Gaffner and P. Wingfield, Native cytokines do not bind to uromodulin (Tamm-Horsfall glycoprotein) (226) 314
Moore, D.C.M. see I.N.M. Day (222) 139
Moore, G.A., D.J. McConkey, G.E.N. Kass, P.J. O'Brien and S. Orrenius, 2,5-Di(*tert*-butyl)-1,4-benzohydroquinone – a novel inhibitor of liver microsomal Ca^{2+} sequestration (224) 331
Moore, G.R. see R.T. Hartshorn (213) 391
Moore, S.E. see J. Thompson (219) 135
Moorhouse, P.C., M. Grootveld, B. Halliwell, J.G. Quinlan and J.M.C. Gutteridge, Allopurinol and oxypurinol are hydroxyl radical scavengers (213) 23
Moosburger, K. see J. Striessnig (212) 247
Morávek, L., O. Kühnemund, J. Havlíček, P. Kopecký and M. Pavlík, Type 1 M protein of *Streptococcus pyogenes*: N-terminal sequence and peptic fragments (208) 435
Moreau, J. and K. Scherrer, Co-evolution of base composition and codon usage in *Xenopus laevis* and human globin genes with long-range DNA organization of their genome (221) 3
Moreau, M. see C. Mazière (218) 243
Morehouse, K.M., W.D. Flitter and R.P. Mason, The enzymatic oxidation of Desferal to a nitroxide free radical (222) 246
Morel-Laurens, N. see P.C. Bibb (209) 169
Mori, H. see E. Takano (208) 199
Mori, H. see J. Yamaguchi (213) 329
Mori, H. see M. Maki (223) 174
Mori, T. see N. Tomita (225) 113
Mori, Y. see T. Kurosaki (214) 253
Mori, Y. see T. Tobimatsu (222) 56
Morich, F. see G. Theiss (218) 159
Morikawa, M. see J. Abe (226) 58
Morimoto, Y.M. see E.F. Sato (214) 181
Morin, J.-P. see D. Chansel (220) 247
Moritz, R.L. see R.J. Simpson (224) 128
Morningstar, J.E. see M.K. Johnson (226) 129
Mornon, J.P. see C. Gaboriaud (224) 149
Morozov, S.Yu., L.I. Lukasheva, B.K. Chernov, K.G. Skryabin and J.G. Atabekov, Nucleotide sequence of the open reading frames adjacent to the coat protein cistron in potato virus X genome (213) 438
Morozov, S.Yu. see V.V. Dolja (214) 308
Morris, A.P., C.M. Fuller and D.V. Gallacher, Cholinergic receptors regulate a voltage-insensitive but Na^+-dependent calcium influx pathway in salivary acinar cells (211) 195
Morrison, C.A., R.V. Fishleigh, D.J. Ward and B. Robson, Computer-aided design and physiological testing of a luteinising hormone-releasing hormone analogue for 'adjuvant-free' immunocastration (214) 65
Morrison, D.F., M.A. Rider and D.J. Takemoto, Modulation of retinal transducin and phosphodiesterase activities by synthetic peptides of the phosphodiesterase γ-subunit (222) 266
Morrow, C. see B. Pearce (211) 73
Mosbach, K. see L. Bülow (210) 147
Moser, B. see M.L. Langsford (225) 163
Motorin, Y.A., A.D. Wolfson, A.F. Orlovsky and K.L. Gladilin, Purification of valyl-tRNA synthetase high-molecular-mass complex from rabbit liver (220) 363
Motta, A., T. Tancredi and P.A. Temussi, Nuclear Overhauser effects in linear peptides: A low-temperature 500 MHz study of Met-enkephalin (215) 215
Moudgil, V.K., Steroid receptors in health and disease: An Oakland University-Serono Symposia, USA Conference, September 20–23, 1987, Meadow Brook Hall, Rochester, MI 48309-9908, USA (226) 213
Moukhtar, M.S. see S. Minvielle (223) 63
Moulinoux, J.P. see J.G. Delcros (220) 236
Moura, I. see G.D. Fauque (215) 63
Moura, J.J.G. see G.D. Fauque (215) 63
Moustaid, N. see M.K. Agarwal (217) 221
Movva, N.R. see P. Wingfield (215) 160
Mowbray, J. see W.E. Thomas (223) 279
Mrigank, see B. Chandrasekhar (225) 151
Mrigank, see K. Gopalakrishna (215) 95
Mrša, V., S. Barbarić, B. Ries and P. Mildner, Role of glycosylation in secretion of yeast acid phosphatase (217) 174
Mukherjee, B.B. see G.R. Laverdure (222) 261

Muldrow, L.L., G.C. Ibeanu, N.I. Lee, N.K. Bose and J. Johnson, Molecular cloning of *Clostridium difficile* toxin A gene fragment in λ gtII (213) 249
Mullany, P.P. see B.W. Wren (225) 82
Müller, D. see M. Montenarh (221) 199
Müller, V. see M. Blaut (215) 53
Müller-Taubenberger, A. see M. Westphal (209) 92
Mullin, J.M. and M.T. McGinn, The phorbol ester, TPA, increases transepithelial epidermal growth factor flux (221) 359
Multimäki, P. see V.-M. Kähäri (215) 331
Muno, D. see T. Isobe (223) 92
Murachi, T. see E. Takano (208) 199
Murachi, T. see M. Maki (223) 174
Muradov, K.G. see Yu.A. Ovchinnikov (223) 169
Murakami, N. see T. Konishi (226) 270
Muramatsu, T. see H.K. Mangold (220) 220
Murataliev, M.B. see Ya.M. Milgrom (212) 63
Murataliev, M.B. see Ya.M. Milgrom (219) 156
Murataliev, M.B. see Y.M. Milgrom (222) 32
Murayama, A. see C. Fukazawa (224) 125
Murazumi, N., K. Yokoyama, Y. Araki and E. Ito, An enzyme catalyzing the liberation of *N*-acetylglucosamine from *N*-acetylglucosaminyl pyrophosphorylpolyprenol in *Bacillus polymyxa* membranes (218) 131
Murdoch, F.A. see M. Bushfield (222) 299
Murer, H. see K. Malmström (216) 257
Murphy, J.B. see M.I. Khamis (211) 155
Murphy, P.M., B. Eide, P. Goldsmith, M. Brann, P. Gierschik, A. Spiegel and H.L. Malech, Detection of multiple forms of $G_{i\alpha}$ in HL60 cells (221) 81
Murphy, S. see B. Pearce (211) 73
Murray, P.G. see J. Ginsberg (226) 223
Muscillo, M. see A.M. Vaccaro (216) 190
Musgrave, A. see W. Homan (215) 323
Müssig, J. see M. Shoham (208) 321
Mustayev, A.A. see Yu.A. Ovchinnikov (217) 111
Mustelin, T., GTP dependence of the transduction of mitogenic signals through the T3 complex in T lymphocytes indicates the involvement of a G-protein (213) 199
Musto, N.A. see G.L. Hammond (215) 100
Muthukrishnan, S. see C.-S. Wang (222) 135
Mutt, V. see Z. Chen (226) 43
Muzya, G.I., E.N. Korobkova, N.K. Golovanova and L.D. Bergelson, Influence of prostaglandins on the lipid transfer between human high density and low density lipoproteins (220) 371
Myatt, J.F. see N.P.J. Cotton (219) 88
Myatt, J.F. see W. Crielaard (225) 6
Mynbeek, G. see A.J. Aarsman (219) 176

N

Nabiev, I.R. see N.G. Abdulaev (213) 113
Naccache, P.H. see L. Beaumier (221) 289
Nagai, Y. see N. Rodrig (221) 315
Nagamura, T. see K. Kuwajima (221) 115
Nagao, M., C. Sakamoto, T. Matozaki and S. Baba, Disulfide bonds within the α-subunit of insulin receptors in rat liver and brain membranes (214) 107
Nagasawa, H. see H. Jhoti (219) 419
Nagasawa, S. see M. Yamazaki (208) 147
Nagase, F. see Y. Kuwana (219) 360
Nagase, H. and K. Brew, Amino acid sequence of a 32-residue region around the thiol ester site in duck ovostatin (222) 83
Nagata, S., H. Günther, J. Bader and H. Simon, Mitochondria catalyze the reduction of NAD by reduced methylviologen (210) 66
Nagy, J.I. see J.D. Geiger (208) 431
Najjar, V.A. see N.J. Bump (226) 303
Nakada, H. see A. Kurosaka (215) 137
Nakagawa, S. see Y. Kuroda (224) 137
Nakajima, H., T. Noguchi, T. Yamasaki, N. Kono, T. Tanaka and S. Tarui, Cloning of human muscle phosphofructokinase cDNA (223) 113
Nakajima, R. see S. Hashimoto (208) 305
Nakaki, T. see N. Sasakawa (223) 413
Nakamura, M. see H. Kai (212) 119
Nakamura, M. see H. Kai (221) 284
Nakamura, M. see H. Kanaide (214) 130
Nakamura, M. see M. Fujii (223) 299
Nakamura, T., K. Nawa, A. Ichihara, N. Kaise anJ T. Nishino, Purification and subunit structure of hepatocyte growth factor from rat platelets (224) 311
Nakamura, T. see K. Akiyama (225) 168
Nakamura, T. see M. Sato (224) 29
Nakano, H. see T. Tanimoto (226) 291
Nakano, M. see K. Sugioka (210) 37
Nakano, M. see Y. Sugioka (223) 251
Nakashima, I. see Y. Kuwana (219) 360
Nakayama, H., H. Shikano, T. Aoyama, T. Amano and Y. Kanaoka, Affinity purification of the opioid receptor in NG 108-15 cells using an avidin-biotin system with a novel elution method (208) 278
Nakayama, H. see Y. Kuroda (224) 137
Nakayama, T. see K. Mikami (223) 273
Nakazato, M. see T. Hiyama (214) 97
Nandan, D. see G.A. Cates (218) 195
Naor, Z. see C.E. Smith (225) 247
Narbonne, J.F. see Z. Amelizad (220) 231
Narindrasorasak, S. see G.A. Cates (218) 195
Närvänen, A. see O. Närvänen (224) 156
Närvänen, O., A. Närvänen, V.-M. Wasenius, P. Partanen and I. Virtanen, A monoclonal antibody against a synthetic peptide reveals common structures among spectrins and α-actinin (224) 156
Närvänen, O. see V.M. Wasenius (221) 73
Nash, B. see S.S. Tate (211) 133
Nasmith, P.E. and S. Grinstein, Are Ca^{2+} channels in neutrophils activated by a rise in cytosolic free Ca^{2+}? (221) 95
Nass, G.S. see L.F. Levinger (209) 340
Nath, A. see G.P. Püschel (219) 145
Natori, S. see K. Itoh (213) 85
Navaroli, F. see L.E. Mallea (218) 143
Navarro, J. see B.F. Dickey (219) 289
Nave, R. see D. Fürst (224) 49
Nawa, K. see T. Nakamura (224) 311
Nazar, R.N. see J. McDougall (209) 52
Neale, P.J. see S. Demeter (214) 370
Nebreda, A.R., C.R. Vazquez, T.G. Villa, J.R. Villanueva and F. del Rey, Heterogeneous glycosylation of the *EXG1* gene product accounts for the two extracellular exo-β-glucanases of *Saccharomyces cerevisiae* (220) 27
Neckers, L. see D. Katsaros (223) 97
Nedergaard, J. see A. Jacobsson (224) 353
Nedospasov, A.A. see J. Šimúth (218) 163
Négrerie, M. see D. Aslanian (219) 202
Neher, E. see R. Penner (226) 307
Neidle, S. see L.H. Pearl (209) 269
Nelson, B. see Z. Dancshazy (209) 44
Nelson, D.A. see Q.A. Vu (220) 79

Nelson, D. see M. Erecińska (213) 61
Nestel, P.J. see P.D. Roach (222) 159
Nettelblad, F.A. and L. Engström, The kinetic effects of in vitro phosphorylation of rabbit muscle enolase by protein kinase C: A possible new kind of enzyme regulation (214) 249
Neumann, J.M. see G. Bloch (219) 464
Neupert, W. see N. Pfanner (209) 152
Neuser, D. and P. Bellemann, Receptor binding, cGMP stimulation and receptor desensitization by atrial natriuretic peptides in cultured A10 vascular smooth muscle cells (209) 347
Neuser, D. see G. Theiss (218) 159
Nevinsky, G.A. see O.I. Lavrik (216) 225
Nevinsky, G.A. see S.V. Doronin (216) 221
Newsholme, E.A. see B. Leighton (225) 93
Newsome, P. see I. Dodd (209) 13
Newton, R.C. see J.J. Huang (223) 294
Nexø, B.A. see E. Boel (219) 181
Nicchitta, C.V., S.K. Joseph and J.R. Williamson, Polyethylene glycol-stimulated microsomal GTP hydrolysis: Relationship to GTP-mediated Ca^{2+} release (209) 243
Nice, E.C. see R.J. Simpson (224) 128
Nicholls, N.J. and J.O. Lampen, Repressor gene, *blaI*, for *Bacillus licheniformis* 749 β-lactamase: (221) 179
Nicholls, P. see C.E. Cooper (223) 155
Nicholson, J.K. see J.D. Bell (215) 311
Nicholson, P., R.W. Osborn and C.J. Howe, Induction of protein synthesis in response to ultraviolet light, nalidixic acid and heat shock in the cyanobacterium *Phormidium laminosum* (221) 110
Nickerson, J.M. see T. van Veen (208) 133
Nicolay, K., E.B. Smaal and B. de Kruijff, Ethylene glycol causes acyl chain disordering in liquid-crystalline, unsaturated phospholipid model membranes, as measured by ^2H NMR (209) 33
Nicotera, P., P. Hartzell, G. Davis and S. Orrenius, The formation of plasma membrane blebs in hepatocytes exposed to agents that increase cytosolic Ca^{2+} is mediated by the activation of a non-lysosomal proteolytic system (209) 139
Niedermann, G. see J. Lasch (214) 13
Nielsen, H.V. see J.E. Celis (220) 1
Nielsen, J.R., H.S. Hansen and B. Jensen, Arginine-vasopressin stimulates the formation of phosphatidic acid in rat Leydig cells (218) 93
Nielsen, L.S. see P.A. Andreasen (209) 213
Nielsen, O.V. see J.J. Holst (211) 169
Nielsen, P.S. and K. Gausing, The precursor of barley plastocyanin: Sequence of cDNA clones and gene expression in different tissues (225) 159
Nieto, J.L. see M.A. Jiménez (221) 320
Nijkamp, F.P. see F. Engels (209) 249
Nikandrov, V.V. see A.A. Krasnovsky (219) 93
Nikiforov, T. see B. Appelhans (224) 14
Nikolsky, N.N. see L.V. Teslenko (221) 105
Nilges, M., G.M. Clore and A.M. Gronenborn, A simple method for delineating well-defined and variable regions in protein structures determined from interproton distance data (219) 11
Nilges, M. see G.M. Clore (213) 269
Nimmo, G.A., G.A.L. McNaughton, C.A. Fewson, M.B. Wilkins and H.G. Nimmo, Changes in the kinetic properties and phosphorylation state of phospho*enol*pyruvate carboxylase in *Zea mays* leaves in reponse to light and dark (213) 18
Nimmo, H.G. see G.A. Nimmo (213) 18
Ninio, J. see P.J. Lecomte (221) 194
Nishida, E. see M. Hoshi (217) 237
Nishida, E. see Y. Ohta (208) 423
Nishida, E. see Y. Ohta (222) 305
Nishii, Y. see J. Abe (226) 58
Nishikawa, Y. see V. Witzemann (223) 104
Nishimoto, S., H. Akutsu and Y. Kyogoku, The presence of the 30 nm filament structure of chromatins in intact chicken erythrocytes observed by ^{31}P NMR (213) 293
Nishimura, C. see D. Carper (220) 209
Nishimura, M. see J. Yamaguchi (213) 329
Nishimura, M. see Y. Isogai (224) 71
Nishino, T. see T. Nakamura (224) 311
Nishitani, H. see M. Ohta (222) 79
Nishizuka, Y. see M. Ido (219) 215
Nishizuka, Y. see U. Kikkawa (217) 227
Nishizuka, Y. see U. Kikkawa (223) 212
Nishizuka, Y. see Y. Ono (226) 125
Nissenson, C. see S.C. Huang (216) 128
Nitschke, W. and G. Hauska, An asymmetric low-spin EPR signal of cytochrome b_6 in the cytochrome b_6f-complex from spinach chloroplasts (213) 453
Nitschke, W. see K. Ziegler (217) 16
Nitschke, W., U. Feiler, W. Lockau and G. Hauska, The photosystem of the green sulfur bacterium *Chlorobium limicola* contains two early electron acceptors similar to photosystem I (218) 283
Nitta, K., H. Tsuge, S. Sugai and K. Shimazaki, The calcium-binding property of equine lysozyme (223) 405
Nixon, P.J., T.A. Dyer, J. Barber and C.N. Hunter, Immunological evidence for the presence of the D1 and D2 proteins in PS II cores of higher plants (209) 83
Niznik, H.B., D.E. Grigoriadis and P. Seeman, Photoaffinity labelling of dopamine D_2 receptors by [^3H]azidomethylspiperone (209) 71
Noegel, A. see M. Westphal (209) 92
Noegel, A., W. Witke and M. Schleicher, Calcium-sensitive non-muscle α-actinin contains EF-hand structures and highly conserved regions (221) 391
Noguchi, S., M. Mishina, M. Kawamura and S. Numa, Expression of functional ($Na^+ + K^+$)-ATPase from cloned cDNAs (225) 27
Noguchi, S. see T. Hiwasa (211) 23
Noguchi, T. see H. Nakajima (223) 113
Noguiez, P. see Y. Le Bouc (222) 181
Nohl, H., A novel superoxide radical generator in heart mitochondria (214) 269
Nojima, H., Molecular evolution of the calmodulin gene (217) 187
Noll, K.M. see D. Ankel-Fuchs (213) 123
Nomura, K., Specificity of prolyl endopeptidase (209) 235
Nomura, Y. and M. Tohda, Inhibitory effects of pertussis toxin on a depolarization-evoked Ca^{2+} influx in NG108-15 cells (216) 40
Nordmann, R. see F. Bauché (219) 296
Nordstedt, C., M. Jondal and B.B. Fredholm, Activation of protein kinase C inhibits prostaglandin- and potentiates adenosine receptor-stimulated accumulation of cyclic AMP in a human T-cell leukemia line (220) 57
Norling, B., B. Hamasur and E. Glaser, Cross-reconstitution of isolated F_1-ATPase from potato tuber mitochondria with F_1-depleted beef heart and yeast submitochondrial particles (223) 309
Norling, B. see E. Glaser (223) 304
Norman, R.I., A.J. Burgess, E. Allen and T.M. Harrison, Monoclonal antibodies against the 1,4-dihydropyridine receptor associated with voltage-sensitive Ca^{2+} channels detect similar polypeptides from a variety of tissues and species (212) 127
Norris, F. see A.J. Moody (212) 302
Norris, J.R. see T.J. Michalski (226) 72
Norris, K. see A.J. Moody (212) 302
North, A.C.T. see D.K. Stammers (218) 178

North, M.J. see P.W. Watt (215) 295
Northoff, H. see T. Andus (221) 18
Norton, J.D., B. Leber and J.C. Yaxley, Patterns of gene expression during plasmacytoid differentiation of chronic lymphocytic leukaemia cells (215) 127
Nose, K. see M. Itami (222) 289
Noteborn, H.P.J.M., J.P.H. Burbach and I. Ebels, Modified forms of vasopressin and oxytocin in a bovine pineal preparation (216) 200
Notton, B.A. see M.J. Barber (213) 372
Noumi, T., M. Maeda and M. Futai, Mode of inhibition of sodium azide on H^+-ATPase of *Escherichia coli* (213) 381
Noumi, T. see S.-Y. Hsu (218) 222
Novoselov, V.I. see E.E. Fesenko (219) 224
Novotný, J. and R.E. Bruccoleri, Correlation among sites of limited proteolysis, enzyme accessibility and segmental mobility (211) 185
Nucci, D. see M. Bigio (218) 271
Nugent, J.H.A. see R.W. Mansfield (220) 74
Nuijs, A.M. see R.V. Danielius (213) 241
Nukada, T., M. Mishina and S. Numa, Functional expression of cloned cDNA encoding the α-subunit of adenylate cyclase-stimulating G-protein (211) 5
Numa, S. see S. Noguchi (225) 27
Numa, S. see T. Kubo (209) 367
Numa, S. see T. Kurosaki (214) 253
Numa, S. see T. Nukada (211) 5
Numa, S. see T. Tobimatsu (222) 56
Numa, S. see V. Witzemann (223) 104
Numata, Y. see A. Kurosaka (215) 137
Nunez, D. see J. Benveniste (226) 371
Nussbaum, O. and A. Loyter, Quantitative determination of virus-membrane fusion events: Fusion of influenza virions with plasma membranes and membranes of endocytic vesicles in living cultured cells (221) 61
Nüße, O. see M. Lindau (222) 317
Nutman, T.B. see A.P. Weetman (211) 69
Nuutinen, M. see J. Londesborough (219) 249
Nybroe, O. see E. Bock (225) 33
Nygren, P., R. Larsson, E. Lindh, S. Ljunghall, J. Rastad, G. Åkerström and E. Gylfe, Bimodal regulation of secretion by cytoplasmic Ca^{2+} as demonstrated by the parathyroid (213) 195

O

Obara, M., M.S. Kang, S. Rocher-Dufour, A. Kornblihtt, J.P. Thiery and K.M. Yamada, Expression of the cell-binding domain of human fibronectin in *E. coli*: Identification of sequences promoting full to minimal adhesive function (213) 261
Oberbäumer, I. see U. Schwarz-Magdolen (208) 203
Oberdieck, U. see F. Hucho (211) 207
Obidin, A.B. see N.V. Gulyaeva (211) 211
O'Brian, C.A., W.L. Arthur and I.B. Weinstein, The activation of protein kinase C by the polyphosphoinositides phosphatidylinositol 4,5-diphosphate and phosphatidylinositol 4-monophosphate (214) 339
O'Brien, E. see E.A. Usanga (209) 23
O'Brien, P.J. see G.A. Moore (224) 331
O'Brien, R.M., K. Siddle, M.D. Houslay and A. Hall, Interaction of the human insulin receptor with the ras oncogene product p21 (217) 253
O'Brien, R.M., M.D. Houslay, G. Milligan and K. Siddle, The insulin receptor tyrosyl kinase phosphorylates holomeric forms of the guanine nucleotide regulatory proteins G_i and G_o (212) 281
Octave, J.N. see M.I. Darville (224) 317
Oda, K., S. Hirose, N. Takami, Y. Misumi and A.T. andYukio Ikehara, Brefeldin A arrests the intracellular transport of a precursor of complement C3 before its conversion site in rat hepatocytes (214) 135
Odani, S. see T. Koide (216) 17
O'Donoghue, D.J. see M.E. Hayes (220) 307
Oehen, S., J. Parsons, D.A. Jans and B.A. Hemmings, Recombinant type I regulatory subunit of the cAMP-dependent protein kinase is biologically active (220) 47
Oesch, F. see Z. Amelizad (220) 231
Oesterhelt, D. see R. Diller (217) 297
Oetting, M., M. LeBoff, L. Swiston, J. Preston and E. Brown, Guanine nucleotides are potent secretagogues in permeabilized parathyroid cells (208) 99
Ogawa, M. see N. Tomita (225) 113
Ogawara, H. see M. Hoshi (217) 237
Ogden, D.C. see T. Capiod (217) 247
Ogg, D. see D.K. Stammers (218) 178
Ogita K. see U. Kikkawa (217) 227
Ogita, K. see U. Kikkawa (223) 212
Ogita, K. see Y. Ono (226) 125
Ogren, W.L. see M.E. Salvucci (221) 215
Oh-oka, H., Y. Takahashi, K. Wada, H. Matsubara, K. Ohyama and H. Ozeki, The 8 kDa polypeptide in photosystem I is a probable candidate of an iron-sulfur center protein coded by the chloroplast gene *frxA* (218) 52
Ohad, I. see A. Gal (221) 205
O'Hara, D.M. and P.E. Reynolds, Antibody used to identify penicillin-binding protein 2′ in methicillin-resistant strains of *Staphylococcus aureus* (MRSA) (212) 237
Ohara, O. and H. Teraoka, Anomalous behavior of human leukocyte interferon subtypes on polyacrylamide gel electrophoresis in the presence of dodecyl sulfate (211) 78
O'Hare, M., L.M. Roberts, P.E. Thorpe, G.J. Watson, B. Prior and J.M. Lord, Expression of ricin A chain in *Escherichia coli* (216) 73
O'Hare, M.M.T. see T.W. Schwartz (225) 209
Ohashi, Y. see M. Ohshima (225) 243
Ohishi, I. see F.A. Al-Mohanna (219) 40
Ohkura, K. see S. Inoue (218) 17
Ohmori, D. see T. Isobe (223) 92
Ohmori, T. see T. Tanimoto (226) 291
Ohmura, Y., H. Teraoka and K. Tsukada, A protein-tyrosine kinase in the nuclear matrix from rat liver (208) 451
Ohnishi, J.-I. and R. Kanai, Na-induced uptake of pyruvate into mesophyll chloroplasts of a C_4 plant, *Panicum miliaceum* (219) 347
Ohnishi, T. see J. Telser (214) 117
Ohno, S., M. Aoshima, S. Matsumoto, I. Yahara and K. Suzuki, A yeast gene coding for a putative protein kinase homologous to *cdc25* suppressing protein kinase (222) 279
Ohno, S. see K. Ishidoh (226) 33
Ohno, S. see K. Kubo (223) 138
Ohno, S. see K. Suzuki (220) 271
Ohshima, M., M. Matsuoka, N. Yamamoto, Y. Tanaka, Y. Kano-Murakami, Y. Ozeki, A. Kato, N. Harada and Y. Ohashi, Nucleotide sequence of the PR-1 gene of *Nicotiana tabacum* (225) 243
Ohta, K. see M. Ohta (222) 79

Ohta, M., K. Ohta, H. Nishitani and K. Hayashi, Primary structure of α-bungarotoxin: Six amino acid residues differ from the previously reported sequence (222) 79
Ohta, S., K. Goto, H. Arai and Y. Kagawa, An extremely acidic amino-terminal presequence of the precursor for the human mitochondrial hinge protein (226) 171
Ohta, T. see H. Adachi (226) 150
Ohta, Y., E. Nishida and H. Sakai, Type II Ca^{2+}/calmodulin-dependent protein kinase binds to actin filaments in a calmodulin-sensitive manner (208) 423
Ohta, Y., T. Akiyama, E. Nishida and H. Sakai, Protein kinase C and cAMP-dependent protein kinase induce opposite effects on actin polymerizability (222) 305
Ohtani, K. see M. Fujii (223) 299
Ohtsuka, E. see H. Inoue (215) 327
Ohtsuka, K. see I. Maruyama (220) 89
Ohyama, K. see H. Fukuzawa (220) 61
Ohyama, K. see H. Oh-oka (218) 52
Oinuma, M. see T. Katada (213) 353
Oka, T. see K. Enomoto (223) 82
Okabe, T. and K. Sobue, Identification of a new 84/82 kDa calmodulin-binding protein, which also interacts with actin filaments, tubulin and spectrin, as synapsin I (213) 184
Okada, H. see M. Ueda (220) 31
Okada, Y. see Y. Watanabe (219) 65
Okon, E.B. and N.N. Vsevolodov, Does bacteriorhodopsin energize the membranes of animal mitochondria under light? (216) 241
Okumura, T. see H. Fukuzawa (220) 61
Okuyama, T. see T. Ichimura (219) 79
Okuyama, T. see T. Isobe (223) 92
Olafsson, I. see M. Abrahamson (216) 229
Olins, G.M., P.P. Mehta, D.J. Blehm, D.R. Patton, M.E. Zupec, D.E. Whipple, F.S. Tjoeng, S.P. Adams, P.O. Olins and J.K. Gierse, Phosphorylation of high- and low-molecular-mass atrial natriuretic peptide analogs by cyclic AMP-dependent protein kinase (224) 325
Olins, P.O. see G.M. Olins (224) 325
Olive, J., I. M'Bina, C. Vernotte, C. Astier and F.A. Wollman, Randomization of the EF particles in thylakoid membranes of *Synechocystis* 6714 upon transition from state I to state II (208) 308
Oliver, M. see M.K. Johnson (226) 129
Olivera, B.M. see D.H. Feldman (214) 295
Olkinuora, M. see L. Keso (215) 105
Olsson, H., P. Strålfors and P. Belfrage, Phosphorylation of the basal site of hormone-sensitive lipase by glycogen synthase kinase-4 (209) 175
Olsson, R.A. see D. Ukena (209) 122
Olsson, R.A. see D. Ukena (215) 203
O'Neill, L.A.J., M.L. Barrett and G.P. Lewis, Induction of cyclo-oxygenase by interleukin-1 in rheumatoid synovial cells (212) 35
Ono, S., S. Lee, Y. Kodera, H. Aoyagi, M. Waki, T. Kato and N. Izumiya, Environment-dependent conformation and antimicrobial activity of a gramicidin S analog containing leucine and lysine residues (220) 332
Ono, T. see I. Vass (211) 215
Ono, Y. see U. Kikkawa (217) 227
Ono, Y. see U. Kikkawa (223) 212
Ono, Y., T. Fujii, K. Ogita, U. Kikkawa, K. Igarashi and Y. Nishizuka, Identification of three additional members of rat protein kinase C family: δ-, ε- and ζ-subspecies (226) 125
Oono, K. see F. Takaiwa (221) 43
Oosawa, M. and K. Maruyama, Reannealing of phalloidin-treated actin filaments during recovery after sonication and its inhibition by β-actinin (213) 433
Oosterwijk, C. see P. de Pagter-Holthuizen (214) 259
Opekarovà, M., A.J.M. Driessen and W.N. Konings, Protonmotive-force-driven leucine uptake in yeast plasma membrane vesicles (213) 45
Oplatka, A. see P.I. Lelkes (208) 357
Oppenheimer, C. see P.A. Andreasen (209) 213
O'Rourke, F., G.B. Zavoico, L.H. Smith, jr and M.B. Feinstein, Stimulus-response coupling in a cell-free platelet membrane system: GTP-dependent release of Ca^{2+} by thrombin, and inhibition by pertussis toxin and a monoclonal antibody that blocks calcium release by IP_3 (214) 176
Oretskaya, T.S. see V.G. Metelev (226) 232
Orlovsky, A.F. see Y.A. Motorin (220) 363
Ornatsky, O.I. see M.A. Glukhova (218) 292
Orrenius, S. see G.A. Moore (224) 331
Orrenius, S. see P. Nicotera (209) 139
Ørskov, C. see J.J. Holst (211) 169
Osanai, T. see N. Rodrig (221) 315
Osborn, M. see D. Fürst (224) 49
Osborn, R.W. see P. Nicholson (221) 110
Oshiro, S., N. Katsura, K. Kitada and N. Gunge, A simple method for extraction of DNA from fungi and yeasts with anhydrous hydrogen fluoride (220) 383
Oshtrakh, M.I. and V.A. Semionkin, Mössbauer spectroscopy of haemoglobins: Study of the relationship of Fe^{2+} electronic and molecular structure of the active site (208) 331
Osuna, C., A. Galván and M. Lucas, Impaired calcium sequestration activity in liver microsomes from fasted rats (211) 41
O'Toole, L. see J.M. Conlon (214) 50
Oursel, A., A. Escoffier, J.C. Kader, J.P. Dubacq and A. Trémolières, Last step in the cooperative pathway for galactolipid synthesis in spinach leaves: formation of mono-galactosyldiacylglycerol with C18 polyunsaturated acyl groups at both carbon atoms of the glycerol (219) 393
Ovádi, J. see P. Tompa (214) 244
Ovchinnikov, Y.A. see E.D. Sverdlov (221) 129
Ovchinnikov, Yu.A., G.S. Monastyrskaya, N.E. Broude, R.L. Allikmets, Yu.A. Ushkaryov, A.M. Melkov, Yu.V. Smirnov, I.V. Malyshev, I.E. Dulubova, K.E. Petrukhin, A.V. Gryshin, V.E. Sverdlov, N.I. Kiyatkin, M.B. Kostina, N.N. Modyanov and E.D. Sverdlov, The family of human Na^+,K^+-ATPase genes: A partial nucleotide sequence related to the α-subunit (213) 73
Ovchinnikov, Yu.A., K.N. Dzhandzhugazyan, S.V. Lutsenko, A.A. Mustayev and N.N. Modyanov, Affinity modification of E_1-form of Na^+,K^+-ATPase revealed Asp-710 in the catalytic site (217) 111
Ovchinnikov, Yu.A., N.M. Arzamazova, E.A. Arystarkhova, N.M. Gevondyan, N.A. Aldanova and N.N. Modyanov, Detailed structural analysis of exposed domains of membrane-bound Na^+,K^+-ATPase: A model of transmembrane arrangement (217) 269
Ovchinnikov, Yu.A. see A.S. Arseniev (213) 283
Ovchinnikov, Yu.A. see E.D. Sverdlov (217) 275
Ovchinnikov, Yu.A. see N.M. Matveeva (217) 42
Ovchinnikov, Yu.A., V.V. Gubanov, N.V. Khramtsov, K.A. Ischenko, V.E. Zagranichny, K.G. Muradov, T.M. Shuvaeva and V.M. Lipkin, Cyclic GMP phosphodiesterase from bovine retina: Amino acid sequence of the α-subunit and nucleotide sequence of the corresponding cDNA (223) 169
Ovchinnikov, Yu.A., V.Z. Slepak, A.N. Pronin, A.B. Shlensky, N.B. Levina, V.L. Voeikov and V.M. Lipkin, Primary structure of bovine cerebellum GTP-binding protein G_{39} and its effect on the adenylate cyclase system (226) 91
Overath, P. see B. Ehlers (225) 53
Overath, P. see F. Jähnig (221) 37
Owada, M.K. see H. Hayashi (223) 267
Owens, R.J. see B. Carnemolla (215) 269

Oxborough, K., P. Lee and P. Horton, Regulation of thylakoid protein phosphorylation by high-energy-state quenching (221) 211
Oyamada, H. see I. Maruyama (220) 89
Ozawa, F. see T. Yamakuni (223) 117
Ozeki, H. see H. Oh-oka (218) 52
Ozeki, Y. see M. Ohshima (225) 243

P

Paccaud, J.-P. see J.A. Schifferli (213) 415
Padgett, W.L. see D. Ukena (209) 122
Padgett, W.L. see D. Ukena (215) 203
Pagala, M.K. see R.J. Bradley (224) 277
Pagès, J.M. see J.M. Bolla (224) 213
Pain, R.H. see P. Wingfield (211) 179
Pairault, J. see B. Feve (219) 56
Palace, G.P., J.E. Franke and J.T. Warden, Is phylloquinone an obligate electron carrier in photosystem I? (215) 58
Paleologue, A., J.P. Reboud and A.-M. Reboud, Modifications of 60 S ribosomal subunits induced by the ricin A chain (208) 373
Pallet, V. see M. Savart (216) 22
Palmblad, J. see C.N. Serhan (217) 242
Palmer, G. see J. Telser (214) 117
Palmqvist, K. see L.-G. Sundblad (209) 28
Palombini, G. see S. Borrello (209) 305
Pan, R.L. see B.-D. Hsu (217) 53
Panchenko, L.F. see V.D. Antonenkov (224) 357
Pandey, V.N., S.B. Amrute, J.G. Satav and M.J. Modak, Inhibition of terminal deoxynucleotidyl transferase by adenine dinucleotides: Unique inhibitory action of Ap5A (213) 204
Pandiella-Alonso, A., A. Malgaroli, L.M. Vicentini and J. Meldolesi, Early rise of cytosolic Ca^{2+} induced by NGF in PC12 and chromaffin cells (208) 48
Pannuti, A. see A. Pascucci (226) 297
Pannuti, A. see L. Lania (219) 400
Papa, S. see F. Guerrieri (213) 67
Papas, T.S. see K.P. Samuel (218) 81
Papini, E., R. Colonna, G. Schiavo, F. Cusinato, M. Tomasi, R. Rappuoli and C. Montecucco, Diphtheria toxin and its mutant *crm* 197 differ in their interaction with lipids (215) 73
Papós, M. see I. Rosztóczy (208) 56
Pappin, D.J.C. see A. Cavaggioni (212) 225
Paquin, J. see N.G. Seidah (211) 144
Parente, J.E. see J. Ginsberg (226) 223
Parés, X. see T. Fairwell (222) 99
Parini, C. see P.L. Manachini (214) 305
Parker, K.R., L.E. Schaechter, J.W. Lewis, K.L. Zeman, D.S. Kliger and E.A. Dratz, Mg^{2+}-ATP induces filament growth from retinal rod outer segments with disrupted plasma membranes (211) 35
Parrack, P., D. Dasgupta, J. Ayyer and V. Sasisekharan, Interaction of synthetic analogs of distamycin with DNA: Role of the conjugated *N*-methylpyrrole system in specificity of binding (212) 297
Parsonage, D. see T.M. Duncan (208) 1
Parsons, J. see S. Oehen (220) 47
Partanen, P. see O. Närvänen (224) 156
Pascall, J.C. see A.J. Harmar (208) 67
Paschenko, V.Z., B.N. Korvatovsky, S.L. Logunov, A.A. Kononenko, P.P. Knox, N.I. Zakharova, N.P. Grishanova and A.B. Rubin, Modification of protein hydrogen bonds influences the efficiency of picosecond electron transfer in bacterial photosynthetic reaction centers (214) 28
Pascucci, A., A. Pannuti, G. La Mantia and L. Lania, Sequences both 5′ and 3′ to the transcription initiation site contribute to the ability of a mouse H-2 gene to respond to type I interferon (226) 297
Pasternak, G. see R. Grosse (210) 123
Pastuszko, A., D.K. Yee and D.F. Wilson, Regulation of calcium uptake in synaptosomes from rat brain by DL-2-amino-5-phosphonovaleric acid (218) 189
Patarroyo, M., E.A. Clark, J. Prieto, C. Kantor and C.G. Gahmberg, Identification of a novel adhesion molecule in human leukocytes by monoclonal antibody LB-2 (210) 127
Patel, H. see M. Beggs (215) 13
Patel, S.D. see F.G.P. Earley (219) 108
Patil, D.S. see E. Samain (216) 140
Patsch, W. see C. Yang (224) 261
Patterson, S. see M.J.O. Wakelam (210) 181
Patthey, J.-P. and J. Deshusses, Accessibility of *Trypanosoma brucei* procyclic glycosomal enzymes to labeling agents of various sizes and charges (210) 137
Patthy, L., Intron-dependent evolution: preferred types of exons and introns (214) 1
Patton, C.L. see P.G. Penketh (221) 427
Patton, D.R. see G.M. Olins (224) 325
Patton, S. and G.E. Huston, Differences between individuals in high-M_r glycoproteins from human mammary epithelia (216) 151
Pattus, F. see B. Dargent (220) 136
Paulsson, M. see K. Mann (218) 167
Pauwels, S. see H. Desmond (210) 185
Pavlík, M. see L. Morávek (208) 435
Pavlovitch, J.H. see C.P. Schelling (214) 21
Payne, W.E. and B.L. Trumpower, A simple, one-step purification of cytochrome *b* from the bc_1 complexes of bacteria (213) 107
Payne, W.J. see M. -C. Liu (218) 227
Pearce, B., J. Jeremy, C. Morrow, S. Murphy and P. Dandona, Inositol phospholipids are probably not the source of arachidonic acid for eicosanoid synthesis in astrocytes (211) 73
Pearce, I.A., M.A. Cambray-Deakin and R.D. Burgoyne, Glutamate acting on NMDA receptors stimulates neurite outgrowth from cerebellar granule cells (223) 143
Pearl, L.H. and S. Neidle, Origins of stereospecificity in DNA damage by *anti*-benzo[*a*]pyrene diol-epoxides: A molecular modelling study (209) 269
Pearl, L.H., The catalytic mechanism of aspartic proteinases (214) 8
Pecceu, F. see P. Ferrara (226) 47
Peck, jr, H.D. see M.-C. Liu, (218) 227
Peck, jr, H.D. see Y.M. Berlier (221) 241
Pedak, A. see V. Tóugu (225) 77
Peisach, J. see D.M. Dooley (214) 274
Pelton, J.T. see J.M. Berman (220) 214
Penketh, P.G., W.P.K. Kennedy, C.L. Patton and A.C. Sartorelli, Trypanosomatid hydrogen peroxide metabolism (221) 427
Penner, R. and E. Neher, Secretory responses of rat peritoneal mast cells to high intracellular calcium (226) 307
Penningroth, S.M., P.M. Rose and D.D. Peterson, Evidence that the 116 kDa component of kinesin binds and hydrolyzes ATP (222) 204
Pépin, D. see C. de Paillerets (219) 113
Pepin, D. see J. Masliah (222) 11
Perbal, B. see C. Kryceve-Martinerie (214) 81

Perea, X. see J.-P. di Rago (208) 208
Pérez-Ortín, J.E., F. Estruch, E. Matallana and L. Franco, Sliding-end-labelling: A method to avoid artifacts in nucleosome positioning (208) 31
Périer, C. see F. Baklouti (223) 59
Periquet, A. see Z. Amelizad (220) 231
Perrotti, N. see S.A. Phillips (212) 141
Perry, M.B. see D.R. Bundle (216) 261
Persaud, S.J. see P.M. Jones (219) 139
Pert, C.B. see M.R. Ruff (211) 17
Perutz, M.F. see M. Brunori (221) 161
Peschek, G.A. see B. Hinterstoisser (217) 169
Peskar, B.A. see U. Hoppe (208) 26
Peskar, B.M. see U. Hoppe (208) 26
Peskin, A.V. and G. Bartosz, One-electron reduction of an anthracycline antibiotic carminomycin by a human erythrocyte redox chain (219) 212
Peterajová, E. see Ľ. Varečka (225) 173
Petersen, J. see D.A. Klærke (216) 211
Petersen, N.O. see P.L. Vandewalle (210) 195
Petersen, O.H. see M.J. Dunne (208) 59
Petersen, V.B. see J. Furmaniak (215) 316
Peterson, D.D. see S.M. Penningroth (222) 204
Petit, J.-F. see J.-P. Tenu (220) 93
Petra, P.H. see B.G. Que (219) 405
Petratos, K., D.W. Banner, T. Beppu, K.S. Wilson and D. Tsernoglou, The crystal structure of pseudoazurin from *Alcaligenes faecalis* S-6 determined at 2.9 Å resolution (218) 209
Petrich, J. see J. Breton (209) 37
Petrukhin, K.E. see E.D. Sverdlov (217) 275
Petrukhin, K.E. see E.D. Sverdlov (221) 129
Petrukhin, K.E. see Yu.A. Ovchinnikov (213) 73
Peumans, W.J. see A.M. Vranken (216) 67
Peumans, W.J. see E.J.M. Van Damme (215) 140
Peumans, W.J. see W.F. Broekaert (220) 116
Pezzella, K. see J.J. Huang (223) 294
Pfaltz, A. see A. Kobelt (214) 265
Pfaltz, A. see J. Ellermann (220) 358
Pfanner, N. and W. Neupert, Transport of F_1-ATPase subunit β into mitochondria depends on both a membrane potential and nucleoside triphosphates (209) 152
Pfanzagl, B. see J. Berenguer (224) 401
Pfeiffer, A. see B. Tocqué (222) 327
Pfeiffer, A. see Y. Shimohigashi (222) 71
Pfleiderer, G. see J. Hönes (212) 193
Phelps, M.E. see S.C. Huang (216) 128
Philippot, J.R. see M. Vidal (216) 159
Philippov, P.P. see V.Yu. Arshavsky (224) 19
Phillips, J.W. see M.N. Berry (224) 201
Phillips, S.A., N. Perrotti and S.I. Taylor, Rat liver membranes contain a 120 kDa glycoprotein which serves as a substrate for the tyrosine kinases of the receptors for insulin and epidermal growth factor (212) 141
Philp, N.J., W. Chang and K. Long, Light-stimulated protein movement in rod photoreceptor cells of the rat retina (225) 127
Picard, J. see J. Gardette (225) 178
Pick, U. see K. Gounaris (211) 94
Pieck, A.C.M., A.A.M. Rijken and F. Wanka, Nuclear matrix and chromosome scaffold preparations of in vitro cultured bovine liver cells have two proteins in common (212) 276
Piefke, J., T. Arad, H.S. Gewitz, A. Yonath and H.G. Wittmann, The growth of ordered two-dimensional sheets of 70 S ribosomes from *Bacillus stearothermophilus* (209) 104
Piiper, A. see V. Henne (218) 153
Pin, S., R. Cortes and B. Alpert, XANES spectroscopy of carp hemoglobin-iron in correlation with the affinity changes of the protein for ligand (208) 325
Piña, B. and P. Suau, Changes in the proportions of histone H1° subtypes in brain cortical neurons (210) 161
Pinna, L.A. see F. Meggio (215) 241
Pinna, L.A. see P. Agostinis (224) 385
Piper, P.W., A. Lockheart and J. Bellatin, The effects of protein synthesis inhibition, and of mutations *rna1.1* and *rna82.1*, on the synthesis of small RNAs in yeast (214) 143
Piper, P.W., B. Curran, W. Davies, K. Hirst and K. Seward, *Saccharomyces cerevisiae* mRNA populations of different intrinsic stability in unstressed and heat shocked cells display almost constant m^7GpppA :m^7GpppG 5'-cap structure ratios (220) 177
Piriou, B. see J. Dexpert (225) 223
Pirozhkov, S.V. see V.D. Antonenkov (224) 357
Plat, H. see R. Wever (216) 1
Plattner, H. see B. Stecher (223) 25
Plattner, J.J. see H.L. Sham (220) 299
Plijter, J.J. see R.V. Danielius (213) 241
Plisetskaya, E.M. see T.P. Mommsen (219) 227
Pliszka, B., Crosslinking of trypsin digested acto-heavy meromyosin as a probe of the affinity of the two myosin heads to actin (212) 254
Plocke, D.J. see S.G. Whiteside (226) 250
Plomp, P.J.A.M. see X.M. Leverve (219) 455
Ploos van Amstel, H.K., A.L. van der Zanden, P.H. Reitsma and R.M. Bertina, Human protein S cDNA encodes Phe-16 and Tyr-222 in consensus sequences for the post-translational processing (222) 186
Pochon, F., M. Tourbez, V. Favaudon and E. Delain, Covalent and non-covalent interaction of chymotrypsin with α_2-macroglobulin (217) 101
Pochon, F. see J. Dexpert (225) 223
Podust, V.N. see O.I. Lavrik (216) 225
Podust, V.N. see S.V. Doronin (216) 221
Pogády, J. see Ľ. Varečka (225) 173
Poggioli, J. see D. Renard (217) 117
Poglazov, B.F. see D.I. Levitsky (221) 77
Polgár, L., The mechanism of action of aspartic proteases involves 'push-pull' catalysis (219) 1Ponnambalam, S., A. Spassky and S. Busby, Studies with the *Escherichia coli* galactose operon regulatory region carrying a point mutation that simultaneously inactivates the two overlapping promoters: Interactions with RNA polymerase and the cyclic AMP receptor protein (219) 189
Politi, L. see R. Scandurra (212) 79
Pollard, H.B. see E. Rojas (217) 25
Pollet, R.J. see D.R. Cooper (214) 122
Pollock, W.K., S.O. Sage and T.J. Rink, Stimulation of Ca^2 efflux from fura-2-loaded platelets activated by thrombin or phorbol myristate acetate (210) 132
Polonovski, J. see C. Mazière (218) 243
Polya, G.M. see A. Lucantoni (221) 33
Pon, C.L. see M. Lammi (215) 115
Ponnambalam, S. and S. Busby, RNA polymerase molecules initiating transcription at tandem promoters can collide and cause premature transcription termination (212) 21
Pontet, M. see R. Buchta (211) 165
Poole, P.L. see D.J. Barlow (213) 423
Poole, R.K. and H.D. Williams, Proposal that the function of the membrane-bound cytochrome a_1-like haemoprotein (cytochrome *b*-595) in *Escherichia coli* is a direct electron donation to cytochrome *d* (217) 49
Portier, M.-M. see A. Lomri (222) 311
Portis, A.R. jr see M.E. Salvucci (221) 215
Portugal, J. and M.J. Waring, Hydroxyl radical footprinting of the sequence-selective binding of netropsin and distamycin to DNA (225) 195

Pöschl, E., I. Lindley, E. Hofer, J.-M. Seifert, W. Brunowsky and J. Besemer, The structure of procalcitonin of the salmon as deduced from its cDNA sequence (226) 96
Possani, L.D. see H.H. Valdivia (226) 280
Potts, J.R. see P.W. Kuchel (219) 5
Pouyet, J. see E. Bertrand-Burggraf (215) 83
Pouysségur, J. see I. Magnaldo (210) 6
Prangishvili, D.A. see R.P. Vashakidze (216) 217
Praškevičius, A. see B. Kholodenko (223) 247
Premecz, G., A. Markovits, G. Bagi, T. Farkas and I. Földes, Inositol phospholipid turnover and protein kinase C translocation are stimulated by poly(I) · poly(C) in human amnion cells (UAC) (226) 13
Premereur, N. see C. Van den Branden (222) 21
Prentki, M., M.C. Glennon, J.-F. Geschwind, F.M. Matschinsky and B.E. Corkey, Cyclic AMP raises cytosolic Ca^{2+} and promotes Ca^{2+} influx in a clonal pancreatic β-cell line (HIT T-15) (220) 103
Preobrazhensky, A.A., A.I. Rodionova, I.N. Trakht and V.S. Rukosuev, Monoclonal antibodies against chordin: Use in structural and immunohistochemical studies (224) 23
Preston, C., J.G. Altin, F.L. Bygrave and C. Critchley, Light induced calcium binding to mangrove PS II particles (210) 27
Preston, C.M. see V.D. Appanna (215) 79
Preston, J. see E. Brown (218) 113
Preston, J. see M. Oetting (208) 99
Pribilla, I. see F. Hucho (211) 207
Prickril, B. see L.C. Sieker (208) 73
Prieto, J.C. see M.J. Carmena (218) 73
Prieto, J. see M. Patarroyo (210) 127
Prieur, B. see A.J. Aarsman (219) 176
Prieur, B. see B. Rothhut (219) 169
Prince, R.C. see W.J. Jackson (215) 171
Printseva, O.Yu. see M.A. Chernousov (217) 124
Prior, B. see M. O'Hare (216) 73
Prochaska, L.J. see P.S. Fink (214) 75
Prochaska, L.J. see P.S. Fink (220) 263
Prockop, D.J. see R. Soininen (225) 188
Prohaska, R. see M. Krahmer (226) 105
Pronin, A.N. see Yu.A. Ovchinnikov (226) 91
Proudfoot, K. see K.H. Cheeseman (209) 191
Ptitsyn, O.B. see G.V. Semisotnov (224) 9
Ptitsyn, O.B. see R.I. Gilmanshin (223) 327
Puget, K. see M.E. Schininà (221) 87
Pullman, A. see Y. Trudelle (216) 11
Pullman, B. see K.-X. Chen (224) 361
Püschel, G.P., A. Nath and K. Jungermann, Increase of urate formation by stimulation of sympathetic hepatic nerves, circulating noradrenaline and glucagon in the perfused rat liver (219) 145
Puustinen, T., M.M. Scheffer and B. Samuelsson, Endogenously generated 5-hydroperoxyeicosatetraenoic acid is the preferred substrate for human leukocyte leukotriene A_4 synthase activity (217) 265
Pyne, N.J. see F. Irvine (208) 455
Pyun, H.Y. see B.F. Dickey (219) 289

Q

Quash, G. see J.G. Delcros (220) 236
Que, B.G. and P.H. Petra, Characterization of a cDNA coding for sex steroid-binding protein of human plasma (219) 405
Quennemet, J. see O. Dogbo (210) 211
Quinlan, J.G. see P.C. Moorhouse (213) 23
Qureshi, N. see C.A. Ellis (218) 238

R

Rabasseda, X., C. Solsona, J. Marsal, G. Egea and B. Bizzini, ATP release from pure cholinergic synaptosomes is not blocked by tetanus toxin (213) 337
Rack, M., D. Richter and N. Rubly, Purification and characterization of a β-toxin from the venom of the African scorpion *Leiurus quinquestriatus* (214) 163
Råden, B. see M. Forsgren (213) 254
Radunović, L. see S. Suzić (216) 287
Ragab-Thomas, J. see J. Fauvel (221) 397
Ragan, C.I. see F.G.P. Earley (219) 108
Rall, T. and B.A. Harris, Identification of the lesion in the stimulatory GTP-binding protein of the uncoupled S49 lymphoma (224) 365
Ramlal, T. see A. Klip (224) 224
Ramos, J.L., M. Gonzalez-Carrero and K.N. Timmis, Broad-host range expression vectors containing manipulated *meta*-cleavage pathway regulatory elements of the TOL plasmid (226) 241
Ramstein, J. see B. Hartmann (225) 11
Rance, M. see W.J. Chazin (222) 109
Randall, R.W., J.E. Tateson, J. Dawson and L.G. Garland, A commentary on the inhibition by retinoids of leukotriene B_4 production in leukocytes (214) 167
Randle, P.J. see M. Beggs (215) 13
Ranjeva, R. see M. Dillenschneider (208) 413
Ranu, R.S., Regulation of protein synthesis in rabbit reticulocyte lysates: Thiophosphorylation of initiation factor eIF-2 by heme-regulated protein kinase (208) 117
Ranu, R.S. see R.C. Roberts (209) 162
Rao, M.V.R. see K. Gopalakrishna (215) 95
Rapier, C., S. Wonnacott, G.G. Lunt and E.X. Albuquerque, The neurotoxin histrionicotoxin interacts with the putative ion channel of the nicotinic acetylcholine receptors in the central nervous system (212) 292
Rapoport, B. see G. Damante (225) 264
Rapoport, B. see R.P. Magnusson (208) 391
Rapoport, H. see D.R.E. Macallan (226) 357
Rapoport, H. see Y. Aracava (222) 63
Rapoport, S., Misunderstanding or misrepresentation?: 'Reticulocyte lipoxygenase, ingensin, and ATP-dependent proteolysis' by Shoichi Ishiura, Tanihiro Yoshimoto and Claude A. Villee [(1986) FEBS Lett. 201, 87–93] (215) 193
Rapoport, S.M. see M.M. Kostić (217) 163
Rappuoli, R. see E. Papini (215) 73
Rappuoli, R. see M. Bigio (218) 271
Rastad, J. see P. Nygren (213) 195
Ratti, G. see M. Bigio (218) 271
Rattigan, S. see E.A. Richter (217) 232
Ratz, G.P. see J.E. Celis (223) 237
Rauvala, H. see J. Laitinen (217) 94
Ray, R. and U. Ganguly, The protein receptor for cholerabacteriophage φ149 (213) 81
Razjivin, A.P. see A.P. Mineyev (223) 187
Reale, M. see P. Conti (225) 103
Rebizova, T.V. see L.P. Sashchenko (226) 261
Reboud, A.-M. see A. Paleologue (208) 373

Reboud, A.-M. see J.-P. Lavergne (216) 83
Reboud, J.P. see A. Paleologue (208) 373
Reboud, J.-P. see J.-P. Lavergne (216) 83
Reece, D. see Y.C. Lee (220) 243
Reeck, G.R. see C.-S. Wang (222) 135
Rees Smith, B. see J. Furmaniak (215) 316
Reglinski, J., W.E. Smith, C.J. Suckling, M. Al-Kabban, I.D. Watson and M.J. Stewart, A ^{1}H spin echo NMR study of the HeLa tumour cell (214) 351
Regoeczi, E. and P.A. Chindemi, Metabolic stability of the fucose in rat transferrin (222) 271
Reinke, H. see T. Jansen (216) 234
Reitsma, P.H. see H.K. Ploos van Amstel (222) 186
Rempel, C.A. see H.L. Sham (220) 299
Renard, D. and J. Poggioli, Does the inositol tris/tetrakisphosphate pathway exist in rat heart? (217) 117
Renganathan, V. see K. Miki (210) 199
Renge, I. see K. Mauring (223) 165
Rennie, M.J. see P.A. MacLennan (215) 187
Rennie, M.J. see P.M. Taylor (221) 370
Repke, H. and M. Bienert, Mast cell activation – a receptor-independent mode of substance P action? (221) 236
Reshef, L. see H. Cohen (223) 347
Revazova, E.S. see L.P. Sashchenko (226) 261
Reynolds, P.E. see D.M. O'Hara (212) 237
Rhodes, C.J., C.A. Lucas and P.A. Halban, Glucose stimulates the biosynthesis of rat I and II insulin to an equal extent in isolated pancreatic islets (215) 179
Ribadeau-Dumas, B. see C. Carles (212) 163
Ribbons, D.W. see A.E.G. Cass (220) 353
Riccio, A. see P.A. Andreasen (209) 213
Rice, K.C. see B. DeCosta (223) 335
Rice, K.C. see D.J.J. Carr (224) 272
Richard, G. see F. Baklouti (223) 59
Richardson, M., A. Yarwood and P. Rougé, The amino acid sequence of an atypical single-chain lectin from seeds of *Lathyrus sphaericus* (Retz) (216) 145
Richter, D. see M. Rack (214) 163
Richter, D. see S. Fehr (210) 45
Richter, E.A., P.J.F. Cleland, S. Rattigan and M.G. Clark, Contraction-associated translocation of protein kinase C in rat skeletal muscle (217) 232
Rico, M. see M.A. Jiménez (221) 320
Rider, M.A. see D.F. Morrison (222) 266
Rider, M.H. see M.I. Darville (224) 317
Ried, G. and U. Henning, A unique amino acid substitution in the outer membrane protein OmpA causes conjugation deficiency in *Escherichia coli* K-12 (223) 387
Riede, I., H. Schwarz and F. Jähnig, Predicted structure of tail-fiber proteins of T-even type phages (215) 145
Rieker, J.P. and J.H. Collins, Phosphorylation of brush border myosin by brush border calmodulin-dependent myosin heavy and light chain kinases (223) 262
Rieker, J.P., H. Swanljung-Collins, J. Montibeller and J.H. Collins, Brush border myosin heavy chain phosphorylation is regulated by calcium and calmodulin (212) 154
Riemens, T. see W. Siffert (212) 123
Rienits, K.G. see U. Schreiber (211) 99
Ries, B. see V. Mrša (217) 174
Riethman, H.C., T.P. Mawhinney and L.A. Sherman, Phycobilisome-associated glycoproteins in the cyanobacterium *Anacystis nidulans* R 2 (215) 209
Rigor, B.M. see A. Schurr (224) 4
Rigoulet, M. see J.-J. Bessoule (214) 158
Rijken, A.A.M. see A.C.M. Pieck (212) 276
Rinaldi, A. see R.L.P. Adams (215) 266
Ringer, D.P., B.A. Howell, J.L. Etheredge, J.A. Clouse and D.E. Kizer, Assessment of salvage pathways utilized for incorporation of exogenous pyrimidine nucleosides into DNA of guinea pig lymphocytes stimulated by Con A (224) 59
Rink, T.J. see A.W.M. Simpson (222) 144
Rink, T.J. see W.K. Pollock (210) 132
Riou, G. see J.-F. Riou (213) 304
Riou, J.-F., M. Dollet, J.-C. Ahomadegbe, D. Coulaud and G. Riou, Characterization of *Phytomonas* sp. kinetoplast DNA: A plant pathogenic trypanosomal species (213) 304
Rizk, M. see C.P. Schelling (214) 21
Roach, P.D., A.M. Kambouris, R.P. Trimble and D.L. Topping and P.J. Nestel, The effects of dietary fish oil on hepatic high density and low density lipoprotein receptor activities in the rat (222) 159
Robert-Nicoud, M. see D.M. Soumpasis (213) 341
Roberts, G.C.K. see B. Birdsall (217) 106
Roberts, L.M. see M. O'Hare (216) 73
Roberts, R.C. and R.S. Ranu, Regulation of protein synthesis in rabbit reticulocyte lysates: Requirement of initiation factor eIF-2 holoprotein for substrate specificity of heme-regulated protein kinase (209) 162
Robertson, L.W. see Z. Amelizad (220) 231
Robillard, G.T., J.M. Schaaf and A.W. Teelken, Dithiols and monothiols are linked with GABA transport in membrane vesicles of rat brain synaptosomes (224) 391
Robins, R.K. see D. Katsaros (223) 97
Robins, R.K. see T. Clair (224) 377
Robinson, A. and B. Austen, GTP-dependent ADP-ribosylation of a 22 kDa protein in the endoplasmic reticulum membrane (218) 63
Robinson, D. see F.-Z. Chung (211) 200
Robinson, J.H. see I. Dodd (209) 13
Robinson, P.A., D. Wion and B.H. Anderton, Isolation of a cDNA for the rat heavy neurofilament polypeptide (NF-H) (209) 203
Robinson, P.A. see J.-P. Brion (226) 28
Robson, B. see C.A. Morrison (214) 65
Robson, B. see R.V. Fishleigh (214) 219
Rocchi, M. see M. Rugolo (212) 313
Roch, A.M. see J.G. Delcros (220) 236
Rochat, H. see J. Fauvel (216) 45
Rochat, H. see J. Fauvel (221) 397
Roche, E., F. Aniento, E. Knecht and S. Grisolía, 2,3-Bisphosphoglycerate inhibits ATP-stimulated proteolysis (221) 231
Roche, E. see J.L. Vargas (224) 182
Rocher-Dufour, S. see M. Obara (213) 261
Rodionov, M.A., S.G. Galaktionov and A.A. Akhrem, Prediction of exposure degree diagram and sites of limited proteolysis in globular proteins as an approach to computer-aided design of protein bioregulators with prolonged action (223) 402
Rodionova, A.I. see A.A. Preobrazhensky (224) 23
Rodionova, N.A. see G.V. Semisotnov (224) 9
Rodrig, N., T. Osanai, M. Iwamori and Y. Nagai, Gangliosides stimulate dome formation in cultured canine kidney epithelial cell line (MDCK) (221) 315
Roels, F. see C. Van den Branden (222) 21
Roersma, E.S. see C.E.M. Voorter (221) 249
Roevens, P. see D. de Chaffoy de Courcelles (219) 283
Rogard, M. see B. de Foresta (216) 107
Rogers, L.J. see P. Hilditch (208) 313
Rogez, D., R. Cerf, R. Andrianjara, S.-T. Salehi and H. Fouladgar, Ultrasonic studies of proton-transfer reactions at the catalytic site of α-chymotrypsin (219) 22

Rögner, M., J.P. Dekker, E.J. Boekema and H.T. Witt, Size, shape and mass of the oxygen-evolving photosystem II complex from the thermophilic cyanobacterium *Synechococcus* sp. (219) 207
Rögner, M. see E.J. Boekema (217) 283
Rögner, M. see I. Witt (221) 260
Rogozin, I.B. see N.A. Kolchanov (214) 87
Röhm, K.-H. see J. Henseling (219) 27
Rojas, E. and H.B. Pollard, Membrane capacity measurements suggest a calcium-dependent insertion of synexin into phosphatidylserine bilayers (217) 25
Rojas, E. see R.M. Santos (220) 342
Rojo, F. see J. Berenguer (224) 401
Romano, V. see D. Fürst (224) 49
Romanova, E.A. see V.G. Metelev (226) 232
Romanowska, A. see E. Romanowska (211) 175
Romanowska, E., A. Romanowska, J. Dabrowski and M. Hauck, Structure determination of the O-specific polysaccharides from *Citrobacter* O4- and O27-lipopolysaccharides by methylation analysis and one- and two-dimensional ^{1}H-NMR spectroscopy (211) 175
Römbach, A. see E. Kojro (212) 83
Romeo, G. see D. Fürst (224) 49
Romeo, G. see M. Rugolo (212) 313
Romhányi, T. see L. Buday (223) 15
Rommerts, F.F.G. see L.E. Mallea (218) 143
Roncero, C., J.M. Estrela and M. Benito, Adenine nucleotide compartmentation in foetal rat hepatocytes: Effects of atractyloside, oligomycin, calcium ionophore and adrenergic agonists (208) 105
Rondinelli, E., R.S. de Moura-Neto, R. Silva, C.M. de Almeida Soares, J.F. Carvalho and F. Torres de Castro, Control of tubulin gene expression during metacyclogenesis of *Trypanosoma cruzi* (208) 379
Roos, B.A. see J.W.M. Höppener (215) 122
Roques, B.P. see J.-M. Zajac (216) 118
Roques, V. see J. Fauvel (216) 45
Roques, V. see J. Fauvel (221) 397
Rose, P.M. see S.M. Penningroth (222) 204
Rosemeyer, M.A. see A.A. Haritos (218) 107
Rosenbusch, J. see B. Dargent (220) 136
Rösener, S., G.S. Chhatwal and K. Aktories, Botulinum ADP-ribosyltransferase C3 but not botulinum neurotoxins C1 and D ADP-ribosylates low molecular mass GTP-binding proteins (224) 38
Rosenheck, K. see P.I. Lelkes (208) 357
Rosenthal, I., C.M. Krishna, G.C. Yang, T. Kondo and P. Riesz, A new approach for EPR detection of hydroxyl radicals by reaction with sterically hindered cyclic amines and oxygen (222) 75
Rosenthal, W., T. Binder and G. Schultz, NADP efficiently inhibits endogenous but not pertussis toxin-catalyzed covalent modification of membrane proteins incubated with NAD (211) 137
Rosie, R. see A.J. Harmar (208) 67
Roskam, W. see P. Ferrara (226) 47
Rospendowski, B.N. see K. Kelly (222) 120
Rossi, R. see M. Bigio (218) 271
Rossier, J. see B. Lambolez (219) 301
Rossier, J. see N.G. Seidah (211) 144
Rossiter, J.T. see A.E.G. Cass (220) 353
Rostapshov, V.M. see E.D. Sverdlov (212) 233
Rosztóczy, I., M. Papós and K. Megyeri, Different interferon-producing capacities of L929 cell sublines and the enhancement of interferon production by priming are controlled pretranslationally (208) 56
Rotberg, J. see E. Brown (218) 113
Roth, B., B. Schwendimann, G.J. Hughes, S.J. Tzartos and T. Barkas, A modified nicotinic acetylcholine receptor lacking the 'ion channel amphipathic helices' (221) 172
Rother, C. see T. Jansen (216) 234
Rothhut, B., C. Comera, B. Prieur, M. Errasfa, G. Minassian and F. Russo-Marie, Purification and characterization of a 32-kDa phospholipase A_2 inhibitory protein (lipocortin) from human peripheral blood mononuclear cells (219) 169
Rothhut, B. see A.J. Aarsman (219) 176
Rothman, R.B. see B. DeCosta (223) 335
Rothschild, K.J. see H. Marrero (223) 289
Rotstein, O.D., K. Houston and S. Grinstein, Control of cytoplasmic pH by Na^+/H^+ exchange in rat peritoneal macrophages activated with phorbol ester (215) 223
Rougé, P. see M. Richardson (216) 145
Rougeon, F. see M. Ekker (222) 337
Roughley, B.S. see H.K. Mangold (220) 220
Rouillé, Y. see M.T. Chauvet (210) 40
Rouimi, P., J. Bonicel, M. Rovery and A. De Caro, Cleavage of the Arg–Ile bond in the native polypeptide chain of human pancreatic stone protein (216) 195
Rouot, B. see M. Toutant (215) 339
Rouot, B. see M. Toutant (222) 51
Rousseau, G.G. see M.I. Darville (224) 317
Rousset, M., G. Trugnan, J.-L. Brun and A. Zweibaum, Inhibition of the post-translational processing of microvillar hydrolases is associated with a specific decreased expression of sucrase-isomaltase and an increased turnover of glucose in Caco-2 cells treated with monensin (208) 34
Rouyer-Fessard, C. see J.-C. Bozou (211) 151
Rovery, M. see P. Rouimi (216) 195
Roy, P. see Nisan Bhattacharyya (208) 386
Royyuru, A.K. see K. Gopalakrishna (215) 95
Rozovskaja, I.A. see L.B. Margolis (220) 288
Rubin, A.B. see V.Z. Paschenko (214) 28
Rubira, M.R. see R.J. Simpson (224) 128
Rubly, N. see M. Rack (214) 163
Rubtsov, N.B. see N.M. Matveeva (217) 42
Rüdiger, W. see R. Grimm (225) 215
Rudy, B. see A. Wolf (212) 203
Rüegg, J.C. see A. Takai (217) 81
Ruff, M.R., B.M. Martin, E.I. Ginns, W.L. Farrar and C.B. Pert, CD4 receptor binding peptides that block HIV infectivity cause human monocyte chemotaxis: Relationship to vasoactive intestinal polypeptide (211) 17
Ruggiero, M. see G. Laffi (220) 217
Rugolo, M., M. Rocchi, G. Lenaz and G. Romeo, Increased chloride efflux in fibroblasts from X-linked muscular dystrophies and clones from Duchenne carriers (212) 313
Ruiz, O. see R.B. Frydman (219) 380
Rukosuev, V.S. see A.A. Preobrazhensky (224) 23
Rümbeli, R., F. Suter, M. Wirth, W. Sidler and H. Zuber, γ-*N*-Methylasparagine in phycobiliproteins from the cyanobacteria *Mastigocladus laminosus* and *Calothrix* (221) 1
Runnebaum, B. see L. Kiesel (217) 85
Russo-Marie, F. see A.J. Aarsman (219) 176
Russo-Marie, F. see B. Rothhut (219) 169
Ryabova, L.A., O.M. Selivanova, V.I. Baranov, V.D. Vasiliev and A.S. Spirin, Does the channel for nascent peptide exist inside the ribosome?: Immune electron microscopy study (226) 255
Ryabushenko, I.L. see V.G. Metelev (226) 232
Ryazanov, A.G., Ca^{2+}/calmodulin-dependent phosphorylation of elongation factor 2 (214) 331
Ryazantsev, S.N. see S.D. Trakhanov (220) 319
Ryba, N.J.P. see R. Wagner (221) 253
Rybkine, T. see J. Masliah (222) 11
Ryskov, A.P. see S.A. Limborska (212) 208

S

Saarma, M. see H.-J. Kärgel (220) 126
Sabbatini, G.P. and C. von Holt, Carrier-free 8-azidoadenosine 5′-[γ-^{32}P]triphosphate (224) 117
Sacchetta, P. see D. Di Cola (210) 81
Sachar, R.C. see R. Kaul (209) 63
Sachse, G. see B. Appelhans (224) 14
Sadée, W. see F.-J. Klinz (224) 43
Sadler, P.J. see J.D. Bell (215) 311
Sadler, P.J. see J.D. Bell (219) 239
Saenger, W. see E.J. Boekema (217) 283
Saenger, W. see I. Witt (221) 260
Sage, S.O. see W.K. Pollock (210) 132
Sainte-Marie, J. see M. Vidal (216) 159
Sakaguchi, K. see Y. Shimohigashi (222) 251
Sakai, H. see M. Hoshi (217) 237
Sakai, H. see S. Maekawa (221) 68
Sakai, H. see Y. Ohta (208) 423
Sakai, H. see Y. Ohta (222) 305
Sakai, K. see S. Dewhurst (213) 138
Sakai, N., T. Umeda, H. Suzuki, Y. Ishimatsu and M. Shikita, Macrophage colony-stimulating factor purified from normal human urine: Amino-terminal sequence and amino acid composition (222) 341
Sakakibara, S. see Y. Hirata (219) 369
Sakamoto, C. see M. Nagao (214) 107
Sakamoto, W., F. Satoh, K. Gotoh and S. Uehara, Ile-Ser-bradykinin (T-kinin) and Met-Ile-Ser-bradykinin (Met-T-kinin) are released from T-kininogen by an acid proteinase of granulomatous tissues in rats (219) 437
Sakihama, N. see A. Hiwatashi (209) 311
Sakihama, T. see E. Takano (208) 199
Sakiyama, S. see T. Hiwasa (211) 23
Saklatvala, J. see T.A. Bird (225) 21
Sakmann, B. see V. Witzemann (223) 104
Saks, V.A. see V.V. Kupriyanov (208) 89
Sakurada, K. see Y. Uehara (208) 352
Sakuragawa, N. see R. Yamagishi (225) 109
Salehi, S.-T. see D. Rogez (219) 22
Sales, N. see J.-M. Zajac (216) 118
Sallenave, J.-M. and R. Bellot, Evidence of an α_2-macroglobulin-like molecule in plasma of *Salamandra salamandra:* Structural and functional similarity with human α_2-macroglobulin (219) 37
Salles, J.-P. see J. Fauvel (216) 45
Salnikow, J. see P. Fromme (218) 27
Salomon, Y. see N.B. Garty (218) 148
Salonen, E.-M. and A. Vaheri, Fibronectin-binding 36 kDa protein in human fibroblasts (221) 381
Salter, A.M., S.C. Fisher and D.N. Brindley, Binding of low-density lipoprotein to monolayer cultures of rat hepatocytes is increased by insulin and decreased by dexamethasone (220) 159
Saluja, D. see R. Kaul (209) 63
Salvioli, R. see A.M. Vaccaro (216) 190
Salvucci, M.E., A.R. Portis jr, U. Heber and W.L. Ogren, Stimulation of thylakoid energization and ribulose-bisphosphate carboxylase/ oxygenase activation in *Arabidopsis* leaves by methyl viologen (221) 215
Samain, E., D.S. Patil, D.V. DerVartanian, G. Albagnac and J. LeGall, Isolation of succinate dehydrogenase from *Desulfobulbus elongatus,* a propionate oxidizing, sulfate reducing bacterium (216) 140
Sample, C.E. and D.F. Steiner, Tissue-specific binding of a nuclear factor to the insulin gene promoter (222) 332
Samuel, K.P., A. Seth, A. Konopka, J.A.Lautenberger and T.S. Papas, The 3′-orf protein of human immunodeficiency virus shows structural homology with the phosphorylation domain of human interleukin-2 receptor and the ATP-binding site of the protein kinase family (218) 81
Samuels, D.S., T. Tojo, M. Homma and N. Shimizu, Inhibition of topoisomerase I by antibodies in sera from scleroderma patients (209) 231
Samuelsson, B. see C.N. Serhan (217) 242
Samuelsson, B. see T. Miyamoto (216) 123
Samuelsson, B. see T. Puustinen (217) 265
Samuelsson, G. see L.-G. Sundblad (209) 28
Sanchez, A. see J.M. Collazos (215) 183
Sánchez, J.A. see A. Guerrero (220) 295
Sanchez, J., B.E. Uhlin, T. Grundström, J. Holmgren and T.R. Hirst, Immunoactive chimeric ST-LT enterotoxins of *Escherichia coli* generated by in vitro gene fusion (208) 194
Sancho, J.I. see M.J. Carmena (218) 73
Sandelius, A.S. see P.-M. Melin (223) 87
Sanders, J.K.M. see R.P. Mason (216) 4
Sandford, C.A. see T. Capiod (217) 247
Santoro, J. see M.A. Jiménez (221) 320
Santoro, L. see R. Scandurra (212) 79
Santos, H. and D.L. Turner, Proton NMR studies of horse ferricytochrome *c*: Completion of the assignment of the well resolved hyperfine shifted resonances (226) 179
Santos, R.M. and E. Rojas, Evidence for modulation of cell-to-cell electrical coupling by cAMP in mouse islets of Langerhans (220) 342
Sanwal, B.D. see G.A. Cates (218) 195
Sapotsky, M.V. see Yu.F. Drygin (215) 247
Saraste, M., CO III gene in bacteria: 'Homology between bacterial DNA and bovine mitochondrial DNA encoding cytochrome *c* oxidase subunit III' by Pamela S. Fink, Tracy Whitford, Michael Leffak and Lawrence J. Prochaska [(1987) FEBS Lett. 214, 75–80] (220) 262
Saraste, M. see V.-M. Wasenius (221) 73
Sarkadi, B. see L. Hunyady (225) 72
Sartorelli, A.C. see P.G. Penketh (221) 427
Sartorio, R. see P.A. Andreasen (209) 213
Sasada, R. see T. Kurokawa (213) 189
Sasakawa, N., T. Nakaki, S. Yamamoto and R. Kato, Inositol trisphosphate accumulation by high K^+ stimulation in cultured adrenal chromaffin cells (223) 413
Sasaki, T. and H. Hasegawa-Sasaki, Activation of polyphosphoinositide phospholipase C by guanosine 5′-*O*-(3-thio)triphosphate and fluoroaluminate in membranes prepared from a human T cell leukemia line, JURKAT (218) 87
Sashchenko, L.P., N.V. Gnuchev, M.A. Kirillova, T.I. Lukjanova, T.V. Rebizova, E.S. Revazova and E.M. Lukanidin, Separation of the pore-forming and cytotoxic activities from natural killer cell cytotoxic factor (226) 261
Sasisekharan, V. see P. Parrack (212) 297
Sassolas, G. see J. Champier (212) 220
Satav, J.G. see V.N. Pandey (213) 204
Sato, A. see M. Maki (223) 174
Sato, E.F., Y.M. Morimoto, T. Matsuno, M. Miyahara and K. Utsumi, Neutrophil specific 33 kDa protein: its Ca^{2+}- and phospholipid-dependent intracellular translocation (214) 181
Sato, M., T. Nakamura and J. Koyama, Different abilities of two distinct Fcγ receptors on guinea pig polymorphonuclear leukocytes to trigger the arachidonic acid metabolic cascade (224) 29
Satoh, F. see W. Sakamoto (219) 437
Satoh, J. see A. Kawamori (217) 134
Satoh, K. see A. Kawamori (217) 134

Satoh, K. see I. Enami (226) 161
Satoh, K. see R.V. Danielius (213) 241
Satoh, K. see Y. Takahashi (208) 347
Satoh, K., Y. Fujii, T. Aoshima and T. Tado, Immunological identification of the polypeptide bands in the SDS-polyacrylamide gel electrophoresis of photosystem II preparations (216) 7
Satoyoshi, E. see Y. Furukawa (208) 258
Saul, S.J. and M. Sugumaran, Protease inhibitor controls prophenoloxidase activation in *Manduca sexta* (208) 113
Saunders, N.R. see D.L. Christie (214) 45
Sauvageot, M. see L.-H. Tessier (208) 183
Savart, M., M. Belamri, V. Pallet and A. Ducastaing, Association of calpains 1 and 2 with protein kinase C activities (216) 22
Sawai, T. see A. Yamaguchi (208) 43
Sawai, T. see A. Yamaguchi (218) 126
Sawano, Y. see H. Fukuzawa (220) 61
Sawyer, W.H. see J.R. Wardlaw (223) 20
Scandurra, R., L. Politi, L. Santoro and V. Consalvi, Covalently bound pyruvate in phosphopantothenoylcysteine decarboxylase from horse liver (212) 79
Scarfò, R. see F. Guerrieri (213) 67
Scarpa, A. see E.C. Wiener (224) 33
Scarpa, A. see M.B. De Young (223) 53
Schaaf, J.M. see G.T. Robillard (224) 391
Schächtele, C. see C. Bruns (221) 23
Schadewaldt, P., N.J. Stapper and W. Staib, Effect of adrenergic agonists on phosphoinositide breakdown in rat skeletal muscle preparations (217) 45
Schaechter, L.E. see K.R. Parker (211) 35
Schägger, H., U. Borchart, W. Machleidt, T.A. Link and G. Von Jagow, Isolation and amino acid sequence of the 'Rieske' iron sulfur protein of beef heart ubiquinol:cytochrome *c* reductase (219) 161
Schairer, H.U. see H.A. Weich (213) 89
Schams, D. see S. Fehr (210) 45
Scheffer, M.M. see T. Puustinen (217) 265
Scheid, P. see W. Siffert (212) 123
Schell, J. see T. Etzold (219) 343
Schelling, C.P., L. Didierjean, M. Rizk, J.H. Pavlovitch, T. Takagi and C.W. Heizmann, Calcium-binding proteins in rat skin (214) 21
Schering, B. see J. Vandekerckhove (225) 48
Scherman, D. see B. Gasnier (222) 215
Scherrer, K. see J. Moreau (221) 3
Schewe, T. see H. Kühn (208) 248
Schiavo, G. see E. Papini (215) 73
Schiefer, H.G. see U. Schummer (224) 79
Schifferli, J.A., G. Hauptmann and J.-P. Paccaud, Complement-mediated adherence of immune complexes to human erythrocytes: Difference in the requirements for C4A and C4B (213) 415
Schininà, M.E., L. Maffey, D. Barra, F. Bossa, K. Puget and A.M. Michelson, The primary structure of iron superoxide dismutase from *Escherichia coli* (221) 87
Schlegel, R.A. see P. Williamson (219) 316
Schleicher, M. see A. Noegel (221) 391
Schlipfenbacher, R., S. Wenzl, F. Lottspeich and M. Sumper, An extremely hydroxyproline-rich glycoprotein is expressed in inverting *Volvox* embryos (209) 57
Schlodder, E. see S. Gerken (223) 376
Schmeissner, U. see H.R. MacDonald (209) 295
Schmidt, H.W.H. see S.D. Haemmerli (220) 149
Schmitt, G. see J. Borg (213) 406
Schnarr, M. see E. Bertrand-Burggraf (215) 83
Schneider, D.L. see F. Mollinedo (217) 158
Schneider, E. see P.P. Kamoun (226 (1988) 285
Schnell, K. see G. Eberl (222) 349
Schoemaker, H.E. see S.D. Haemmerli (220) 149
Schoepfer, R., M. Luther and J. Lindstrom, The human medulloblastoma cell line TE671 expresses a muscle-like acetylcholine receptor: Cloning of the α-subunit cDNA (226) 235
Schofield, P.N. see P. Hammonds (213) 149
Schofield, P.N. see P. Hammonds (223) 131
Schön, R. see R. Grosse (210) 123
Schönert, H. see M. Heinrichs (223) 255
Schreiber, U. and K.G. Rienits, ATP-induced photochemical quenching of variable chlorophyll fluorescence (211) 99
Schreier, P.H. see T. Etzold (219) 343
Schreier, T., K.H. Winterhalter and B. Trüeb, The tissue form of chicken type VI collagen (213) 319Segrest, J.P. see G. von Heijne (213) 238
Schröder, W. see G. Theiss (218) 159
Schulick, R. see K.A. Jacobson (225) 97
Schulte-Frohlinde, D. see H. Wefers (211) 49
Schultz, G. see W. Rosenthal (211) 137
Schulz, G.E. see D. Dreusicke (208) 301
Schumacher, L. see J. Hoppe (223) 243
Schummer, U. and H.G. Schiefer, Transmembrane proton-motive potential of *Spiroplasma floricola* (224) 79
Schurr, A. and B.M. Rigor, The mechanism of neuronal resistance and adaptation to hypoxia (224) 4
Schuster, G. see A. Gal (221) 205
Schuster, G. see T.G. Dunahay (215) 25
Schwartz, J.-C. see F. Vargas (211) 234
Schwartz, T.W., S.P. Sheikh and M.M.T. O'Hare, Receptors on phaeochromocytoma cells for two members of the PP-fold family – NPY and PP (225) 209
Schwartz, T.W. see E. Boel (219) 181
Schwartz, T.W. see J.J. Holst (211) 169
Schwarz, H. see I. Riede (215) 145
Schwarz, M. see B.A. Hemmings (209) 219
Schwarz-Magdolen, U., I. Oberbäumer and K. Kühn, cDNA and protein sequence of the NC1 domain of the α_2-chain of collagen IV and its comparison with α_1(IV) (208) 203
Schweitz, H. see R.M. Metrione (218) 59
Schwendimann, B. see B. Roth (221) 172
Scimonelli, T. and A.N. Eberle, Photoaffinity labelling of melanoma cell MSH receptors (226) 134
Scislowski, P.W.D. and E.J. Davis, Sulfur oxidation of free methionine by oxygen free radicals (224) 177
Scrutton, M.C. see D.E. Knight (223) 47
Searle, M.S. see B. Birdsall (217) 106
Sebastio, G., W. Hunziker, A. Ballabio, S. Auricchio and G. Semenza, On the primary site of control in the spontaneous development of small-intestinal sucrase-isomaltase after birth (208) 460
Sedelnikova, S.E., S.C. Agalarov, M.B. Garber and M.M. Yusupov, Proteins of the *Thermus thermophilus* ribosome: Purification of several individual proteins and crystallization of protein TL7 (220) 227
Sedgwick, E.G. and P.D. Bragg, The fluorescence intensity of the lipophilic probe *N*-phenyl-1-naphthylamine responds to the oxidation-reduction state of the respiratory chain in everted membrane vesicles of *Escherichia coli* (218) 22
Seeman, P. see H.B. Niznik (209) 71
Segade, F. see J. Gómez-Márquez (226) 217
Segawa, K. see K. Tobe (215) 345
Segond, N. see S. Minvielle (223) 63
Segovia-Quinson, B. see P. Hossenlopp (208) 439
Šegović, R. see S. Suzić (216) 287

Seidah, N.G., G.N. Hendy, J. Hamelin, J. Paquin, C. Lazure, K.M. Metters, J. Rossier and M. Chrétien, Chromogranin A can act as a reversible processing enzyme inhibitor: Evidence from the inhibition of the IRCM-serine protease 1 cleavage of pro-enkephalin and ACTH at pairs of basic amino acids (211) 144
Seidah, N.G. see S. Benjannet (224) 142
Seifert, J.-M. see E. Pöschl (226) 96
Seiki, M. see J. Inoue (209) 187
Seino, Y., K. Tanaka, J. Takeda, H. Takahashi, T. Mitani, M. Kurono, T. Kayano, G. Koh, H. Fukumoto, H. Yano, J. Fujita, N. Inagaki, Y. Yamada and H. Imura, Sequence of an intestinal cDNA encoding human motilin precursor (223) 74
Seitz, W. see J. Striessnig (212) 247
Sekiguchi, K. see M. Ido (219) 215
Sekiguchi, K. see U. Kikkawa (217) 227
Sekiguchi, K. see U. Kikkawa (223) 212
Selden, L.A., L.C. Gershman, H.J. Kinosian and J.E. Estes, Conversion of ATP-actin to ADP-actin reverses the affinity of monomeric actin for Ca^{2+} vs Mg^{2+} (217) 89
Selivanova, O.M. see L.A. Ryabova (226) 255
Selman, B.R. see M.S. Abbott (209) 157
Selman-Reimer, S. see M.S. Abbott (209) 157
Semenov, A.Yu. see L.A. Drachev (213) 128
Semenza, G. see G. Sebastio (208) 460
Semionkin, V.A. see M.I. Oshtrakh (208) 331
Semisotnov, G.V., N.A. Rodionova, V.P. Kutyshenko, B. Ebert, J. Blanck and O.B. Ptitsyn, Sequential mechanism of refolding of carbonic anhydrase B (224) 9
Senior, A.E. see T.M. Duncan (208) 1
Sensenbrenner, M. see C. Gensburger (217) 1
Senshu, M. see M. Takahashi (213) 345
Seprödi, J. see L. Buday (223) 15
Serhan, C.N., U. Hirsch, J. Palmblad and B. Samuelsson, Formation of lipoxin A by granulocytes from eosinophilic donors (217) 242
Serov, O.L. see N.M. Matveeva (217) 42
Serrano, R., C. Montesinos and A. Cid, A temperature-sensitive mutant of the yeast plasma membrane ATPase obtained by in vitro mutagenesis (208) 143
Serrano, R. see P. Eraso (224) 193
Serratosa, J. see M.J. Coll (208) 418
Seth, A. see K.P. Samuel (218) 81
Šetlík, I. see S.I. Allakhverdiev (226) 186
Šetlíková, E. see S.I. Allakhverdiev (226) 186
Seto, P. see R.P. Magnusson (208) 391
Seurin, D. see P. Hossenlopp (208) 439
Ševčík, J. see S.V. Shlyapnikov (209) 335
Seward, K. see P.W. Piper (220) 177
Seyama, Y. see S. Kitamura (213) 169
Shabarova, Z.A. see V.G. Metelev (226) 232
Shahak, Y. see A. Gal (221) 205
Sham, H.L., H. Stein, C.A. Rempel, J. Cohen and J.J. Plattner, Highly potent and specific inhibitors of human renin (220) 299
Shamim, M.T. see D. Ukena (209) 122
Sharon, N., Bacterial lectins, cell–cell recognition and infectious disease (217) 145
Sharp, G.W.G., C. Hannah-White, M. El-Sabban, M.E. Cohen and M. Donowitz, Effects of Ca^{2+}, theophylline and promethazine on protein phosphorylation in intact cells of rabbit ileum: Correlation with active Na and Cl absorption (221) 309
Shastri, K. see C.-S. Wang (222) 135
Shavit, N. see M.S. Abbott (209) 157
Shaw, A. see H.R. MacDonald (209) 295
Shaw, E.K. see J. Bennett (210) 22
Shearman, M.S. see U. Kikkawa (223) 212
Shechter, E. see A. Ghazi (209) 325
Sheikh, S.P. see T.W. Schwartz (225) 209
Sherman, L.A. see H.C. Riethman (215) 209
Sherry, A.D. see C.R. Malloy (212) 58
Shevack, A. see M. Shoham (208) 321
Shibai, H. see N. Tsuchimori (218) 205
Shibolet, S. see M.S. Meyer (212) 138
Shichida, Y. see M. Ariki (225) 255
Shikano, H. see H. Nakayama (208) 278
Shikita, M. see N. Sakai (222) 341
Shiku, H. see T. Tanimoto (226) 291
Shima, H., M. Takano, K. Shimotohno and M. Miwa, Identification of $p26^{Xb}$ and $p24^{Xb}$ of human T-cell leukemia virus type II (209) 289
Shimada, M., T. Hattori, T. Umezawa, T. Higuchi and K. Uzura, Regiospecific oxygenations during ring cleavage of a secondary metabolite, 3,4-dimethoxybenzyl alcohol catalyzed by lignin peroxidase (221) 327
Shimasaki, S. see P. Whiting (219) 459
Shimazaki, K. see K. Nitta (223) 405
Shimazu, T. see A. Takeda (210) 169
Shimizu, N. see D.S. Samuels (209) 231
Shimizu, S. see J.M. Kim (210) 77
Shimizu, T. see S. Kitamura (213) 169
Shimizu, Y. see N. Takasu (225) 43
Shimohigashi, Y., H. Kodama, S. Imazu, H. Horimoto, K. Sakaguchi, M. Waki, H. Uchida, M. Kondo, T. Kato and N. Izumiya, [4,4′-(*Z*)-Dehydrophenylalanine]gramicidin S with stabilized bioactive conformation and strong antimicrobial activity (222) 251
Shimohigashi, Y., T. Costa, A. Pfeiffer, A. Herz, H. Kimura and C.H. Stammer, V^5Phe^4-enkephalin analogs: Delta receptors in rat brain are different from those in mouse vas deferens (222) 71
Shimonishi, Y., Y. Hidaka, M. Koizumi, M. Hane, S. Aimoto, T. Takeda, T. Miwatani and Y. Takeda, Mode of disulfide bond formation of a heat-stable enterotoxin (ST_h) produced by a human strain of enterotoxigenic *Escherichia coli* (215) 165
Shimosegawa, Y. see K. Sugioka (210) 37
Shimotohno, K. see H. Shima (209) 289
Shimotohno, K. see T. Gojobori (208) 231
Shimura, K. see H. Ishihara (226) 319
Shin, M. see A. Hiwatashi (209) 311
Shinohara, T. see D. Carper (220) 209
Shinohara, T. see T. van Veen (208) 133
Shinozaki, K. see K. Yamaguchi-Shinozaki (215) 132
Shinozaki, K. see N. Zaita (210) 153
Shinozawa, T., S. Terada, H. Matsusaka and S. Yamashita, Detection of Ca^{2+}-dependent cyclic GMP binding protein in frog rod outer segments (219) 293
Shirokov, V.A. see S.D. Trakhanov (220) 319
Shivaji, S., Interaction of seminalplasmin with chlortetracycline, a fluorescent chelate probe of Ca^{2+} (218) 97
Shkuropatov, A.Y. see S.V. Chekalin (216) 245
Shlensky, A.B. see Yu.A. Ovchinnikov (226) 91
Shlyapnikov, S.V., V. Both, V.A. Kulikov, A.A. Dementiev, J. Ševčík and J. Zelinka, Amino acid sequence determination of guanyl-specific ribonuclease Sa from *Streptomyces aureofaciens* (209) 335
Shogakiuchi, Y. see H. Kai (212) 119
Shogakiuchi, Y. see H. Kanaide (214) 130
Shoham, M., J. Müssig, A. Shevack, T. Arad, H.G. Wittmann and A. Yonath, A new crystal form of large ribosomal subunits from *Halobacterium marismortui* (208) 321
Shuin, T. see Y. Kubota (212) 159
Shuvaeva, T.M. see Yu.A. Ovchinnikov (223) 169
Shuvalov, V.A. see S.V. Chekalin (216) 245
Shuvalova, L.A. see D.I. Levitsky (221) 77
Sibley, J.T. see B.L. Haug (215) 252

Siddle, K. see R.M. O'Brien (212) 281
Siddle, K. see R.M. O'Brien (217) 253
Sidler, W. see R. Rümbeli (221) 1
Siebert, F. see K. Gerwert (213) 39
Sieker, L.C., L.H. Jensen and J. LeGall, Preliminary X-ray studies of the tetra-heme cytochrome c_3 and the octa-heme cytochrome c_3 from *Desulfovibrio gigas* (209) 261
Sieker, L.C., R.E. Stenkamp, L.H. Jensen, B. Prickril and J. LeGall, Structure of rubredoxin from the bacterium *Desulfovibrio desulfuricans* (208) 73
Sies, H. see H. Wefers (211) 49
Sies, H. see J.C. Griffiths (213) 34
Siffert, G. see W. Siffert (212) 123
Siffert, W., G. Siffert, P. Scheid, T. Riemens, G. Gorter and J.W.N. Akkerman, Inhibition of Na^+/H^+ exchange reduces Ca^{2+} mobilization without affecting the initial cleavage of phosphatidylinositol 4,5-bisphosphate in thrombin-stimulated platelets (212) 123
Sigler, P.B. see T. Wagner (212) 317
Sigon, C. see W. Homan (215) 323
Silva, C.D. see F. Heymans (218) 35
Silva, J., S. Zinker and P. Gariglio, Isolation and partial characterization of 2-μm yeast plasmid as a transcriptionally active minichromosome (214) 71
Silva, R. see E. Rondinelli (208) 379
Simmons, T.W., I.S. Jamall and R.A. Lockshin, The effect of selenium deficiency on peroxidative injury in the house fly, *Musca domestica*: A role for glutathione peroxidase (218) 251
Simon, D. see R.V. Guntaka (221) 332
Simon, H.-G., U. Fruth, M.D. Kramer and M.M. Simon, A secretable serine proteinase with highly restricted specificity from cytolytic T lymphocytes inactivates retrovirus-associated reverse transcriptase (223) 352
Simon, H. see S. Nagata (210) 66
Simon, M.M. see H.-G. Simon (223) 352
Simonis, W. see M. Thaler (219) 351
Simonneau, M. see E. Leneveu (209) 165
Simpkin, D. see J. Telser (214) 117
Simpson, A.W.M. and T.J. Rink, Elevation of pH_i is not an essential step in calcium mobilisation in fura-2-loaded human platelets (222) 144
Simpson, J., I.H. Milne, J.O. Gardner, C.M. Yates, K. James and G. Fink, Antibodies to normal and Alzheimer human brain structures from non-immunised mice of various ages (217) 62
Simpson, R.J., R.L. Moritz, C.J. Lloyd, L.J. Fabri, E.C. Nice, M.R. Rubira and A.W. Burgess, Primary structure of ovine pituitary basic fibroblast growth factor (224) 128
Šimúth, J., H. Sternbach, J. Zelinka, R.M. Chomutov and A.A. Nedospasov, DNA-dependent RNA polymerase from the chlorotetracycline producing strain of *Streptomyces aureofaciens* (218) 163
Singh, B. see M.L. Langsford (225) 163
Singh, J., J.M. Thornton, M. Snarey and S.F. Campbell, The geometries of interacting arginine-carboxyls in proteins (224) 161
Singh, S. see T. Braun (215) 233
Sirois, P. see J. Gutkowska (214) 17
Sjövall, J. see H.-U. Marschall (213) 411
Sklenář, V., B.R. Brooks, G. Zon and A. Bax, Absorption mode two-dimensional NOE spectroscopy of exchangeable protons in oligonucleotides (216) 249
Sklenář, V., H. Miyashiro, G. Zon, H.T. Miles and A. Bax, Assignment of the ^{31}P and ^1H resonances in oligonucleotides by two-dimensional NMR spectroscopy (208) 94
Skripkin, E.A. see A.S. Mankin (219) 269
Skryabin, K.G. see S.Yu. Morozov (213) 438
Skulachev, V.P., Energy transduction by the photosynthetic reaction center complex from *Rhodopseudomonas viridis* (225) 1
Skulachev, V.P. see A.Yu. Andreyev (226) 265
Skulachev, V.P. see L.A. Drachev (209) 316
Skulachev, V.P. see L.A. Drachev (226) 139
Skulachev, V.P. see L.B. Margolis (220) 288
Slater, T.F. see K.H. Cheeseman (209) 191
Slebos, R.J.C. see J.W.M. Höppener (215) 122
Slepak, V.Z. see Yu.A. Ovchinnikov (226) 91
Slobin, L.I. see J.R. Greenberg (224) 54
Slominsky, P.A. see S.A. Limborska (212) 208
Smaal, E.B. see K. Nicolay (209) 33
Small, J.V. see R.A. Cross (219) 306
Smarrelli, jr, J. see D. Castignetti (209) 147
Smirnov, I. see V.L. Makarov (212) 263
Smirnov, V.N. see M.A. Glukhova (218) 292
Smirnov, V.N. see V.V. Kushnirov (215) 257
Smirnov, Yu.V. see E.D. Sverdlov (217) 275
Smirnov, Yu.V. see Yu.A. Ovchinnikov (213) 73
Smirnova, I.A., E.N. Vulfson and V.A. Kostyrko, The ATP-dependent generation of membrane potential by sub-bacterial vesicles from the marine bacterium, *Vibrio alginolyticus* (214) 343
Smith, C.E., I. Wakefield, J.A. King, Z. Naor, R.P. Millar and J.S. Davidson, The initial phase of GnRH-stimulated LH release from pituitary cells is independent of calcium entry through voltage-gated channels (225) 247
Smith, C.L. see G.L. Hammond (215) 100
Smith, J.A. see E.H.A. Wong (213) 419
Smith, J.S. see H.H. Valdivia (226) 280
Smith, jr, L.H. see F. O'Rourke (214) 176
Smith, L.D., K.J. Stevenson, D.W. Hough and M.J. Danson, Citrate synthase from the thermophilic archaebacteria *Thermoplasma acidophilum* and *Sulfolobus acidocaldarius* (225) 277
Smith, P.J., R.M. Warren and C. von Holt, The quantitation of biotinylated compounds by a solid-phase assay using a ^{125}I-labelled biotin derivative (215) 305
Smith, W.E. see J. Reglinski (214) 351
Smith, W.E. see K. Kelly (222) 120
Snarey, M. see J. Singh (224) 161
Snyderman, R. see J.R. Didsbury (211) 160
Snyderman, R. see J.R. Didsbury (219) 259
Sobieszek, A. see D. Szczęsna (210) 177
Sobue, K. see T. Okabe (213) 184
Soininen, R., T. Haka-Risku, D.J. Prockop and K. Tryggvason, Complete primary structure of the α_1-chain of human basement membrane (type IV) collagen (225) 188
Soleilhac, J.-M. see J.-M. Zajac (216) 118
Söling, H.-D. see V. Henne (218) 153
Söll, D. see G. Krupp (212) 271
Solomonson, L.P. see M.J. Barber (213) 372
Solovyov, V.V. see N.A. Kolchanov (214) 87
Solsona, C. see X. Guitart (219) 219
Solsona, C. see X. Rabasseda (213) 337
Sommarin, M. see P.-M. Melin (223) 87
Sondermeijer, P. see Y. Le Bouc (222) 181
Song, M.D., M. Wachi, M. Doi, F. Ishino and M. Matsuhashi, Evolution of an inducible penicillin-target protein in methicillin-resistant *Staphylococcus aureus* by gene fusion (221) 167
Sonobe, S. see H. Hayashi (223) 267
Sonthayanon, B. see C.-S. Wang (222) 135
Sorbi, R.T. see A. Cavaggioni (212) 225
Sørensen, A.R. see L. Thim (212) 307
Soret, J. see C. Kryceve-Martinerie (214) 81
Sorkin, A.D. see L.V. Teslenko (221) 105
Sorsa, T. see S. Lindy (208) 23

Sottrup-Jensen, L. see J. Gliemann (221) 55
Soudijn, W. see P.J.M. van Galen (223) 197
Soumpasis, D.M., M. Robert-Nicoud and T.M. Jovin, B-Z DNA conformational transition in 1:1 electrolytes: dependence upon counterion size (213) 341
Soute, B.A.M. see L.J.M. van Haarlem (222) 353
Southan, C., A. Aitken, R.A. Childs, W.M. Abbott and T. Feizi, Amino acid sequence of β-galactoside-binding bovine heart lectin: Member of a novel class of vertebrate proteins (214) 301
Southwick, W. see I. Dodd (209) 13
Souvannavong, V. see J.-P. Tenu (220) 93
Spach, G. see G. Molle (224) 208
Spandidos, D.A. and L. Holmes, Transcriptional enhancer activity in the variable tandem repeat DNA sequence downstream of the human Ha-*ras*1 gene (218) 41
Spangfort, M., U.K. Larsson, J.M. Anderson and B. Andersson, Isolation of two different subpopulations of the light-harvesting chlorophyll *a*/*b* complex of photosystem II (LHCII) (224) 343
Spassky, A. see S. Ponnambalam (219) 189
Spät, A. see L. Hunyady (225) 72
Spät, A. see L. Kiesel (217) 85
Spatz, M. see T. Yamamoto (219) 326
Spector, I. see M. Coué (213) 316
Spiegel, A. see J. Falloon (209) 352
Spiegel, A. see M.R. Brann (222) 191
Spiegel, A. see P.M. Murphy (221) 81
Spirin, A.S. see L.A. Ryabova (226) 255
Sprinzl, M. see R.L. Joshi (208) 189
Stackebrandt, E. see H.V. den Eynde (213) 301
Staehelin, L.A. see T.G. Dunahay (215) 25
Staehelin, M. see R.-J. Box (214) 323
Stahl, J. see H.-J. Kärgel (220) 126
Staib, W. see P. Schadewaldt (217) 45
Stammer, C.H. see Y. Shimohigashi (222) 71
Stammers, D.K., J.N. Champness, C.R. Beddell, J.G. Dann, E. Eliopoulos, A.J. Geddes, D. Ogg and A.C.T. North, The structure of mouse L1210 dihydrofolate reductase-drug complexes and the construction of a model of human enzyme (218) 178
Stammers, D.K. see R. Wootton (209) 129
Standaert, M.L. see D.R. Cooper (214) 122
Stapper, N.J. see P. Schadewaldt (217) 45
Stasch, J.-P. see G. Theiss (218) 159
Stecher, B., B. Höhne, U. Gras, M. Momayezi, R. Glas-Albrecht and H. Plattner, Involvement of a 65 kDa phosphoprotein in the regulation of membrane fusion during exocytosis in *Paramecium* cells (223) 25
Steenbergh, P.H., J.W.M. Höppener, J. Zandberg, A. Visser, C.J.M. Lips and H.S. Jansz, Structure and expression of the human calcitonin/CGRP genes (209) 97
Steenbergh, P.H. see J.W.M. Höppener (215) 122
Steigner, W. see M. Thaler (219) 351
Stein, H. see H.L. Sham (220) 299
Stein, P.J. see A. Caretta (219) 97
Steiner, D.F. see C.E. Sample (222) 332
Steiner, V. and J.-Y. Chang, Chemical modification of the carboxyl groups of protein substrates enhances their thrombin susceptibility (222) 6
Steinert, A. see D. Brömme (219) 441
Steinschneider, A.Y. see V.V. Kupriyanov (208) 89
Stender, H. see H. Kühn (208) 248
Stenkamp, R.E. see L.C. Sieker (208) 73
Stent, G.S., Cell lineage in development (215) 1
Steplewski, Z. see J. Thurin (208) 17
Steppuhn, J. see T. Jansen (216) 234
Sternbach, H. see J. Šimúth (218) 163
Sternberg, M.J.E., G.J. Barton, M.J. Zvelebil, J. Cookson and A.R.M. Coates, Prediction of antigenic determinants and secondary structures of the major AIDS virus proteins (218) 231
Sternlicht, H., M.B. Yaffe and G.W. Farr, A model of the nucleotide-binding site in tubulin (214) 226
Stevenson, K.J. see L.D. Smith (225) 277
Stevenson, M. see S. Dewhurst (213) 133
Stevenson, M. see S. Dewhurst (213) 138
Stewart, B.W. see C.M. Ireland (212) 173
Stewart, M.J. see J. Reglinski (214) 351
Stinson, R.A. see K. Hawrylak (212) 289
Stirm, S. see F. Altmann (221) 145
Stitt, M. see S. Krömer (226) 352
Stock, A., S. Clarke, C. Clarke and J. Stock, N-terminal methylation of proteins: structure, function and specificity (220) 8
Stock, J. see A. Stock (220) 8
Stockburger, M. see R. Diller (217) 297
Stoppani, A.O.M. see M. Dubin (220) 197
Stoppini, M. see M. Galliano (208) 364
Strålfors, P. see H. Olsson (209) 175
Stratman, F.W. see L.L. Ji (208) 297
Strid, Å., I.-M. Karlsson and M. Baltscheffsky, Demonstration of ΔpH- and Δψ-induced synthesis of inorganic pyrophosphate in chromatophores from *Rhodospirillum rubrum* (224) 348
Striessnig, J., H.-G. Knaus, M. Grabner, K. Moosburger, W. Seitz, H. Lietz and H. Glossmann, Photoaffinity labelling of the phenylalkylamine receptor of the skeletal muscle transverse-tubule calcium channel (212) 247
Strnad, C.F. and R.A. Carchman, Human T lymphocyte mitogenesis in response to the B oligomer of pertussis toxin is associated with an early elevation in cytosolic calcium concentrations (225) 16
Strömqvist, M., Brain spectrin fragments and crosslinks actin filaments (213) 102
Strong, P.N. see N.A. Castle (209) 117
Strosberg, A.D. see S. Bon (209) 206
Strosznajder, J. see H. Wikieł (216) 57
Strotmann, H., S. Kleefeld and D. Lohse, Control of ATP hydrolysis in chloroplasts (221) 265
Stuart, D.S. see M.E. Houston (219) 469
Stubbs, M. see S.J.H. Ashcroft (219) 311
Sturgess, N.C., C.N. Hales and M.L.J. Ashford, Inhibition of a calcium-activated, non-selective cation channel, in a rat insulinoma cell line, by adenine derivatives (208) 397
Styring, S. see Y. Takahashi (223) 371
Suau, P. see B. Piña (210) 161
Subbarao, K. see D.J. Cash (217) 129
Subirana, J.A. and J. Colom, Comparison of protamines from freshwater and marine bivalve molluscs: evolutionary implications (220) 193
Subirana, J.A. see M. Chiva (215) 237
Suckling, C.J. see J. Reglinski (214) 351
Suda, T. see J. Abe (226) 58
Suda, T. see T. Hayashi (218) 200
Sugai, S. see K. Kuwajima (221) 115
Sugai, S. see K. Nitta (223) 405
Sugamura, K. see M. Fujii (223) 299
Sugimoto, E. see Y. Aratani (218) 47
Sugimoto, K. see T. Kubo (209) 367
Sugioka, K. see Y. Sugioka (223) 251
Sugioka, K., Y. Shimosegawa and M. Nakano, Estrogens as natural antioxidants of membrane phospholipid peroxidation (210) 37

Sugioka, Y., M. Suzuki, K. Sugioka and M. Nakano, A ferriprotoporphyrin IX-chloroquine complex promotes membrane phospholipid peroxidation: A possible mechanism for antimalarial action (223) 251
Sugita, H. see S. Imajoh (215) 274
Sugiura, M. see K. Yamaguchi-Shinozaki (215) 132
Sugiura, M. see N. Zaita (210) 153
Sugumaran, M. see S.J. Saul (208) 113
Suki, W.N., J. Abramowitz, R. Mattera, J. Codina and L. Birnbaumer, The human genome encodes at least three non-allellic G proteins with α_i-type subunits (220) 187
Sumbilla, C. and G. Inesi, Rapid filtration measurements of Ca^{2+} release from cisternal sarcoplasmic reticulum vesicles (210) 31
Summers, L.J., T.L. Blundell, G.G. Gause and S.I. Tomarev, A computer graphics model of frog γ-crystallin based on the three-dimensional structure of calf γ-II crystallin (208) 11
Sumper, M. see R. Schlipfenbacher (209) 57
Sundblad, L.-G., K. Palmqvist and G. Samuelsson, Luminescence decay kinetics in relation to the relaxation of the transthylakoid ΔpH from high and low CO_2 adapted cells of *Scenedesmus obliquus* (209) 28
Sundby, C. see P. Mäenpää (215) 31
Suomalainen, K. see S. Lindy (208) 23
Surguchov, A.P. see V.V. Kushnirov (215) 257
Sushkov, D.G., N.P. Gritsan and L.M. Weiner, Generation of OH radical during enzymatic reduction of 9,10-anthraquinone-2-sulphonate: Can semiquinone decompose hydrogen peroxide? (225) 139
Sussenbach, J.S. see J.W.M. Höppener (215) 122
Sussenbach, J.S. see P. de Pagter-Holthuizen (214) 259
Suter, F., P. Füglistaller, D.J. Lundell, A.N. Glazer and H. Zuber, Amino acid sequences of α-allophycocyanin B from *Synechococcus* 6301 and *Mastigocladus laminosus* (217) 279
Suter, F. see R. Rümbeli (221) 1
Suter, F. see T.D. Wechsler (210) 189
Sutton, R. see F.M. Ashcroft (215) 9
Sutton, R. see P. Hammonds (223) 131
Suyama, A. see R. Hanai (226) 247
Suzić, S., L. Radunović, V. Janković and R. Šegović, Effects of protein-free diet in amino acid homeostatis of rat blood plasma and gut contents (216) 287
Suzuki, A. see F. Inagaki (212) 91
Suzuki, A. see F. Inagaki (219) 45
Suzuki, A. see H. Jhoti (219) 419
Suzuki, H. see N. Sakai (222) 341
Suzuki, K., S. Imajoh, Y. Emori, H. Kawasaki, Y. Minami and S. Ohno, Calcium-activated neutral protease and its endogenous inhibitor: Activation at the cell membrane and biological function (220) 271
Suzuki, K. see A.M. Vaccaro (216) 190
Suzuki, K. see K. Ishidoh (223) 69
Suzuki, K. see K. Ishidoh (226) 33
Suzuki, K. see K. Kubo (223) 138
Suzuki, K. see S. Imajoh (215) 274
Suzuki, K. see S. Ohno (222) 279
Suzuki, M. see F. Inagaki (219) 45
Suzuki, M. see Y. Sugioka (223) 251
Sverdlov, E.D., G.S. Monastyrskaya, N.E. Broude, Yu.A. Ushkaryov, R.L. Allikmets, A.M. Melkov, Yu.V. Smirnov, I.V. Malyshev, I.E. Dulobova, K.E. Petrukhin, A.V. Grishin, N.I. Kijatkin, M.B. Kostina, V.E. Sverdlov, N.N. Modyanov and Yu.A. Ovchinnikov, The family of human Na^+,K^+-ATPase genes: No less than five genes and/or pseudogenes related to the α-subunit (217) 275
Sverdlov, E.D., N.E. Broude, V.E. Sverdlov, G.S. Monastyrskaya, A.V. Grishin, K.E. Petrukhin, N.S. Akopyanz, N.N. Modyanov and Y.A. Ovchinnikov, Family of Na^+, K^+-ATPase genes: Intra-individual tissue-specific restriction fragment length polymorphism (221) 129
Sverdlov, E.D., S.A. Tsarev, R.A. Krykbaev, I.P. Chernov and V.M. Rostapshov, trp operon induction during the expression in *E. coli* of two IFN-γ sequences (212) 233
Sverdlov, E.D. see N.M. Matveeva (217) 42
Sverdlov, E.D. see Yu.A. Ovchinnikov (213) 73
Sverdlov, V.E. see E.D. Sverdlov (217) 275
Sverdlov, V.E. see E.D. Sverdlov (221) 129
Sverdlov, V.E. see Yu.A. Ovchinnikov (213) 73
Svinukhova, I.A. and A.A. Boldyrev, Na^+ transport by reconstituted Na^+,K^+-ATPase in the presence of various nucleotides (214) 335
Swanljung-Collins, H. see J.P. Rieker (212) 154
Swanson, K.L. see D.R.E. Macallan (226) 357
Swanson, K.L. see Y. Aracava (222) 63
Swiston, L. see M. Oetting (208) 99
Syoto, B. see I. Matsuoka (216) 295
Szczepan, E.W., D. Kaller, J.F. Honek and T. Viswanatha, Direct evidence for the participation of pyruvate in *N*-hydroxylation of lysine (211) 239
Szczęsna, D., A. Sobieszek and I. Kąkol, Binding of phosphorylated and dephosphorylated heavy meromyosin to F-actin (210) 177
Szekeres, M. and A.V. Matveyev, Cleavage and sequence recognition of 2,6-diaminopurine-containing DNA by site-specific endonucleases (222) 89
Szyszka, R. see W. Kudlicki (215) 16

T

Tabaqchali, S. see B.W. Wren (225) 82
Tabata, T. see K. Mikami (223) 273
Tado, T. see K. Satoh (216) 7
Tagliaferri, P. see D. Katsaros (223) 97
Tagliaferri, P. see T. Clair (224) 377
Taguchi, R., M. Kawase and H. Ikezawa, Determination of covalently bound *myo*-inositol in bovine erythrocyte acetylcholinesterase and porcine kidney alkaline phosphatase (225) 273
Takagi, T. see C.P. Schelling (214) 21
Takagi, Y. see Y. Hirata (219) 369
Takahashi, H. see T. Kubo (209) 367
Takahashi, H. see Y. Seino (223) 74
Takahashi, M. and M. Senshu, Two distinct DNA ligases from *Drosophila melanogaster* embryos (213) 345
Takahashi, M. see Y. Takahashi (208) 347
Takahashi, Y. and S. Styring, A comparative study of the reduction of EPR signal II_{slow} by iodide and the iodo-labeling of the D2-protein in photosystem II (223) 371
Takahashi, Y., M. Takahashi and K. Satoh, Identification of the site of iodide photooxidation in the photosystem II reaction center complex (208) 347
Takahashi, Y. see H. Oh-oka (218) 52
Takahashi, Y. see T. Yamakuni (223) 117
Takai, A., C. Bialojan, M. Troschka and J.C. Rüegg, Smooth muscle myosin phosphatase inhibition and force enhancement by black sponge toxin (217) 81

Takai, T., K. Wada and T. Tanabe, Primary structure of the biotin-binding site of chicken liver acetyl-CoA carboxylase (212) 98
Takai, Y. see K. Kariya (217) 69
Takai, Y. see K. Kariya (219) 119
Takai, Y. see T. Tanimoto (226) 291
Takai, Y. see T. Tsuda (208) 39
Takaiwa, F., H. Ebinuma, S. Kikuchi and K. Oono, Nucleotide sequence of a rice glutelin gene (221) 43
Takaku, F. see K. Tobe (215) 345
Takami, N. see K. Oda (214) 135
Takamiya, S., M.A. Lindorfer and R.A. Capaldi, Purification of all thirteen polypeptides of bovine heart cytochrome *c* oxidase from one aliquot of enzyme: Characterization of bovine fetal heart cytochrome *c* oxidase (218) 277
Takano, E., M. Maki, M. Hatanaka, H. Mori, K. Zenita, T. Sakihama, R. Kannagi, T. Marti, K. Titani and T. Murachi, Evidence for the repetitive domain structure of pig calpastatin as demonstrated by cloning of complementary DNA (208) 199
Takano, E. see M. Maki (223) 174
Takano, M. see H. Shima (209) 289
Takano, T. see K. Tobe (215) 345
Takasu, N., T. Yamada and Y. Shimizu, Epidermal growth factor (EGF), tumor promoter 12-*O*-tetradecanoylphorbol 13-acetate (TPA) and calcium ionophore A23187 increase cytoplasmic free calcium and stimulate arachidonic acid release and PGE_2/6-keto $PGF_{1\alpha}$ production in cultured porcine thyroid cells (225) 43
Takata, S. see Y. Hirata (219) 369
Takatsuki, A. see K. Oda (214) 135
Takayama, H. see T. Hayashi (218) 200
Takayama, K. see C.A. Ellis (218) 238
Takeda, A., E. Hashimoto, H. Yamamura and T. Shimazu, Phosphorylation of liver gap junction protein by protein kinase C (210) 169
Takeda, J. see Y. Seino (223) 74
Takeda, T. see Y. Shimonishi (215) 165
Takeda, Y. see Y. Shimonishi (215) 165
Takemoto, D.J. see D.F. Morrison (222) 266
Takeuchi, A. see M. Maeshima (220) 23
Takeyama, M. see S.-Y. Hsu (218) 222
Takio, K. see T. Marti (219) 415
Talwar, H. see I. Magnaldo (210) 6
Tamai, N. see M. Mimuro (213) 119
Tanabe, T. see K. Wada (209) 330
Tanabe, T. see T. Takai (212) 98
Tanaka, A. see M. Ueda (220) 31
Tanaka, H. see T. Hayashi (218) 200
Tanaka, K.I. see T. Kurosaki (214) 253
Tanaka, K. see T. Tobimatsu (222) 56
Tanaka, K. see Y. Seino (223) 74
Tanaka, M. see M. Doi (213) 265
Tanaka, T. see H. Nakajima (223) 113
Tanaka, Y. see M. Ohshima (225) 243
Tancredi, T. see A. Motta (215) 215
Tanimoto, T., M. Hoshijima, M. Kawata, K. Yamamoto, T. Ohmori, H. Shiku, H. Nakano and Y. Takai, Binding of *ras* p21 to bands 4.2 and 6 of human erythrocyte membranes (226) 291
Tanokura, M., M. Imaizumi and K. Yamada, A calorimetric study of Ca^{2+} binding by the parvalbumin of the toad (*Bufo*): distinguishable binding sites in the molecule (209) 77
Tanoue, K. see A. Yamaguchi (225) 228
Tarasevich, M.R. see A.M. Kuznetsov (215) 219
Tarui, S. see H. Nakajima (223) 113
Tas, P.W.L. see H.G. Kress (221) 28
Tastenoy, M. see M. Waelbroeck (226) 287
Tate, S.S. and B. Nash, Membrane translocation and insertion of NH_2-terminally anchored γ-glutamyl transpeptidase require a signal recognition particle (211) 133
Tateson, J.E. see R.W. Randall (214) 167
Tatischeff, I. see R. Thiery (223) 381
Tatsuno, Y. see S. Hashimoto (208) 305
Tatti, M. see A.M. Vaccaro (216) 190
Taurog, A. see R.P. Magnusson (208) 391
Taylor, A., C.C. Allende, R. Weinmann and J.E. Allende, The phosphorylation of nucleoplasmin by casein kinase-2 is resistant to heparin inhibition (226) 109
Taylor, D.M. see B.N. Zaba (213) 49
Taylor, P.M. and M.J. Rennie, Perivenous localisation of Na-dependent glutamate transport in perfused rat liver (221) 370
Taylor, P. see D. Aslanian (219) 202
Taylor, S.I. see S.A. Phillips (212) 141
Tedeschi, H. see K.W. Kinnally (226) 83
Teelken, A.W. see G.T. Robillard (224) 391
Telfer, A. see J. Barber (220) 67
Telser, J., B.M. Hoffman, R. LoBrutto, T. Ohnishi, A.-L. Tsai, D. Simpkin and G. Palmer, Evidence for N coordination to Fe in the [2Fe-2S] center in yeast mitochondrial complex III: Comparison with similar findings for analogous bacterial [2Fe-2S] proteins (214) 117
Tempest, P.R., C.S. Cooper and G.N. Major, The activated human *met* gene encodes a protein tyrosine kinase (209) 357
Temussi, P.A. see A. Motta (215) 215
Tennant, L. see C. Dalvit (213) 289
Tenu, J.-P., A. Adam, V. Souvannavong, G. Barratt, A. Yapo, J.-F. Petit, M. Level, M. Clemance and K. Douglas, A novel muramyl peptide derivative stimulates tumoricidal activity of macrophages and antibody production by B cells (220) 93
Ter-Avanesyan, M.D. see V.V. Kushnirov (215) 257
Terada, S. see T. Shinozawa (219) 293
Teranishi, Y. see M. Ueda (220) 31
Teraoka, H. see O. Ohara (211) 78
Teraoka, H. see Y. Ohmura (208) 451
Tertoolen, L.G.J., B.C. Tilly, R.F. Irvine and W.H. Moolenaar, Electrophysiological responses to bradykinin and microinjected inositol polyphosphates in neuroblastoma cells: Possible role of inositol 1,3,4-trisphosphate in altering membrane potential (214) 365
Terzi, G. see T. Marti (219) 415
Teslenko, L.V., E.S. Kornilova, A.D. Sorkin and N.N. Nikolsky, Recycling of epidermal growth factor in A431 cells (221) 105
Tessier, L.-H., S. Jallat, M. Sauvageot, R.G. Crystal and M. Courtney, RNA structural elements for expression in *Escherichia coli:* α_1-Antitrypsin synthesis using translation control elements based on the cII ribosome-binding site of phage λ (208) 183
Testa, I. see M. Clementi (221) 11
Tew, D.G. see M.J. Green (216) 31
Thaler, M., W. Steigner, K. Köhler, W. Simonis and W. Urbach, Release of repetitive transient potentials and opening of potassium channels by barium in *Eremosphaera viridis* (219) 351
Thauer, R.K. see A. Kobelt (214) 265
Thauer, R.K. see D. Ankel-Fuchs (213) 123
Thauer, R.K. see H.P.C. Hogenkamp (219) 197
Thauer, R.K. see J. Ellermann (220) 358
Theiss, G., A. John, F. Morich, D. Neuser, W. Schröder, J.-P. Stasch and S. Wohlfeil, α-h-ANP is the only form of circulating ANP in humans (218) 159
Thelestam, M. see L. Blomqvist (211) 127
Thiery, J.P. see M. Obara (213) 261

Thiery, R., R. Klein and I. Tatischeff, Increase of DPH fluorescence polarization during development of *Dictyostelium discoideum* cells (223) 381
Thim, L., M.T. Hansen and A.R. Sørensen, Secretion of human insulin by a transformed yeast cell (212) 307
Thim, L. see A.J. Moody (212) 302
Thim, L. see J.M. Conlon (208) 445
Thim, L. see J.M. Conlon (214) 50
Thom, R.E. and J.E. Casnellie, Demonstration that LSTRA cells have an elevated level of proteins phosphorylated on tyrosine residues (222) 104
Thomas, H. see P. Hilditch (208) 313
Thomas, V. see J.G. Delcros (220) 236
Thomas, W.E. and J. Mowbray, Evidence for ADP-ribosylation in the mechanism of rapid thyroid hormone control of mitochondria (223) 279
Thomazy, V. see L. Fesus (224) 104
Thompson, J., S.E. Moore and F.S. Walsh, Thyroid hormones regulate expression of the neural cell adhesion molecule in adult skeletal muscle (219) 135
Thompson, N.T. see J. Dawson (214) 171
Thompson, N.T. see R.W. Bonser (209) 134
Thompson, R.J. see I.N.M. Day (210) 157
Thompson, R.J. see I.N.M. Day (222) 139
Thompson, R.J. see U.S. Vogel (218) 261
Thomson, A.J. see N. Foote (214) 347
Thomson, A.J. see R.S. Blackmore (219) 244
Thörnström, P.-E. see M. Fabian (213) 396
Thornton, J.M. see J. Singh (224) 161
Thorpe, P.E. see E.J. Wawrzynczak (219) 51
Thorpe, P.E. see M. O'Hare (216) 73
Thurin, J., M. Thurin, M. Herlyn, D.E. Elder, Z. Steplewski, W.H. Clark, jr and H. Koprowski, GD2 ganglioside biosynthesis is a distinct biochemical event in human melanoma tumor progression (208) 17
Thurin, M. see J. Thurin (208) 17
Tiemann, K., W. Hinderer and W. Barz, Isolation of NADPH:isoflavone oxidoreductase, a new enzyme of Pterocarpan phytoalexin biosynthesis in cell suspension cultures of *Cicer arietinum* (213) 324
Tilly, B.C. see L.G.J. Tertoolen (214) 365
Timmerman, M.P. and C.C. Ashley, Fura-2 diffusion and its use as an indicator of transient free calcium changes in single striated muscle cells (209) 1
Timmerman, M.P. see A.P. Jackson (216) 35
Timmis, K.N. see J.L. Ramos (226) 241
Timpl, R. see K. Mann (218) 167
Timpmann, K.E. see Z.G. Fetisova (223) 161
Tischenko, S.V. see S.D. Trakhanov (220) 319
Titani, K. see E. Takano (208) 199
Tittor, J. see R. Diller (217) 297
Tjoeng, F.S. see G.M. Olins (224) 325
Toazara, J. see J. Borg (213) 406
Tobe, K., M. Kasuga, H. Kitasato, F. Takaku, T. Takano and K. Segawa, Differential effects of DNA tumor virus nuclear oncogene products on adipocyte differentiation (215) 345
Tobimatsu, T., Y. Fujita, K. Fukuda, K. Tanaka, Y. Mori, T. Konno, M. Mishina and S. Numa, Effects of substitution of putative transmembrane segments on nicotinic acetylcholine receptor function (222) 56
Tocqué, B., A. Pfeiffer and A. Klee, Transfer of functional opiate receptors from membranes to recipient cells by polyethylene glycol-induced fusion (222) 327
Toh, H., A. Imamura and K. Kanda, Role of Mg^{2+} in the ribozyme system (219) 279
Tohda, M. see Y. Nomura (216) 40
Tojo, T. see D.S. Samuels (209) 231
Tokuda, H., T. Udagawa, M. Asano, T. Yamamoto and T. Unemoto, Conjugation-dependent recovery of the Na^+ pump in a mutant of *Vibrio alginolyticus* lacking three subunits of the Na^+ pump (215) 335
Tokunaga, F. see H. Higashiyama (218) 287
Toleikis, A. see B. Kholodenko (223) 247
Tomarev, S.I. see L.J. Summers (208) 11
Tomasi, M. see E. Papini (215) 73
Tominaga, S., A 63 kDa protein is secreted from BALB/c-3T3 cells entering the G_1 phase from the G_0 state (226) 53
Tomita, N., A. Horii, T. Yamamoto, M. Ogawa, T. Mori and K. Matsubara, Expression of pancreatic secretory trypsin inhibitor gene in neoplastic tissues (225) 113
Tomiyama, N. see A. Yamaguchi (208) 43
Tommassen, J. and T. de Kroon, Subcellular localization of a PhoE-LacZ fusion protein in *E. coli* by protease accessibility experiments reveals an inner-membrane-spanning form of the protein (221) 226
Tomoda, A., Y. Yoneyama, T. Yamaguchi, K. Kakinuma, K. Kawasaki and D. Yonemura, Spectroscopic studies of brunescent cataractous lenses (219) 472
Tompa, P., J. Batke and J. Ovádi, How to determine the efficiency of intermediate transfer in an interacting enzyme system? (214) 244
Tonevitsky, A.G. see T.L. Bushueva (215) 155
Tongio, M.-M. see B. Uring-Lambert (217) 65
Tonk, W.J.M. see J.J.S. van Rensen (226) 347
Töpfer-Petersen, E. and A. Henschen, Acrosin shows zona and fucose binding, novel properties for a serine proteinase (226) 38
Topping, D.L. see P.D. Roach (222) 159
Torazawa, K. see N. Zaita (210) 153
Torchilin, V.P. see J. Lasch (214) 13
Torrence, P.F., D. Alster, S. Huss, G. Gosselin and J.-L. Imbach, Degradation by the 2′,5′-phosphodiesterase activity of mouse cells requires the presence of a *ribo* hydroxyl group in the penultimate position of the oligonucleotide substrate (212) 267
Torres de Castro, F. see E. Rondinelli (208) 379
Tortora, G. see D. Katsaros (223) 97
Tortora, P. see S. Lamponi (216) 265
Totsuka, A. see C. Fukazawa (224) 125
Tóugu, V., A. Pedak, T. Kesvatera and A. Aaviksaar, Acetylcholinesterase as polyelectrolyte in reaction with cationic substrates (225) 77
Tourbez, M. see F. Pochon (217) 101
Toutant, M., D. Aunis, J. Bockaert, V. Homburger and B. Rouot, Presence of three pertussis toxin substrates and $G_o\alpha$ immunoreactivity in both plasma and granule membranes of chromaffin cells (215) 339
Toutant, M., J. Bockaert, V. Homburger and B. Rouot, G-proteins in *Torpedo marmorata* electric organ: Differential distribution in pre- and post-synaptic membranes and synaptic vesicles (222) 51
Towatari, T. see K. Ishidoh (223) 69
Trakhanov, S.D., M.M. Yusupov, S.C. Agalarov, M.B. Garber, S.N. Ryazantsev, S.V. Tischenko and V.A. Shirokov, Crystallization of 70 S ribosomes and 30 S ribosomal subunits from *Thermus thermophilus* (220) 319
Trakht, I.N. see A.A. Preobrazhensky (224) 23
Travers, M.T. and J.T. Knowler, Oestrogen-induced expression of oncogenes in the immature rat uterus (211) 27
Trayer, I.P. see J.S. Evans (208) 211
Trémeau, O., J.-C. Boulain, J. Couderc, P. Fromageot and A. Ménez, A monoclonal antibody which recognized the functional site of snake neurotoxins and which neutralizes all short-chain variants (208) 236
Trémolières, A. see A. Oursel (219) 393
Trewavas, A.J. see S. Gilroy (212) 133

Trimble, R.P. see P.D. Roach (222) 159
Trimble, W.S. and N. Hozumi, Deletion analysis of the c-Ha-*ras* oncogene promoter (219) 70
Trommer, W.E. see A. Wolf (212) 203
Troschka, M. see A. Takai (217) 81
Trudelle, Y., P. Daumas, F. Heitz, C. Etchebest and A. Pullman, Experimental and theoretical study of gramicidin P, an analog of gramicidin A with a methylamine C-terminal (216) 11
Trüeb, B. see T. Schreier (213) 319
Trugnan, G. see M. Rousset (208) 34
Truman, J.W. see T. Marti (219) 415
Trumpower, B.L. see W.E. Payne (213) 107
Tryggvason, K. see R. Soininen (225) 188
Tryggvason, K. see S.L. Hostikka (216) 281
Tryggvason, K. see S.L. Hostikka (224) 297
Tsai, A.-L. see J. Telser (214) 117
Tsarev, S.A. see E.D. Sverdlov (212) 233
Tschank, G. see H.M. Hanauske-Abel (214) 236
Tschopp, J. see D. Masson (208) 84
Tselepis, A.D. see M. Lekka (208) 52
Tsernoglou, D. see K. Petratos (218) 209
Tsetlin, V.I. see A.S. Arseniev (213) 283
Tsou, K. see Y.X. Zhu (208) 253
Tsoukatos, D. see M. Lekka (208) 52
Tsuchimori, N., S. Miyashiro, H. Shibai and S. Ikegami, Isolation and identification of a specific and reversible inhibitor of starfish development (218) 205
Tsuda, T. see K. Kariya (217) 69
Tsuda, T., Y. Hamamori, T. Yamashita, Y. Fukumoto and Y. Takai, Involvement of three intracellular messenger systems, protein kinase C, calcium ion and cyclic AMP, in the regulation of c-*fos* gene expression in Swiss 3T3 cells (208) 39
Tsuge, H. see K. Nitta (223) 405
Tsukada, K. see Y. Ohmura (208) 451
Tsunoda, Y. and H. Matsumiya, Calcium-activated membrane depolarization via modulation of chloride efflux from parietal cells during gastrin stimulation (222) 149
Tsvetkov, Yu.D. see S.A. Dikanov (224) 75
Tu, A.T. see A. Maurer (224) 89
Tucker, M.M. see M.L. Ernst-Fonberg (215) 261
Tuite, M.F., B.S. Cox and C.S. McLaughlin, A ribosome-associated inhibitor of in vitro nonsense suppression in [*psi*$^-$] strains of yeast (225) 205
Turner, A.J. see K.B. Widdows (222) 125
Turner, D.L. see H. Santos (226) 179
Turner, P.R. see J.D. Bell (219) 239
Turto, H. see S. Lindy (208) 23
Tushima, S. see Y. Kuroda (224) 137
Tyihák, E. see Z. Huszti (209) 362
Tysnes, O.-B., A.J.M. Verhoeven, G.M. Aarbakke and H. Holmsen, Phosphoinositide metabolism in resting and thrombin-stimulated human platelets: Evidence against metabolic heterogeneity (218) 68
Tzartos, S.J. see B. Roth (221) 172

U

Uchida, H. see Y. Shimohigashi (222) 251
Udagawa, T. see H. Tokuda (215) 335
Udaka, K. see C. Fukazawa (224) 125
Ueda, M., H. Okada, T. Hishida, Y. Teranishi and A. Tanaka, Isolation of several cDNAs encoding yeast peroxisomal enzymes (220) 31
Uehara, S. see W. Sakamoto (219) 437
Uehara, Y., S. Gasa, A. Makita, K. Sakurada and T. Miyazaki, Protein phosphorylation of lysosomal arylsulfatase B in normal and leukemic leukocytes (208) 352
Uejima, Y., T. Koga and T. Kamihara, Enhanced metabolism of phosphatidylinositol in *Candida tropicalis* in association with filamentous growth caused by ethanol (214) 127
Ueno, K. see K. Itoh (213) 85
Uhlin, B.E. see J. Sanchez (208) 194
Uhl, R. see R. Wagner (221) 253
Ui, M. see H. Kurose (219) 375
Ui, M. see T. Katada (213) 353
Ukena, D., K.A. Jacobson, W.L. Padgett, C. Ayala, M.T. Shamim, K.L. Kirk, R.A. Olsson and J.W. Daly, Species differences in structure-activity relationships of adenosine agonists and xanthine antagonists at brain A1 adenosine receptors (209) 122
Ukena, D., W.L. Padgett, O. Hong, J.W. Daly, D.T. Daly and R.A. Olsson, N^6-substituted 9-methyladenines: a new class of adenosine receptor antagonists (215) 203
Ulbrich, N. see R.K. Hartmann (218) 215
Umeda, T. see N. Sakai (222) 341
Umezawa, T. and T. Higuchi, Mechanism of aromatic ring cleavage of *β-O*-4 lignin substructure models by lignin peroxidase (218) 255
Umezawa, T. see M. Shimada (221) 327
Umezawa, T. see S. Kawai (210) 61
Underhill, D.A. see G.L. Hammond (215) 100
Unemoto, T. see H. Tokuda (215) 335
Unger, F.M. see F. Altmann (221) 145
Unson, C. see J. Falloon (209) 352
Urbach, W. see M. Thaler (219) 351
Uring-Lambert, B., N. Vegnaduzzi, M.C. Carroll, M.-M. Tongio, J. Goetz and G. Hauptmann, Heterogeneity in the structural basis of the human complement C4A null allele (C4A*Q0) as revealed by *Hin*dIII restriction fragment length polymorphism analysis (217) 65
Usanga, E.A., E. O'Brien and L. Luzzato, Mitotic inhibitors arrest the growth of *Plasmodium falciparum* (209) 23
Ushkaryov, Yu.A. see E.D. Sverdlov (217) 275
Ushkaryov, Yu.A. see Yu.A. Ovchinnikov (213) 73
Utsumi, K. see E.F. Sato (214) 181
Utsumi, T., Y. Aizono and G. Funatsu, Receptor-mediated interaction of ricin with the lipid bilayer of ganglioside G_{M1}-liposomes (216) 99
Uzura, K. see M. Shimada (221) 327

V

Vaccaro, A.M., M. Muscillo, R. Salvioli, M. Tatti, E. Gallozzi and K. Suzuki, The binding of glucosylceramidase to glucosylceramide is promoted by its activator protein (216) 190
Vaeck, M. see H. Höfte (226) 364
Vaheri, A. see E.-M. Salonen (221) 381
Valdivia, H.H., J.S. Smith, B.M. Martin, R. Coronado and L.D. Possani, Charybdotoxin and noxiustoxin, two homologous peptide inhibitors of the $K^+(Ca^{2+})$ channel (226) 280

Valeins, H., T. Volker, O. Viratelle and J. Labouesse, A quenched-flow study of a receptor-triggered second messenger response: cyclic AMP burst elicited by isoproterenol in C6 glioma cell membranes (226) 331
Valente, L. see J.K. French (212) 242
Vallee, B.L. see T. Fairwell (222) 99
Vamecq, J. see C. Van den Branden (222) 21
Van Batenburg, M.J. see G.R. Elliott (217) 6
Van Beeumen, J.J. see J.R. Vanfleteren (211) 59
Van Belle, H. see D. de Chaffoy de Courcelles (219) 283
Van Boom, J.H. see P.D. Baas (218) 119
Van Bun, S.M. see J.R. Vanfleteren (211) 59
Van Damme, E.J.M., A.K. Allen and W.J. Peumans, Isolation and characterization of a lectin with exclusive specificity towards mannose from snowdrop (*Galanthus nivalis*) bulbs (215) 140
Van Damme, E.J.M. see A.M. Vranken (216) 67
Van Damme, J. see M.I. Darville (224) 317
Van den Bosch, H. see A.J. Aarsman (219) 176
Van den Brande, J.L. see J.W.M. Höppener (215) 122
Van den Brande, J.L. see P. de Pagter-Holthuizen (214) 259
Van den Branden, C., J. Vamecq, G. Dacremont, N. Premereur and F. Roels, Short and long term influence of phenothiazines on liver peroxisomal fatty acid oxidation in rodents (222) 21
Van den Briel, W. see W. Homan (215) 323
Van den Ende, H. see W. Homan (215) 323
Van den Eijnden, D.H. see A.H.L. Koenderman (222) 42
Van den Eijnden, D.H. see D.H. Joziasse (221) 139
Van den Eynde, H. and R. De Wachter, Variable base pairing in a helix of eubacterial 5 S ribosomal RNA points to the existence of a conformational switch (217) 191
Van den Eynde, H., E. Stackebrandt and R. De Wachter, The structure of the 5 S ribosomal RNA of a member of the phylum of green non-sulfur bacteria and relatives (213) 301
Van der Kammen, R. see P. de Pagter-Holthuizen (214) 259
Van der Meer, R.A., J.A. Jongejan and J.A. Duine, Phenylhydrazine as probe for cofactor identification in amine oxidoreductases: Evidence for PQQ as the cofactor in methylamine dehydrogenase (221) 299
Van der Zanden, A.L. see H.K. Ploos van Amstel (222) 186
Van Dorsselaer, A. see G.J. Arlaud (222) 129
Van Dun, J. see W. Wouters (213) 359
Van Esch, J.H. see A.M. Batenburg (223) 148
Van Galen, P.J.M., A.P. IJzerman and W. Soudijn, Adenosine derivatives with N^6-alkyl, -alkylamine or -alkyladenosine substituents as probes for the A_1-receptor (223) 197
Van Gorkom, H.J. see R.V. Danielius (213) 241
Van Haarlem, L.J.M., B.A.M. Soute and C. Vermeer, Vitamin K-dependent carboxylase: Possible role for thioredoxin in the reduction of vitamin K metabolites in liver (222) 353
Van Heel, M.G. see E.J. Boekema (217) 283
Van Hoek, A. see A.J.W.G. Visser (224) 406
Van Kan, P.J.M. see R.V. Danielius (213) 241
Van Kuik, J.A., J. Breg, C.E.M. Kolsteeg, J.P. Kamerling and J.F.G. Vliegenthart, Primary structure of the acidic carbohydrate chain of hemocyanin from *Panulirus interruptus* (221) 150
Van Lint, J. see J.R. Vandenheede (211) 190
Van Lookeren-Campagne, M.M. see J.R. Vanfleteren (211) 190
Van Lookeren Campagne, M.M. see R.J. Aerts (220) 366
Van Mansfeld, A.D.M. see P.D. Baas (218) 119
Van Pee, K.-H. see W. Wiesner (209) 321
Van Rensen, J.J.S., W.J.M. Tonk and S.M. de Bruijn, Involvement of bicarbonate in the protonation of the secondary quinone electron acceptor of photosystem II via the non-haem iron of the quinone-iron acceptor complex (226) 347
Van Schaftingen, E. see E. Mertens (221) 124
Van Schaik, F.M.A. see P. de Pagter-Holthuizen (214) 259
Van Teeffelen, H.A.A.M. see P.D. Baas (218) 119
Van Veen, T., A. Katial, T. Shinohara, D.J. Barrett, B. Wiggert, G.J. Chader and J.M. Nickerson, Retinal photoreceptor neurons and pinealocytes accumulate mRNA for interphotoreceptor retinoid-binding protein (IRBP) (208) 133
Van Walraven, H.S., N.P. Haak, K. Krab and R. Kraayenhof, Evidence for a high proton translocation stoichiometry of the H^+-ATPase complex in well coupled proteoliposomes reconstituted from a thermophilic cyanobacterium (208) 138
Vandekerckhove, J., B. Schering, M. Bärmann and K. Aktories, *Clostridium perfringens* iota toxin ADP-ribosylates skeletal muscle actin in Arg-177 (225) 48
Vandekerckhove, J. see M.I. Darville (224) 317
Vandekerckhove, J. see N. Geisler (221) 403
Vanden Abeele, C. see J.R. Vandenheede (211) 190
Vanden Abeele, C. see J.R. Vandenheede (216) 291
Vandenheede, J.R., C. Vanden Abeele and W. Merlevede, On the dephosphorylation of the ATP,Mg-dependent protein phosphatase modulator (216) 291
Vandenheede, J.R., J.Van Lint, C.Vanden Abeele and W. Merlevede, Interaction of myelin basic protein with the different components of the ATP,Mg-dependent protein phosphatase system (211) 190
Vandenheede, J.R. see P. Agostinis (224) 385
Vandewalle, P.L. and N.O. Petersen, Oxidation of reduced cytochrome *c* by hydrogen peroxide: Implications for superoxide assays (210) 195
Vanfleteren, J.R., S.M.V. Bun and J.J.V. Beeumen, The primary structure of histone H3 from the nematode *Caenorhabditis elegans* (211) 59
Vänngård, T. see R. Aasa (221) 245
Varečka, Ľ., E. Peterajová and J. Pogády, Polymyxin B, a novel inhibitor of red cell Ca^{2+}-activated K^+ channel (225) 173
Varela, I., M.M.V.L. Campagne, J.F. Alvarez and J.M. Mato, The developmental regulation of phosphatidylinositol kinase in *Dictyostelium discoideum* (211) 64
Varetto, L., J.-M. Frère and J.-M. Ghuysen, The importance of the negative charge of β-lactam compounds for the inactivation of the active-site serine DD-peptidase of *Streptomyces* R61 (225) 218
Vargas, F. and J.-C. Schwartz, Apparent identity of cerebral tyrosylsulfotransferase activities using either a cholecystokinin derivative or an acidic amino acid polymer as substrate (211) 234
Vargas, J.L., E. Roche, E. Knecht and S. Grisolía, Differences in the half-lives of some mitochondrial rat liver enzymes may derive partially from hepatocyte heterogeneity (224) 182
Varro, A. see H. Desmond (210) 185
Vashakidze, R.P. and D.A. Prangishvili, Simple repetitive sequences in the genomes of archaebacteria (216) 217
Vasilevskaya, T.D. see M.A. Glukhova (218) 292
Vasiliev, V.D. see L.A. Ryabova (226) 255
Vass, I., T. Ono and Y. Inoue, Removal of 33 kDa extrinsic protein specifically stabilizes the $S_2Q_A^-$ charge pair in photosystem II (211) 215
Vassalli, J.-D. and D. Belin, Amiloride selectively inhibits the urokinase-type plasminogen activator (214) 187
Vasta, V. see P. Bruni (222) 27
Vazquez, C.R. see A.R. Nebreda (220) 27
Vedeler, A. see G. Burns (208) 217
Vegnaduzzi, N. see B. Uring-Lambert (217) 65
Venter, J.C. see F.-Z. Chung (211) 200
Venton, D.L. see E.J. Kattelman (213) 179
Venturoli, G., M. Virgili, B.A. Melandri and A.R. Crofts, Kinetic measurements of electron transfer in coupled chromatophores from photosynthetic bacteria: A method of correction for the electrochromic effects (219) 477
Verbert, A. see R. Cecchelli (208) 407

Verhoeven, A.J.M. see O.-B. Tysnes (218) 68
Verkleij, A.J. see A.M. Batenburg (223) 148
Vermeer, C. see L.J.M. van Haarlem (222) 353
Vermeulen, J.A.W.H., R.M.J.N. Lamerichs, L.J. Berliner, A.De Marco, M. Llinás, R. Boelens, J. Alleman and R. Kaptein, ^{1}H NMR characterization of two crambin species (219) 426
Vernotte, C. see J. Olive (208) 308
Vianello, A. see F. Macrì (215) 47
Vicendo, P. see J. Fauvel (221) 397
Vicentini, L.M. see A. Pandiella-Alonso (208) 48
Vidal, M., J. Sainte-Marie, J.R. Philippot and A. Bienvenue, The influence of coupling transferrin to liposomes or minibeads on its uptake and fate in leukemic L_2C cells (216) 159
Vidali, G. see A. De Ambrosis (225) 120
Viel, A., M.K. Djé, A. Mazabraud, H. Denis and M. le Maire, Thesaurin a, the major protein of *Xenopus laevis* previtellogenic oocytes, present in the 42 S particles, is homologous to elongation factor EF-1α (223) 232
Vihko, P. see L. Keso (215) 105
Vile, G.F. and C.C. Winterbourn, Iron binding to microsomes and liposomes in relation to lipid peroxidation (215) 151
Vilgrain, I. see J. Fauvel (221) 397
Villa, T.G. see A.R. Nebreda (220) 27
Villanueva, J.R. see A.R. Nebreda (220) 27
Ville, A.L. see J.D. Bell (219) 239
Villee, C.A. see S. Ishiura (215) 195
Vince, G.S. and R.T. Dean, Is enhanced free radical flux associated with increased intracellular proteolysis? (216) 253
Vincent, J.-P. see J.-C. Bozou (211) 151
Viratelle, O. see H. Valeins (226) 331
Virgili, M. see G. Venturoli (219) 477
Virtanen, I. see O. Närvänen (224) 156
Visser, A.J.W.G., A. van Hoek, T. Kulinski and J. Le Gall, Time-resolved fluorescence studies of flavodoxin: Demonstration of picosecond fluorescence lifetimes of FMN in *Desulfovibrio* flavodoxins (224) 406
Visser, A.J.W.G. see A. de Kok (218) 135
Visser, A. see P.H. Steenbergh (209) 97
Visser, F.R. see D.R. Biggs (220) 223
Viswanatha, T. see A.P. Jahagirdar (219) 83
Viswanatha, T. see E.W. Szczepan (211) 239
Vita, N. see P. Ferrara (226) 47
Vize, P.D. and J.R.E. Wells, Spacer alterations which increase the expression of porcine growth hormone in *E. coli* (213) 155
Vliegenthart, J.F.G. see J.A. van Kuik (221) 150
Voeikov, V.L. see Yu.A. Ovchinnikov (226) 91
Vogel, D.W. see R.K. Hartmann (218) 215
Vogel, U.S. and R.J. Thompson, Molecular cloning of the myelin specific enzyme 2′,3′-cyclic-nucleotide 3′-phosphohydrolase (218) 261
Vogt, B. see H. Bartels (221) 277
Volker, T. see H. Valeins (226) 331
Volkov, N.I. see A.Yu. Andreyev (226) 265
Volpe, P. see A. Maurer (224) 89
Volpe, P. see C. Delfini (210) 17
Volsky, D.J. see S. Dewhurst (213) 133
Volsky, D.J. see S. Dewhurst (213) 138
Von Bahr-Lindström, H. see O.U. Beg (216) 270
Von Heijne, G. and J.P. Segrest, The leader peptides from bacteriorhodopsin and halorhodopsin are potential membrane-spanning amphipathic helices (213) 238
Von Holt, C. see G.P. Sabbatini (224) 117
Von Holt, C. see P.J. Smith (215) 305
Von Jagow, G. see H. Schägger (219) 161
Von Janta-Lipinsky, M. see N. Dyatkina (219) 151
Voorter, C.E.M., E.S. Roersma, H. Bloemendal and W.W. de Jong, Age-dependent deamidation of chicken αA-crystallin (221) 249
Vranken, A.M., E.J.M. Van Damme, A.K. Allen and W.J. Peumans, Purification and properties of an *N*-acetylgalactosamine specific lectin from the plant pathogenic fungus *Rhizoctonia solani* (216) 67
Vsevolodov, N.N. see E.B. Okon (216) 241
Vu, Q.A., D. Zhang, Z.C. Chroneos and D.A. Nelson, Polyamines inhibit the yeast histone deacetylase (220) 79
Vulfson, E.N. see I.A. Smirnova (214) 343
Vuorio, E. see V.-M. Kähäri (215) 331
Vuust, J. see T. Lund (208) 369
Vygodina, T. and A.A. Konstantinov, Evidence for two H_2O_2-binding sites in ferric cytochrome *c* oxidase: Indication to the O-cycle? (219) 387

W

Wachi, M. see M.D. Song (221) 167
Wada, A. see R. Hanai (226) 247
Wada, K. and T. Tanabe, N-terminal amino acid sequences of the heavy and light chains of chicken liver cathepsin L (209) 330
Wada, K. see H. Oh-oka (218) 52
Wada, K. see T. Takai (212) 98
Waelbroeck, M., J. Camus, M. Tastenoy and J. Christophe, 80% of muscarinic receptors expressed by the NB-OK 1 human neuroblastoma cell line show high affinity for pirenzepine and are comparable to rat hippocampus M1 receptors (226) 287
Wagner, A.F.V. see J. Hönes (212) 193
Wagner, H. see R.V. Guntaka (221) 332
Wagner, J.-C. and D.H. Wolf, Hormone (pheromone) processing enzymes in yeast: The carboxy-terminal processing enzyme of the mating pheromone α-factor, carboxypeptidase yscα, is absent in α-factor maturation-defective *kex1* mutant cells (221) 423
Wagner, J.-C., C. Escher and D.H. Wolf, Some characteristics of hormone (pheromone) processing enzymes in yeast (218) 31
Wagner, R., N.J.P. Ryba and R. Uhl, The amplified P-signal, an extremely photosensitive light scattering signal from rod outer segments, which is not affected by pre-activation of phosphodiesterase with Gα-GTP-γ-S (221) 253
Wagner, T., M. Gross and P.B. Sigler, CpA containing oligoribonucleotides specifically inhibit protein synthesis in rabbit reticulocytes (212) 317
Wagter, R. see W. Homan (215) 323
Wakefield, I. see C.E. Smith (225) 247
Wakelam, M.J.O., S. Patterson and M.R. Hanley, L6 skeletal muscle cells have functional V_1-vasopressin receptors coupled to stimulated inositol phospholipid metabolism (210) 181
Waki, M. see S. Ono (220) 332
Waki, M. see Y. Shimohigashi (222) 251
Walaas, O. see S.I. Walaas (220) 311
Walaas, S.I., R.S. Horn, A. Adler, K.A. Albert and O. Walaas, Insulin increases membrane protein kinase C activity in rat diaphragm (220) 311
Walker, G.H. and S.C. Huber, Spinach leaf 6-phosphofructo-2-kinase: Isolation of a new enzyme form that undergoes ATP-dependent modification (213) 375
Wallace, P.G. see M.N. Berry (224) 201
Wallace, R., E. Knecht and S. Grisolía, Turnover of rat liver ornithine transcarbamylase (208) 427

Waller, J.-P. see M. Lazard (216) 27
Walsh, F.S. see J. Thompson (219) 135
Walsh, K.A. see D.R. Eyre (220) 337
Walsh, K.A. see L.H. Ericsson (218) 11
Walsh, K.A. see R.M. Metrione (218) 59
Walsh, K.A. see T. Marti (219) 415
Wang, C.-S., K. Shastri, L. Wen, J.-K. Huang, B. Sonthayanon, S. Muthukrishnan and G.R. Reeck, Heterogeneity in cDNA clones encoding rice glutelin (222) 135
Wang, J. see T. Minegishi (214) 139
Wanka, F. see A.C.M. Pieck (212) 276
Ward, D.J. see C.A. Morrison (214) 65
Warden, J.T. see G.P. Palace (215) 58
Wardlaw, J.R., W.H. Sawyer and K.P. Ghiggino, Vertical fluctuations of phospholipid acyl chains in bilayers (223) 20
Waring, M.J. see J. Portugal (225) 195
Warren, M.J. see P.M. Jordan (225) 87
Warren, R.A.J. see M.L. Langsford (225) 163
Warren, R.M. see P.J. Smith (215) 305
Wasenius, V.-M., O. Närvänen, V.-P. Lehto and M. Saraste, α-Actinin and spectrin have common structural domains (221) 73
Wasenius, V.-M. see O. Närvänen (224) 156
Wasil, M. see B. Halliwell (213) 15
Watanabe, T. see T. Hiyama (214) 97
Watanabe, T.X. see Y. Hirata (219) 369
Watanabe, Y., T. Meshi and Y. Okada, Infection of tobacco protoplasts with in vitro transcribed tobacco mosaic virus RNA using an improved electroporation method (219) 65
Waterhouse, R.N., D. Boulter and J.A. Gatehouse, An organ-specific hypomethylation of cotyledon genomic rDNA in *Pisum sativum* L. (209) 223
Watsfeldt, P. see G. Larson (214) 41
Watson, G.J. see E.J. Wawrzynczak (219) 51
Watson, G.J. see M. O'Hare (216) 73
Watson, H.C. see J.A. Littlechild (225) 123
Watson, I.D. see J. Reglinski (214) 351
Watson, M.D. see J.N. Yarwood (222) 175
Watt, P.W. and M.J. North, The stress-shock response of the bacterium *Methylophilus methylotrophus* (215) 295
Watts, A. see C.E. Dempsey (218) 173
Wawrzynczak, E.J., A. Falasca, W.A. Jeffery, G.J. Watson and P.E. Thorpe, Identification of a tyrosine residue in the saccharide binding site of ricin B-chain using *N*-[^{14}C]acetylimidazole (219) 51
Weakland, L.L. see J.S. Davis (208) 287
Webb, A. see K.H. Cheeseman (209) 191
Weber, K. see D. Fürst (224) 49
Weber, K. see N. Geisler (221) 403
Weber, N. and H.K. Mangold, Radioactively labeled ether lipids by biotransformation of symmetrical alkylglycerols in cell suspension cultures of rape (211) 225
Weber, N. see H.K. Mangold (220) 220
Wechsler, T.D., R.A. Brunisholz, G. Frank, F. Suter and H. Zuber, The complete amino acid sequence of the antenna polypeptide B806–866-β from the cytoplasmic membrane of the green bacterium *Chloroflexus aurantiacus* (210) 189
Weetman, A.P., Recombinant γ-interferon stimulates iodide uptake and cyclic AMP production by the FTRL$_5$ thyroid cell line (221) 91
Weetman, A.P., T.B. Nutman, J.R. Baker, jr and K.D. Burman, Demonstration by immunoblotting of heterogeneity in the autoantibody response directed against fat cells in Graves' disease (211) 69
Wefers, H., D. Schulte-Frohlinde and H. Sies, Loss of transforming activity of plasmid DNA (pBR322) in *E. coli* caused by singlet molecular oxygen (211) 49
Weglicki, W.B. see C.M. Arroyo (221) 101
Weich, H.A., D. Herbst, H.U. Schairer and J. Hoppe, Platelet-derived growth factor: Phorbol ester induces the expression of the B-chain but not of the A-chain in HEL cells (213) 89
Weich, H.A. see J. Hoppe (223) 243
Weiner, L.M. see D.G. Sushkov (225) 139
Weiner, L.M. see Ya.Yu. Woldman (212) 53
Weingarten, D.P. see S. Kumar (208) 151
Weinmann, R. see A. Taylor (226) 109
Weinstein, I.B. see C.A. O'Brian (214) 339
Weintraub, H. see C. de Paillerets (219) 113
Welch, G.R. see M.N. Berry (224) 201
Welinder, K.G. see P.A. Andreasen (209) 213
Welle, R. see D.R. Biggs (220) 223
Weller, U. see K. Aktories (212) 109
Wells, J.R.E. see P.D. Vize (213) 155
Wen, L. see C.-S. Wang (222) 135
Weng, S. see C. Yang (224) 261
Wenzl, S. see R. Schlipfenbacher (209) 57
West, L.A. see J.S. Davis (208) 287
Westphal, M., A. Müller-Taubenberger, A. Noegel and G. Gerisch, Transcript regulation and carboxyterminal extension of ubiquitin in *Dictyostelium discoideum* (209) 92
Wettenhall, R.E.H. see W. Kudlicki (215) 16
Wever, R., E. de Boer, H. Plat and B.E. Krenn, Vanadium – an element involved in the biosynthesis of halogenated compounds and nitrogen fixation (216) 1
Whipple, D.E. see G.M. Olins (224) 325
White, R.D. see R. Wootton (209) 129
Whiteside, S.G. and D.J. Plocke, Identification and isolation of core histones from *Schizosaccharomyces pombe* (226) 250
Whitford, T. see P.S. Fink (214) 75
Whitford, T. see P.S. Fink (220) 263
Whiting, P. and J. Lindstrom, Affinity labelling of neuronal acetylcholine receptors localizes acetylcholine-binding sites to their β-subunits (213) 55
Whiting, P., F. Esch, S. Shimasaki and J. Lindstrom, Neuronal nicotinic acetylcholine receptor β-subunit is coded for by the cDNA clone α$_4$ (219) 459
Wickremasinghe, R.G., A.R. Mire-Sluis and A.V. Hoffbrand, Interleukin-2 binding to activated human T lymphocytes triggers generation of cyclic AMP but not of inositol phosphates (220) 52
Wickremasinghe, R.G. see B.S. Hall (223) 6
Widdows, K.B., E.F. Kirkness and A.J. Turner, Modification of the GABA/benzodiazepine receptor with the arginine reagent, 2,3-butanedione (222) 125
Wiemken, A. see T. Hottiger (220) 113
Wiener, E.C., M.C. Griffor and A. Scarpa, Antibody-induced cAMP accumulation in splenocytes from athymic nude mice (224) 33
Wiesner, R. see H. Kühn (208) 248
Wiesner, W., K.-H. van Pee and F. Lingens, Detection of a new chloroperoxidase in *Pseudomonas pyrrocinia* (209) 321
Wiggert, B. see T. van Veen (208) 133
Wigglesworth, N.M. see S. Kellie (213) 428
Wikieł, H. and J. Strosznajder, Phosphatidylinositol degradation in ischemic brain specifically activated by synaptosomal enzymes (216) 57
Wilcox, M. see K. Gull (219) 31
Wild, F. see E. Malvoisin (215) 175
Wildner, G.F. see U. Johanningmeier (211) 221
Wilkins, M.B. see G.A. Nimmo (213) 18
Wilkinson, M.C. see B.N. Zaba (213) 49
Willems, H. see F. Engels (209) 249
Willhardt, I. see E. Kojro (212) 83
Williams, B.A. see S.C. Huang (216) 128
Williams, H.D. see R.K. Poole (217) 49

Williams, R.J.P., P. Esnouf, M. Lawrence and S.A. Cederholm-Williams, The similarities and differences in structures between kringle 1 of prothrombin and kringle 4 of plasminogen (209) 111
Williams, R.J.P., The mechanism of cytochrome oxidase and other reaction centres for electron/proton pumping (226) 1
Williams, R.S., J.F. Allen, A.P.R. Brain and R.J. Ellis, Effect of Mg^{2+} on excitation energy transfer between LHC II and LHC I in a chlorophyll-protein complex (225) 59
Williams, S.R. see A.E.G. Cass (220) 353
Williamson, J.R. see C.V. Nicchitta (209) 243
Williamson, J.R. see S.K. Joseph (219) 125
Williamson, K.C. see B.F. Dickey (219) 289
Williamson, P., R. Antia and R.A. Schlegel, Maintenance of membrane phospholipid asymmetry: Lipid-cytoskeletal interactions or lipid pump? (219) 316
Wilson, D.F. see A. Pastuszko (218) 189
Wilson, E. see E. Deli (221) 365
Wilson, K.S. see K. Petratos (218) 209
Wilson, P.B. see S.B. Bocckino (225) 201
Winand, J. see J.-P. Dehaye (219) 451
Wingfield, P., P. Graber, N.R. Movva, A.M. Gronenborn, G.M. Clore and H.R. MacDonald, N-terminal-methionylated interleukin-1β has reduced receptor-binding affinity (215) 160
Wingfield, P., R.H. Pain and S. Craig, Tumour necrosis factor is a compact trimer (211) 179
Wingfield, P. see H.R. MacDonald (209) 295
Wingfield, P. see P. Moonen (226) 314
Winterbourn, C.C. see G.F. Vile (215) 151
Winterhalter, K.H. see T. Schreier (213) 319
Wion, D. see P.A. Robinson (209) 203
Wirth, M. see R. Rümbeli (221) 1
Wise, J.G., B.J. Hicke and P.D. Boyer, Catalytic and noncatalytic nucleotide binding sites of the *Escherichia coli* F_1 ATPase: Amino acid sequences of β-subunit tryptic peptides labeled with 2-azido-ATP (223) 395
Wise, P.J., C.J. Danpure and P.R. Jennings, Immunological heterogeneity of hepatic alanine:glyoxylate aminotransferase in primary hyperoxaluria type 1 (222) 17
Wistow, G. see D. Carper (220) 209
Witke, W. see A. Noegel (221) 391
Witt, H.T. see E.J. Boekema (217) 283
Witt, H.T. see I. Witt (221) 260
Witt, H.T. see M. Rögner (219) 207
Witt, H.T. see S. Gerken (223) 376
Witt, I., H.T. Witt, S. Gerken, W. Saenger, J.P. Dekker and M. Rögner, Crystallization of reaction center I of photosynthesis: Low-concentration crystallization of photoactive protein complexes from the cyanobacterium *Synechococcus* sp. (221) 260
Witt, I. see E.J. Boekema (217) 283
Wittmann, H.G. see J. Piefke (209) 104
Wittmann, H.G. see M. Shoham (208) 321
Witzemann, V., B. Barg, Y. Nishikawa, B. Sakmann and S. Numa, Differential regulation of muscle acetylcholine receptor γ- and ϵ-subunit mRNAs (223) 104
Wohlfeil, S. see G. Theiss (218) 159
Woldman, Ya.Yu., L.M. Weiner, L.F. Gulayeva and V.V. Lyakhovich, NMR study of the interaction of P-450 with 4-methoxypyridine (212) 53
Wolf, A., T.M. Fritzsche, B. Rudy and W.E. Trommer, Synthesis of spin-labeled photoaffinity derivatives of NAD^+ and their interaction with lactate dehydrogenase (212) 203
Wolf, C.R. see K. Kelly (222) 120
Wolf, D.H. see J.-C. Wagner (218) 31
Wolf, D.H. see J.-C. Wagner (221) 423
Wolfe, R.S. see D. Ankel-Fuchs (213) 123
Wolfson, A.D. see Y.A. Motorin (220) 363
Wollman, F.A. see J. Olive (208) 308
Wong, E.H.A., J.A. Smith and L. Jarett, Adenosine and oxytocin reverse antagonism of cyclic AMP elevating agents to insulin activation of adipocyte pyruvate dehydrogenase (213) 419
Wong, K. see J. Ginsberg (226) 223
Wong, S.S. see A. Chen (214) 192
Wonnacott, S. see C. Rapier (212) 292
Wonnacott, S. see D.R.E. Macallan (226) 357
Wonnacott, S. see Y. Aracava (222) 63
Woolley, G.A., M.K. Kapral and C.M. Deber, Potential-sensitive membrane association of a fluorescent dye (224) 337
Woolley, P. see B. Epe (213) 443
Wootton, R., D.K. Stammers and R.D. White, Reduction in potency and reversal of left-shifting activity of BW12C with the major and minor components of chicken hemoglobin (209) 129
Worthington, J., C.-T.J. Chan and P.G.H. Byfield, Novel separation method for serum immunoglobulins: Application to thyroid related antibodies (211) 123
Wouters, W., J. Van Dun and P.M. Laduron, Identification of the serotonin-S_2 receptor ligand binding site by photoaffinity labelling with 7-azido-8-[^{125}I]ketanserin ([^{125}I]AZIK) (213) 359
Wren, B.W., C.L. Clayton, P.P. Mullany and S. Tabaqchali, Molecular cloning and expression of *Clostridium difficile* toxin A in *Escherichia coli* K12 (225) 82
Wright, P.E. see C. Dalvit (213) 289
Wright, P.E. see W.J. Chazin (222) 109
Wrotek, M. see Yu.S. Borovikov (223) 409
Wu, J.-J. see D.R. Eyre (220) 337
Wu, S.X. see Y.X. Zhu (208) 253
Wun, T.-C. and K.K. Kretzmer, cDNA cloning and expression in *E. coli* of a plasminogen activator inhibitor (PAI) related to a PAI produced by Hep G2 hepatoma cell (210) 11
Wijermans, P.W. see A.H.L. Koenderman (222) 42

X

Xanthopoulos, K.G. see D.-A. Lidholm (226) 8
Xavier, A.V. see G.D. Fauque (215) 63
Xue, Z., C.G. Miller, J.-M. Zhou and P.D. Boyer, Catalytic and noncatalytic nucleotide binding sites of chloroplast F_1 ATPase: Photoaffinity labeling and peptide sequencing (223) 391

Y

Yaffe, M.B. see H. Sternlicht (214) 226
Yagi, K. see M. Ikura (219) 17
Yagi, T. see J.P. Gayda (217) 57
Yahara, I. see S. Ohno (222) 279
Yamada, H. see J.M. Kim (210) 77
Yamada, K.M. see M. Obara (213) 261
Yamada, K. see M. Tanokura (209) 77
Yamada, S. see T. Hayashi (218) 200

Yamada, T. see N. Takasu (225) 43
Yamada, Y. see Y. Seino (223) 74
Yamagishi, R., T. Koide and N. Sakuragawa, Binding of heparin or dermatan sulfate to thrombin is essential for the sulfated polysaccharide-accelerated inhibition of thrombin by heparin cofactor II (225) 109
Yamaguchi, A., H. Adachi and T. Sawai, Identification of the active site of *Citrobacter freundii* β-lactamase using dansyl-penicillin (218) 126
Yamaguchi, A., M. Yanai, N. Tomiyama and T. Sawai, Effects of magnesium and sodium ions on the outer membrane permeability of cephalosporins in *Escherichia coli* (208) 43
Yamaguchi, A., N. Yamamoto, H. Kitagawa, K. Tanoue and H. Yamazaki, Ca^{2+} influx mediated through the GPIIb/IIIa complex during platelet activation (225) 228
Yamaguchi, J., H. Mori and M. Nishimura, Biosynthesis and intracellular transport of glyoxysomal malate dehydrogenase in germinating pumpkin cotyledons (213) 329
Yamaguchi, T. see A. Tomoda (219) 472
Yamaguchi-Shinozaki, K., K. Shinozaki and M. Sugiura, Processing of precursor tRNAs in a chloroplast lysate: Processing of the 5′-end involves endonucleolytic cleavage by an RNase P-like enzyme and precedes 3′-end maturation (215) 132
Yamakuni, T., F. Ozawa, F. Hishinuma, R. Kuwano, Y. Takahashi and T. Amano, Expression of β-nerve growth factor mRNA in rat glioma cells and astrocytes from rat brain (223) 117
Yamakura, F. see T. Isobe (223) 92
Yamamoto, K. see T. Hayashi (218) 200
Yamamoto, K. see T. Tanimoto (226) 291
Yamamoto, K., The binding of skeletal muscle C-protein to regulated actin (208) 123
Yamamoto, N. see A. Yamaguchi (225) 228
Yamamoto, N. see M. Ohshima (225) 243
Yamamoto, S. see N. Sasakawa (223) 413
Yamamoto, T. see H. Tokuda (215) 335
Yamamoto, T. see K. Kariya (217) 69
Yamamoto, T. see N. Tomita (225) 113
Yamamoto, T., Y. Furuki, J.W. Kebabian and M. Spatz, α-Melanocyte-stimulating hormone secretion from permeabilized intermediate lobe cells of rat pituitary gland: The role of guanine nucleotides (219) 326
Yamamoto, Y., Assignment of hyperfine shifted haem methyl carbon resonances in paramagnetic low-spin met-cyano complex of sperm whale myoglobin (222) 115
Yamamoto, Yasusi see Y. Isogai (224)
Yamamoto, Yasuyuki see Y. Isogai (224) 71
Yamamura, H. see A. Takeda (210) 169
Yamasaki, T. see H. Nakajima (223) 113
Yamashina, I. see A. Kurosaka (215) 137
Yamashita, I. see T. Itoh (219) 339
Yamashita, S. see T. Shinozawa (219) 293
Yamashita, T. see T. Tsuda (208) 39
Yamauchi, F. see H. Kagawa (226) 145
Yamauchi, T. see T. Ichimura (219) 79
Yamaya, H. see K. Kuwajima (221) 115
Yamazaki, H. see A. Yamaguchi (225) 228
Yamazaki, I. see M. Mimuro (213) 119
Yamazaki, I. see S. Hashimoto (208) 305
Yamazaki, M., C. Ichihara and S. Nagasawa, Evidence that a nicked C4b, C4b′, is a functionally active C4b derivative (208) 147
Yamazaki, T. see M. Mimuro (213) 119
Yanai, M. see A. Yamaguchi (208) 43
Yang, C., Z.-W. Gu, W. Patsch, S. Weng, T.W. Kim and L. Chan, The complete amino acid sequence of proapolipoprotein A-I of chicken high density lipoproteins (224) 261
Yano, H. see Y. Seino (223) 74
Yano, J. see H. Hayashi (223) 267
Yao, M. see Y. Kubota (212) 159
Yapo, A. see J.-P. Tenu (220) 93
Yartzev, A.P. see S.V. Chekalin (216) 245
Yarwood, A. see M. Richardson (216) 145
Yarwood, J.N., N. Harris, A. Delauney, R.R.D. Croy, J.A. Gatehouse, M.D. Watson and D. Boulter, Construction of a hybrid cDNA encoding a major legumin precursor polypeptide and its expression and localization in *Saccharomyces cerevisiae* (222) 175
Yasuda, H. see H. Kawaguchi (221) 305
Yatani, A. see J. Codina (216) 104
Yates, C.M. see J. Simpson (217) 62
Yaxley, J.C. see J.D. Norton (215) 127
Yazawa, M. see M. Ikura (219) 17
Yeaman, S.J. see A.P. Bradford (222) 211
Yee, D.K. see A. Pastuszko (218) 189
Yialouris, P.P. see A.A. Haritos (218) 107
Yokoyama, K. see N. Murazumi (218) 131
Yokoyama, S. see T. Hiwasa (211) 23
Yonath, A. see J. Piefke (209) 104
Yonath, A. see M. Shoham (208) 321
Yonemura, D. see A. Tomoda (219) 472
Yonetani, T. see C. Balny (221) 349
Yoneyama, Y. see A. Tomoda (219) 472
Yonuschot, G., E.K. Matthews, A.N. Corps and J.C. Metcalfe, Permeabilization of thymocytes by photon activation of erythrosin (213) 401
Yoshida, M. see J. Inoue (209) 187
Yoshida, T. see H. Fukuzawa (220) 61
Yoshikami, D. see D.H. Feldman (214) 295
Yoshimi, H. see Y. Hirata (219) 369
Yoshimoto, T. see S. Ishiura (215) 195
Yoshioka, T. see Y. Kubota (212) 159
Yoshizawa, T. see M. Ariki (225) 255
Young, D.A. see A. Klip (224) 224
Young, J. see H. Desmond (210) 185
Young, M. and J. Cullum, A plausible mechanism for large-scale chromosomal DNA amplification in streptomycetes (212) 10
Young, W.S. iii, Corticotropin-releasing factor mRNA in the hypothalamus is affected differently by drinking saline and by dehydration (208) 158
Yu, Q.S., A. Brossi and J.L. Flippen-Anderson, *S*-configuration of (+)-primaquine by X-ray analysis of the urea obtained with *R*-(+)-1-phenylethylisocyanate (221) 325
Yu, V.C. see F.-J. Klinz (224) 43
Yusupov, M.M. see S.D. Trakhanov (220) 319
Yusupov, M.M. see S.E. Sedelnikova (220) 227

Z

Zaba, B.N., M.C. Wilkinson, D.M. Taylor, T.J. Lewis and D.L. Laidman, Electrochemical characteristics of platinum electrodes coated with cytochrome b_5-phospholipid monolayers (213) 49
Zachowski, A., P. Fellmann, P. Hervé and P.F. Devaux, Labeling of human erthrocyte membrane proteins by photoactivatable radioiodinated phosphatidylcholine and phosphatidylserine: A search for the aminophospholipid translocase (223) 315

Zagranichny, V.E. see Yu.A. Ovchinnikov (223) 169
Zahradka, P., Probing DNA polymerase α with monoclonal antibodies (212) 259
Zaidi, Z.H. see O.U. Beg (216) 270
Zaita, N., K. Torazawa, K. Shinozaki and M. Sugiura, *Trans* splicing in vivo: joining of transcripts from the 'divided' gene for ribosomal protein S12 in the chloroplasts of tobacco (210) 153
Zajac, J.-M., Y. Charnay, J.-M. Soleilhac, N. Sales and B.P. Roques, Enkephalin-degrading enzymes and angiotensin-converting enzyme in human and rat meninges (216) 118
Zakharova, N.I. see V.Z. Paschenko (214) 28
Zalewski, P.D. see J.K. French (212) 242
Zamai, M. see D. Masson (208) 84
Zammit, V.A. see D. Carling (223) 217
Zandberg, J. see P.H. Steenbergh (209) 97
Zanetti, G. see G. Merati (215) 37
Zanotti, F. see F. Guerrieri (213) 67
Zardi, L. see B. Carnemolla (215) 269
Zavoico, G.B. see F. O'Rourke (214) 176
Zayakina, G.V. see V.G. Metelev (226) 232
Zeelen, J.P. see G.D.F. Maessen (223) 181
Zelinka, J. see J. Šimúth (218) 163
Zelinka, J. see S.V. Shlyapnikov (209) 335
Zeman, K.L. see K.R. Parker (211) 35
Zenita, K. see E. Takano (208) 199
Zeuthen, J. see T. Lund (208) 369
Zhang, D. see Q.A. Vu (220) 79
Zhang, H.L. see Y.X. Zhu (208) 253
Zhang, S.Y. see Y.X. Zhu (208) 253
Zhou, J.-M. see Z. Xue (223) 391
Zhu, Y.X., K.L. Hsi, Z.G. Chen, H.L. Zhang, S.X. Wu, S.Y. Zhang, P.F. Fang, S.Y. Guo, Y.S. Kao and K. Tsou, Neo-kyotorphin, an analgesic peptide isolated from human lung carcinoma (208) 253
Ziegler, K., W. Lockau and W. Nitschke, Bound electron acceptors of photosystem I: Evidence against the identity of redox center A_1 with phylloquinone (217) 16
Žilinskienė, V. see B. Kholodenko (223) 247
Zimmet, J. see K.A. Jacobson (225) 97
Zinker, S. see J. Silva (214) 71
Živković, R. see M.M. Kostić (217) 163
Zlatanov, I.V., M. Foley, J. Birmingham and P.B. Garland, Developmental changes in the lateral diffusion of Leydig cell membranes measured by the FRAP method (222) 47
Zon, G. see V. Sklenář (208) 94
Zon, G. see V. Sklenář (216) 249
Zorina, V.V. see L.A. Drachev (209) 316
Zorina, V.V. see L.A. Drachev (226) 139
Zuber, H. see F. Suter (217) 279
Zuber, H. see R. Rümbeli (221) 1
Zuber, H. see T.D. Wechsler (210) 189
Zueva, M.Y. see V.V. Kupriyanov (208) 89
Zupec, M.E. see G.M. Olins (224) 325
Zvelebil, M.J. see M.J.E. Sternberg (218) 231
Zweibaum, A. see M. Rousset (208) 34
Zwingelstein, G. see E. Malvoisin (215) 175

FEBS Letters Volumes 208–226
Master Subject Index

A site, (213) 443
Absolute configuration, (223) 335
Absorbance difference spectroscopy, (223) 376
Absorption spectrum, (218) 102; (221) 221
Accessibility, (211) 185
Acetaldehyde, (216) 113
Acetaldehyde metabolism, (216) 113
Acetate, (213) 209
N-Acetylated sugar, (215) 311
Acetylation, (222) 99; (222) 322
Acetyl-CoA carboxylase, (212) 98; (223) 217
Acetylcholine, (211) 195; (220) 327; (222) 63
Acetylcholine receptor, (208) 236; (213) 55; (221) 172; (222) 79; (223) 104; (226) 235
Acetylcholine release, (213) 337; (219) 219
Acetylcholinesterase, (209) 206; (213) 159; (219) 202; (225) 77; (225) 273
N-Acetylgalactosamine, (216) 67
N-Acetylglucosamine liberation, (218) 131
N-Acetylglucosaminyl pyrophosphorylpolyprenol hydrolase, (218) 131
N-Acetylglucosaminyltransferase III, (222) 42
N-Acetylglucosaminyltransferase IV, (222) 42
N-Acetylglucosaminyltransferase V, (222) 42
N-[^{14}C]Acetylimidazole, (219) 51
Acid phosphatase, (217) 174
Acid proteinase, (219) 437
Acid-base jump, (224) 348
Acidic amino acid transmitter, (213) 61
Acidic peptide, (226) 217
Acridine orange, (215) 47
Acrosin, (226) 38
Acrosome reaction, (218) 247
ACTH receptor, (219) 56
Actin, (208) 357; (208) 423; (210) 173; (211) 31; (213) 102; (213) 433; (222) 305; (222) 311; (225) 48; (225) 269
Actin binding, (210) 177; (221) 73
Actin-binding protein, (219) 306; (221) 391
Actin-bound divalent cation, (217) 89
Actin-bound nucleotide, (217) 89
Actin crosslinking, (217) 237
Actin filament, (208) 123; (213) 184; (213) 433
Actin filament organization, (221) 387
Actin-myosin interaction, (212) 254; (221) 77
Actin polymerization, (213) 102; (213) 316; (219) 40
α-Actinin, (220) 291; (221) 73; (224) 156
β-Actinin, (213) 433
Activation, (208) 23
Activation energy, (221) 349; (225) 255
Activator protein, (219) 79
Active site, (212) 63; (214) 8; (218) 126
Active transport, (208) 143; (209) 325; (224) 251
Acute-phase protein, (215) 311; (218) 11; (221) 18
Acyl carrier protein, (215) 261; (224) 133
Acyl exchange, (220) 323
Acylation, (213) 428
ADP, (208) 58; (212) 213; (221) 270
ADP-ribosylation, (208) 217; (211) 5; (211) 137; (216) 295; (218) 63; (223) 279; (224) 38; (224) 219; (225) 48
ADP-ribosyltransferase, (212) 109
azido-ADP, (209) 157
H^+-ADPase, (215) 47
ADP/ATP carrier, (212) 1
Adaptation, (224) 4
Adenine derivative, (208) 397
Adenine dinucleotide inhibitor, (213) 204
Adenine nucleotide, (208) 105
Adenine nucleotide translocator, (223) 247; (224) 251
Adenosine, (208) 431; (212) 119; (213) 419; (218) 205; (219) 296
Adenosine A_1-receptor, (223) 197
Adenosine agonist, (209) 122
Adenosine deaminase, (208) 431
Adenosine derivative, (225) 97
Adenosine receptor, (209) 122; (215) 203; (225) 97
Adenosylcobalamin, (219) 197
S-Adenosyl-L-methionine, (209) 362
S-Adenosylmethionine, (220) 167
Adenoviral promoter, (219) 181
Adenovirus, (214) 153
Adenylate cyclase, (208) 455; (211) 113; (211) 151; (215) 203; (216) 107; (218) 148; (219) 56; (219) 296; (220) 57; (220) 98; (222) 327; (224) 365; (226) 91
Adenylate cyclase-inhibiting G-protein, (211) 160
Adenylate cyclase-stimulating G-protein, (211) 5
Adenylate kinase, (208) 301
Adenylyl cyclase, (220) 187
Adhesion molecule, (210) 127
Adipocyte, (209) 238
Adipogenic differentiation, (215) 345
Adipose conversion, (218) 47
Adipose tissue, (209) 175
Adjuvant, (220) 93
Adrenal cortex, (216) 107
Adrenalectomy, (208) 158; (223) 223
Adrenergic receptor, (208) 258
α_1-Adrenergic receptor, (212) 149
β-Adrenergic agonist, (211) 5
β-Adrenergic receptor, (214) 323; (226) 331
α-Adrenoceptor, (218) 7
β-Adrenoceptor, (217) 287
Adrenocortex, (209) 311
Aequorin, (211) 44
Affinity chromatography, (208) 278; (211) 123; (216) 211; (217) 180; (224) 71
Affinity labeling, (213) 55; (216) 221; (216) 225
Affinity modification, (217) 111
Age dependence, (221) 249; (222) 47
Aging, (210) 161; (217) 62
α-Agonist, (217) 45
Agonist binding, (214) 253
AIDS, (213) 133; (213) 138; (218) 231
Alanine transport, (223) 315; (225) 6
Alanine:glyoxylate aminotransferase, (222) 17
Albumin, (208) 364; (220) 206; (221) 18
Albumin secretion, (219) 75
Alcohol dehydrogenase, (222) 99; (223) 33
Aldehyde, (226) 101
Aldehyde dehydrogenase, (222) 95; (224) 357
Aldehyde dehydrogenase I, (215) 233
Aldehyde reductase, (220) 209
Aldolase crosslinking, (221) 408
Aldose reductase, (220) 209
Alfalfa mosaic virus, (209) 284; (221) 337
Alginate, (212) 199; (223) 207

Alginate epimerase, (212) 199
Alginate lyase, (212) 199
Aliphatic amidase, (215) 285; (215) 291
Alkali light chain, (211) 31
Alkali metal, (213) 341
Alkaline phosphatase, (212) 289; (220) 163; (225) 273
Alkane-utilizing yeast, (220) 31
Alkyl analog, (218) 35
Alkylacetylglycerophosphocholine, (220) 220
Alkylglycerophosphocholine, (220) 220
Alkylmethylglyceroglucopyranoside, (220) 220
Alkylmethylglycerophosphocholine, (220) 220
C-Alkylputrescine, (219) 380
N-Alkylputrescine, (219) 380
Allantoin, (219) 145
α-Allophycocyanin B, (217) 279
Allopurinol, (213) 23
Allosteric transition, (221) 161
Gm Allotype, (208) 221
Alternative splicing, (209) 97; (223) 138
Alu sequence, (212) 208
Alveolar macrophage, (209) 249
Alzheimer's disease, (224) 361; (226) 28
Alzheimer-type dementia, (217) 62
Amiloride, (212) 123; (214) 187
Amiloride analogue, (225) 72
Amine oxidase, (214) 274
Amino acid analog, (212) 83
Amino acid analysis, (215) 21; (218) 159; (219) 426
Amino acid homeostatis, (216) 287
Amino acid sequence, (208) 364; (208) 435; (209) 213; (209) 330; (209) 335; (210) 185; (210) 189; (211) 59; (212) 1; (213) 438; (214) 301; (215) 16; (215) 100; (216) 145; (216) 270; (217) 279; (218) 11; (218) 17; (218) 27; (218) 59; (218) 126; (218) 167; (219) 405; (219) 415; (220) 43; (220) 337; (221) 1; (221) 415; (222) 79; (222) 83; (222) 99; (222) 135; (222) 341; (223) 69; (223) 169; (223) 232; (224) 65; (224) 372; (225) 188; (226) 8; (226) 33; (226) 91; (226) 96; (226) 145; (226) 176; (226) 280
Amino acid sequence analysis, (211) 127
Amino acid sequence homology, (223) 340
Amino acid sequencing, (214) 57
Amino acid substitution, (211) 221
Amino acid transport, (213) 45; (214) 327
Amino-terminal anchor, (226) 150
9-Aminoacridine fluorescence, (221) 265
Aminoacylase, (219) 27
γ-Aminobutyric acid, (224) 391
δ-Aminolevulinic acid, (220) 231
DL-2-Amino-5-phosphonovalerate, (218) 189
6-Amino-1,2-benzopyrone site, (212) 73
Ammonium assimilation, (223) 37
Ammonium chloride, (219) 130
cyclic AMP, (208) 39; (210) 169; (211) 5; (213) 365; (213) 419; (215) 183; (216) 40; (217) 32; (217) 163; (218) 7; (218) 73; (219) 139; (219) 227; (219) 254; (219) 326; (220) 103; (220) 155; (220) 342; (222) 299; (223) 47; (223) 97; (223) 131; (223) 321; (224) 33; (224) 377; (225) 178; (225) 251
cyclic AMP dependence, (208) 352; (209) 219; (216) 211
cyclic AMP-dependent protein kinase, (221) 309; (222) 279; (222) 305; (224) 322
cyclic-AMP phosphodiesterase, (208) 455
cyclic AMP production, (221) 91
cyclic AMP receptor protein, (219) 189
cyclic AMP stimulation, (220) 366
cyclic AMP synthesis, (226) 331
Amphibian development, (223) 330
Amphipathic helix, (213) 238; (221) 172
Amphipathic peptide, (224) 208
Amphipathic protein, (216) 183
Amphiphilic helix, (215) 261
Amplification mechanism, (212) 10
α-Amylase, (219) 339
Amyloid protein A, (218) 11
Amyloidosis, (218) 11
Analgesic activity, (208) 253
Analgesic peptide, (208) 253
Anatoxin, (222) 63
Anatoxin-a, (226) 357
Androgen, (226) 213
Anerobic respiration, (218) 209
Angiotensin II, (212) 40
Angiotensin-converting enzyme, (216) 118; (221) 305
Angiotensin I-converting enzyme, (220) 247
Animal model, (208) 431
Anion activation, (223) 304
Antenna, (223) 161
Antenna domain, (223) 187
Antenna size, (215) 31
B806–866 Antenna complex, (210) 189
Anthracycline antibiotic, (219) 212
Anthroyloxy fatty acid, (223) 20
Anti-Fcγ receptor antibody, (224) 29
Anti-sickling, (209) 129
Antibiotic, (213) 443
Antibiotic-associated colitis, (213) 249
Antibiotic biosynthesis, (226) 319
Antibiotic-DNA binding, (225) 195
Antibiotic resistance, (219) 343
Antibody, (209) 83; (212) 213; (213) 309; (216) 7; (222) 11; (224) 33; (224) 156
Antibody production, (220) 93
LB-2 Antibody, (210) 127
Antigen, (216) 89
M Antigen, (216) 261
O Antigen structure, (216) 261
T4 Antigen, (213) 133
Antileucoprotease, (224) 14
Antimalarial activity, (214) 291; (223) 77
Antimycin A, (223) 315; (225) 6
Antioxidant, (216) 253
Antipeptide antibody, (209) 357
Antiport, (208) 407
Antiprotease, (219) 37
Antithrombin-III, (219) 431
α_1-Antitrypsin, (216) 79
α_1-Antitrypsin secretion, (216) 79
Antitumor agent, (223) 294
Antiviral activity, (226) 13
Antiviral state, (214) 153
Apamin, (209) 117; (217) 247; (219) 451
Apolipoprotein A-I, (215) 105
Apolipoprotein-binding protein, (215) 105
Apoliprotein B receptor, (216) 275
Apoliprotein E receptor, (216) 275

Apoptosis, (224) 104
Apurinic site, (208) 155
Apyrimidinic site, (208) 155
Arabinogalactan, (209) 57
O-Arabinosylation, (221) 293
Arachidonic acid, (211) 73; (213) 174; (214) 139; (217) 242; (217) 265; (218) 93; (221) 155; (221) 289; (224) 29
Archaebacteria, (216) 217; (219) 269; (225) 277
Arginine-carboxyl interaction, (224) 161
Arginine deiminase, (226) 285
Arginine residue, (222) 125
Arginine-vasopressin, (218) 93
Arginine-vasotocin, (220) 98
Aromatic ring cleavage, (210) 199; (218) 255; (221) 327
Aromatization, (218) 143
Arrestin, (225) 127
Aryl cation radical, (210) 199; (220) 149
β-Aryl ether, (210) 199
Arylsulfatase B, (208) 352
Ascorbic acid, (213) 15
Asparagine, (219) 431
Aspartic protease, (219) 1
Aspartic proteinase, (214) 8
Aspartyl-prolyl deavage, (218) 1
Aspirin, (218) 243
Astrocyte, (211) 73; (213) 133; (213) 138; (223) 117
AT-base specificity, (212) 297
A + T-rich sequence, (221) 3
ATG start codon, (218) 119
ATP, (208) 58; (209) 152; (211) 190; (213) 375; (219) 375; (221) 231; (221) 270; (223) 53
ATP analog, (225) 145
ATP binding, (222) 204
ATP-binding site, (218) 81; (225) 145
ATP citrate-lyase, (224) 322
ATP hydrolysis, (211) 99; (221) 77; (222) 204
ATP-phosphocreatine exchange, (208) 89
ATP release, (213) 337
ATP sensitivity, (215) 9
ATP synthase, (218) 27; (223) 366
ATP synthesis, (221) 265; (222) 163; (223) 315; (225) 6; (220) 358
[α-^{32}P] ATP, (222) 204
azido-ATP, (223) 395
2-azido-ATP, (223) 391
extracellular ATP, (221) 155
F_1F_0-H^+-ATP synthase, (209) 197
Mg^{2+}-ATP, (211) 35
P_i-ATP exchange, (212) 323
ATP/ADP antiporter, (226) 265
ATP/ADP ratio, (212) 114
ATPase, (209) 157; (214) 343; (219) 274; (221) 265; (224) 187; (224) 193; (225) 269
ATPase inhibitor protein, (215) 300
ATPase proteoliposome, (208) 138
Ca^{2+}-ATPase, (217) 32; (224) 89
(Ca^{2+} + Mg^{2+})-ATPase, (224) 331
F_1-ATPase, (208) 1; (209) 152; (212) 63; (213) 333; (218) 222; (219) 15; (222) 32; (223) 304; (223) 309; (223) 391
H^+-ATPase, (212) 323; (213) 67; (213) 381; (219) 321; (220) 366; (223) 309; (223) 395; (224) 348
(Na^+ + K^+)-ATPase, (213) 73; (214) 335; (217) 42; (217) 111; (217) 163; (217) 269; (217) 275; (220) 84; (221) 129; (222) 163; (224) 83; (225) 27; (225) 145
ATP,Mg-dependent phosphatase, (224) 385
ATP,Mg-dependent protein phosphatase, (216) 291
Atherosclerosis, (224) 261
Atrial natriuretic factor, (208) 263; (219) 375; (220) 214; (224) 396
Atrial natriuretic factor secretion, (214) 17
Atrial natriuretic peptide, (218) 159; (219) 369; (224) 325; (225) 251
Atypic ribonucleotide, (219) 464
Autoantibody, (211) 69
Autoimmunity, (215) 252
Autophosphorylation, (208) 451
Autoradiography, (216) 118
Avidin, (208) 278
Axon, (221) 403
7-Azido-8-ketanserin, (213) 359
Azurin, (212) 168

B-cell line, (219) 311
B2 sequence, (219) 400
Ba^{2+} effect, (219) 351
Bacitracin, (226) 319
Bacterial expression vector, (220) 47
Bacterial haloperoxidase, (209) 321
Bacterial photosynthesis, (209) 37; (218) 283; (219) 477; (223) 161
Bacterial receptor, (214) 41
Bacterial vaccine, (208) 194
Bacteriochlorophyll, (216) 245; (219) 477
Bacteriophage, (221) 145
Bacteriopheophytin, (216) 245
Bacteriorhodopsin, (208) 337; (209) 44; (209) 316; (213) 238; (213) 283; (216) 155; (218) 287; (222) 275; (226) 139
Barley stripe mosaic virus, (214) 313
Basement membrane, (208) 203; (212) 49; (216) 281; (224) 297; (225) 188
Basic 7 S globulin, (226) 145
BCECF, (222) 144
Becker muscular dystrophy, (212) 313
Benzo[a]pyrene, (209) 269
Benzodiazepine, (219) 301
Benzodiazepine receptor, (222) 125
2′,3′-O-(4-Benzoyl)benzoyl-ADP, (209) 157
Bicarbonate, (218) 148
Bicarbonate effect, (226) 347
Bifunctional enzyme, (217) 212
Bifunctional protein, (226) 364
Bilayer dynamics, (223) 20
Bile acid, (213) 411
Bile salt-stimulated lipase, (217) 37
Bimodal regulation, (213) 195
Binding inhibition, (208) 273
Binding site, (219) 301
Biological activity, (225) 251
Bioluminescence, (224) 348
Biotin, (208) 278; (215) 305
Biotin analog, (215) 305
Biotin-binding site, (212) 98
Biotin enzyme, (220) 121
Biotinyl-UTP, (215) 305
Biotinylglycyltyrosine, (215) 305

Biotinylinsulin, (215) 305
Biotransformation, (220) 353
Biphasic association kinetics, (219) 103
Biphasic dissociation kinetics, (219) 103
2,3-Bisphosphoglycerate, (221) 231
Black lipid membrane, (224) 83
Blastula, (218) 205
Blebbing, (209) 139
Bleomycin assay, (214) 362
Blood coagulation, (220) 129
Blood group, (214) 41
Blue bacteriorhodopsin, (213) 39
Bohr effect, (221) 161
Bolton-Hunter reagent, (224) 198
Bone-resorbing activity, (226) 58
Botulinum ADP-ribosyltransferase C3, (224) 38
Botulinum C2 toxin, (212) 109
Botulinum neurotoxin, (216) 295; (219) 219
Botulinum neurotoxin C1, (224) 38
Botulinum neurotoxin D, (224) 38
Bradykinin, (214) 365; (219) 441; (220) 302; (222) 27
Branched-chain ketoacid dehydrogenase complex, (215) 13
Branched-chain ketoacid dehydrogenase kinase, (215) 13
Brefeldin A, (214) 135
Brevetoxin, (219) 355
8-Bromo cyclic GMP, (221) 284
Brown fat, (208) 128
Brunescent cataractous lens, (219) 472
Brush border enzyme, (211) 133
Brush border membrane, (212) 154
Brush border myosin, (223) 262
BSF-2, (221) 18
Bumping, (212) 21
α-Bungarotoxin, (222) 79; (224) 277; (226) 357
α-Bungarotoxin binding, (214) 253; (222) 56
t-Butyl hydroperoxide, (209) 305
n-Butyrate, (220) 79
BW12C, (209) 129

Clr, (222) 129
C2 toxin, (225) 48
C3 cinvertase, (298) 147
C4, (213) 415
nicked C4b, (208) 147
C-peptide, (208) 445; (212) 307
C-reactive protein, (211) 165
C-terminal localization, (213) 29
C-terminal peptide, (212) 163
C-terminal propeptide, (212) 229
C-terminus, (226) 166
C_4 photosynthesis, (213) 18; (219) 331; (219) 347
Ca^{2+}, (208) 39; (208) 47; (208) 51; (208) 99; (208) 287; (208) 423; (209) 1; (209) 169; (210) 132; (211) 41; (211) 44; (212) 242; (214) 139; (214) 176; (214) 331; (214) 365; (215) 279; (216) 35; (216) 45; (217) 117; (218) 113; (218) 143; (219) 83; (219) 119; (219) 125; (221) 23; (221) 33; (222) 144; (222) 317; (222) 349; (223) 47; (225) 16; (225) 43; (226) 67; (226) 307
Ca^{2+}-activated K^+ channel, (225) 173
Ca^{2+}-activated neutral protease, (215) 274
Ca^{2+}-activated potassium channel, (219) 451
Ca^{2+} activation, (209) 117; (209) 165; (216) 211; (217) 247
Ca^{2+} binding, (209) 77; (223) 405
Ca^{2+}-binding protein, (214) 21; (215) 261; (218) 167; (221) 397
Ca^{2+}-binding site, (218) 97
Ca^{2+} calmodulin-dependent protein kinase, (221) 309
Ca^{2+}/calmodulin-dependent protein kinase, (219) 79
Ca^{2+} channel, (212) 127; (212) 247; (214) 295; (214) 347; (219) 351; (221) 28; (221) 95; (225) 247
Ca^{2+} channel blocker, (220) 103
Ca^{2+} dependence, (213) 184; (214) 181
Ca^{2+}-dependent phospholipid binding, (223) 267
Ca^{2+}-dependent protease, (220) 271
Ca^{2+} effect, (214) 35
Ca^{2+} efflux, (210) 132
Ca^{2+} influx, (211) 195; (216) 40; (224) 4; (225) 247
Ca^{2+} mobilization, (212) 123; (213) 95; (221) 289
Ca^{2+}-mobilizing hormone, (225) 201
Ca^{2+} phospholipid-dependent protein kinase, (217) 232
Ca^{2+} regulation, (223) 409
Ca^{2+} release, (208) 283; (209) 243; (217) 85; (218) 153; (221) 119; (222) 149
Ca^{2+} sensitivity, (217) 11
Ca^{2+} sequestration, (224) 331
Ca^{2+} signal, (225) 72
Ca^{2+} store, (214) 130
Ca^{2+} transient, (223) 53
Ca^{2+} uptake, (211) 41; (213) 337; (218) 189; (221) 119; (223) 413
cytoplasmic Ca^{2+}, (213) 195
cytoplasmic free Ca^{2+}, (221) 28
cytosolic Ca^{2+}, (209) 139; (212) 133
cytosolic free Ca^{2+}, (220) 103; (224) 396
intracellular Ca^{2+}, (212) 40; (215) 183; (219) 40
intracellular free Ca^{2+}, (225) 228
Cadmium, (214) 62
Cadmium-bisphosphoglycerate interaction, (217) 21
Caffeine, (214) 130; (221) 119
Calcitonin, (209) 97; (226) 96
Calcitonin gene, (223) 63
Caldesmon, (218) 139; (218) 292; (219) 306
Calmodulin, (208) 418; (212) 133; (212) 154; (214) 331; (215) 241; (216) 211; (217) 187; (219) 17; (220) 36; (223) 262; (225) 269
Calmodulin-binding protein, (213) 184
Calmodulin dependence, (208) 423; (213) 184
Calmodulin-dependent protein kinase, (219) 249
Calmodulin gene, (217) 187
Calmodulin inhibitor, (216) 211
Calpactin, (223) 267
Calpain, (215) 274
Calpain 1, (216) 22
Calpain 2, (216) 22
Calpastatin, (208) 199; (223) 174
Calycosin, (214) 101
Cancer, (214) 351; (220) 1; (220) 236; (223) 237
Cancer-associated carbohydrate antigen, (215) 137
Cannabis, (211) 119
5′-Cap, (220) 177
Capsular polysaccharide, (221) 145
Carbamoyl-phosphate synthase, (224) 182
Carbamyl phosphate synthetase, (222) 95
Carbamylcholine, (210) 204; (219) 451
Carbene, (221) 408; (226) 134
Carbohydrate attachment, (219) 431
Carbohydrate structure, (224) 97
β-Carboline, (219) 301

Carbon-centered radical, (221) 101
Carbon enrichment, (220) 257
Carbon monoxide, (221) 241
Carbonic anhydrase B, (224) 9
Carboxyatractylate, (226) 265
γ-Carboxyglutamic acid, (222) 353
Carboxyl group, (213) 39
Carboxylase, (222) 353
Carboxylation, (222) 186
Carboxypeptidase A, (212) 83
Carboxypeptidase P digestion, (216) 195
Carboxypeptidase Y, (212) 163
Carboxypeptidase Y digestion, (216) 195
Carboxypeptidase yscα, (218) 31; (221) 423
Carcinoid tumour, (208) 67
Carcinoma, (208) 253
Cardiac enzyme, (214) 192
Cardiodilatin, (225) 251
Cardiotoxin, (218) 17
Carotenoid, (210) 211; (219) 477
Carrier protein, (220) 129
Casein, (218) 55
Casein kinase, (215) 241
Casein kinase II, (220) 143
Casein kinase-1, (224) 385
Casein kinase-2, (224) 385; (226) 109
β-Casein, (212) 163
Catabolic plasmid, (226) 241
Catabolite inactivation, (216) 265
Catalase, (218) 251; (220) 31
Catalase compound I, (215) 228
Catalepsy, (211) 119
Catalytic histidyl residue, (219) 22
Catalytic iron, (214) 362
Catalytic mechanism, (214) 8
Catalytic site structure, (217) 111
Catecholamine, (208) 258; (219) 145; (222) 215
Cathepsin B, (211) 23; (219) 441
Cathepsin H, (226) 33
Cathepsin L, (209) 330; (211) 23; (223) 69
Cathode reduction, (215) 219
Cation binding, (223) 289
Cation channel, (208) 397
Cation depletion, (213) 39
Cation radical, (218) 255
CCB, (224) 142
CCCP, (223) 315; (225) 6
CDP-choline, (214) 149
CDP-diacylglycerol, (212) 159
Cecropin A, (226) 8
Cecropin D, (226) 8
Cell activation, (217) 158
Cell adhesion, (213) 261; (217) 94; (225) 33
Cell culture, (211) 225; (216) 257; (220) 223
Cell cycle, (208) 143; (209) 277; (219) 400; (220) 1; (221) 48; (222) 322; (223) 237; (226) 53
Cell density, (222) 311
Cell division, (209) 191
Cell-free system, (212) 317
Cell growth, (226) 53
Cell harvestor, (214) 285
Cell lineage, (215) 1
Cell nucleus, (210) 165
Cell proliferation, (209) 277; (210) 123; (217) 69; (219) 119; (220) 1; (223) 237
Cell surface, (208) 418
Cell transfection, (219) 181
Cell wall, (220) 27; (220) 383
Cellular adherence, (219) 311
Cellular receptor, (221) 55
Cellulase, (225) 163
Central nervous system, (213) 138; (222) 63
Cephalosporin, (208) 43; (214) 357
Cerebral hemorrhage, (216) 229
CF_1, (219) 321
CF_0 subunit, (218) 27
CF_0F_1, (218) 27
CGRP, (209) 97
CGRP gene, (223) 63
CHAPS, (208) 401
Chain length, (226) 101
Chain melting, (218) 173
B-Chain, (219) 51
γ-Chain, (224) 291
Channel activity, (214) 253; (222) 56
Channeling, (214) 244
Characterization, (215) 140
Charge recombination, (217) 134
Charge separation, (209) 37; (214) 370; (216) 245; (226) 186
Charge transfer, (224) 406
Charybdotoxin, (209) 117; (226) 280
Chelator, (215) 151
Chemical modification, (212) 193; (222) 125; (223) 361
Chemiosmotic coupling, (222) 235
Chemiosmotic hypothesis, (219) 88
Chemiosmotic theory, (209) 325
Chemotactic factor, (221) 95
Chemotactic peptide, (222) 289
Chemotaxis, (211) 17
Chimeric oligonucleotide, (215) 327
Chimeric probe, (226) 232
Chiral deuteration, (223) 191
Chiral fluorinated compound, (220) 353
Chloramine-T, (224) 198
Chloramphenicol acetyltransferase, (219) 70
Chloroperoxidase, (209) 321
Chlorophyll a, (213) 119
Chlorophyll a′, (214) 97
Chlorophyll a/b-light harvesting complex, (215) 25
Chlorophyll a/b-protein, (213) 29
Chlorophyll fluorescence, (211) 99; (221) 211
Chlorophyll protein, (224) 343
Chlorophyll-protein complex, (225) 59; (226) 275
Chlorophyllase, (226) 72
Chloroplast, (209) 157; (209) 181; (210) 153; (212) 114; (213) 453; (218) 27; (219) 343; (219) 393; (220) 61; (221) 211; (221) 265
Chloroplast lysate, (215) 132
Chloroplast membrane, (208) 401; (219) 331; (219) 347
Chloroplast origin, (221) 189
Chloroquine, (216) 51; (223) 251
Chlorpromazine, (222) 21
Chlortetracycline, (218) 97
Cholecystokinin, (211) 234; (220) 327; (226) 67
Cholera toxin, (220) 327; (224) 1; (224) 219

Cholera toxin dependence, (211) 5
Choleraphage φ149, (213) 81
Cholesterol esterification, (220) 371
Cholesterol metabolism, (220) 159
Cholesterol synthesis, (222) 159; (223) 217
Cholinergic synaptosome, (213) 337
Chondroitin sulfate, (209) 299
Chordin, (224) 23
Chromaffin, (211) 44
Chromaffin cell, (208) 357; (226) 337
Chromaffin granule, (215) 339; (222) 215; (222) 358
Chromatin, (208) 31; (213) 293; (215) 237; (222) 322; (226) 88
Chromatin structure, (212) 263; (213) 293; (225) 120
Chromatofocusing, (208) 439
Chromatography, (220) 243
Chromatophore, (219) 88
Chromogranin, (211) 144; (224) 142
Chromolipid, (226) 101
Chromosomal localization, (217) 42
Chromosome banding, (217) 184
Chromosome scaffold, (212) 276
Chronic lymphocytic leukaemia, (215) 127
Chronic myelogenous leukemia, (208) 352
Chronic myeloid leukemia, (222) 293
Chymotrypsin, (217) 101
Chymotrypsin-like enzyme, (225) 168
α-Chymotrypsin, (214) 13; (219) 22
Ciguatera, (219) 355
Ciguatoxin, (219) 355
Circular dichroism, (213) 283; (221) 115; (223) 327
Cirrhosis, (220) 217
Citrate synthase, (225) 277
L-Citrulline synthesis, (226) 285
Cl^- conductance, (217) 247
Cl^- current, (217) 11
Cl^- efflux, (212) 313; (222) 149
Cl^- uptake, (217) 129
Clathrin β-light chain, (220) 143
Cleavage site, (209) 9
Clonidine, (218) 7
Cloning, (212) 208; (213) 438
CNPase, (218) 261
CO_2 accumulation, (209) 28
COSY, (219) 426
Coat protein regulation, (209) 284; (221) 337
Coated vesicle, (219) 113; (220) 143
Coding sequence, (215) 291
Codon alteration, (213) 155
Codon usage, (215) 291; (217) 184; (221) 3
Codon usage bias, (220) 36
Coenzyme F430, (220) 358
Coherence transfer, (219) 45
Colchicine analog, (218) 102
Colchicine-tubulin, (219) 103
Cold acclimation, (208) 128
Cold activation, (212) 213
Colicin, (216) 183
Collagen, (212) 49; (213) 319; (215) 331; (216) 281; (218) 195; (220) 337; (224) 297; (225) 188; (225) 228
Collagen IV, (208) 203
Collagen synthesis, (220) 163
Collagenase, (209) 9
Colonic cancer, (215) 137
Colony-stimulating factor, (222) 341
Comovirus, (215) 247
Compartmentation, (208) 105
Competitive activator, (212) 83
Competitive binding assay, (208) 439
Complement, (208) 147
Complement C3, (214) 135
Complement component C4, (217) 65
Complement precursor accumulation, (214) 135
Complement system, (222) 129
Complementary DNA, (223) 212
Complementary peptide, (224) 272
Complex III, (214) 117
bc_1 Complex, (219) 161
T-p53 Complex formation, (221) 199
Component B, (213) 123; (214) 265; (220) 358
Compound ES, (221) 349
Compound 48/80, (216) 89
Computer graphics model, (208) 11
Computer simulation, (215) 95
Concanavalin A, (215) 209
Concanavalin A-stimulated T-lymphocyte, (224) 59
Conductance, (216) 11
Conformation, (215) 215
Conformation homology, (224) 149
Conformation-limited proteolysis, (217) 180
E_1-Conformation, (217) 111
Conformational change, (212) 213; (216) 183; (223) 289
Conformational restriction, (222) 71
Conformational similarity, (213) 265
Conformational switch, (217) 191
Conjugation, (215) 335; (223) 387
Connective tissue breakdown, (209) 9
Connective tissue formation, (214) 236
Consensus box, (219) 70
Continuous ambulatory peritoneal dialysis, (220) 307
Continuous chromatography, (209) 13
Contractile activity, (219) 469
Control coefficient, (223) 247
Cooperativity, (223) 304
Coordination geometry, (213) 289
Copeptin, (210) 40
Copper, (219) 335; (226) 1
Copper protein, (218) 209
Core histone, (226) 250
Corticotropin, (216) 107
Counterion, (216) 155
Coupling factor, (209) 157; (211) 99
Coupling protein, (216) 104
Covalent flavinylation, (224) 121
Covalent interaction, (217) 101
Covalent modification, (223) 279
CP29, (215) 25
Crambin, (219) 426
Creatine kinase, (208) 89; (214) 192
Creatine phosphate, (221) 270
Critical phenomenon, (217) 212
Cross-linking, (214) 107; (220) 337; (224) 54; (226) 161
Cross-reactivity, (222) 11
Cross-reconstitution, (223) 309
Cryoelectron microscopy, (219) 274

Cryoprotection, (214) 28
Cryoprotective mixture, (215) 215
Crystal growth, (225) 123
Crystal structure, (218) 178; (218) 209
Crystallin, (221) 134; (221) 249
α-Crystallin, (222) 1
γ-Crystallin, (208) 11
ρ-Crystallin, (220) 209
Crystallization, (208) 321; (220) 227; (221) 260
Crystallography, (208) 73; (209) 261
Cu center, (222) 235
Cu loop mechanism, (222) 235
Curare, (224) 277
Cyanelle, (224) 401
Cyanobacteria, (208) 308; (215) 209; (217) 169; (221) 110; (223) 37
Cyclic amine, (222) 75
Cyclic voltammetry, (213) 49
Cyclin, (209) 277
Cycloheximide, (214) 143
Cyclooxygenase, (209) 249; (212) 35
Cystatin, (211) 23
Cystatin C, (216) 229
Cystatin superfamily, (216) 17
Cysteine proteinase, (209) 330; (216) 229; (223) 69; (226) 33
Cysteine proteinase inhibitor, (216) 17
Cytidine 2′-phosphate, (225) 183
Cytochalasin, (208) 357
Cytochrome, (209) 261
Cytochrome a, (213) 396; (226) 343
Cytochrome a_1, (217) 49
Cytochrome a_3, (226) 343
Cytochrome b, (213) 107
Cytochrome b_5, (209) 49; (213) 49; (213) 391
Cytochrome b_6-f, (208) 317; (213) 453; (220) 61; (221) 205; (225) 67
Cytochrome b-558, (214) 279
Cytochrome b-559, (211) 99
Cytochrome b-590, (217) 49
Cytochrome b-595, (217) 49
Cytochrome c, (213) 391; (213) 396; (223) 155; (226) 179; (226) 343
Cytochrome c_3, (217) 57
Cytochrome-c oxidase, (214) 75; (218) 277; (219) 387; (223) 255
Cytochrome c peroxidase, (221) 349
Cytochrome d complex, (217) 49
Cytochrome oxidase, (213) 396; (217) 49; (220) 262; (222) 235; (223) 155; (226) 1; (226) 343
Cytochrome P-448, (209) 305
Cytochrome P-450, (212) 53; (222) 120
Cytochrome P-450 synthesis, (220) 231
Cytoskeleton, (208) 423; (209) 203; (211) 35; (213) 102; (213) 428; (221) 391; (221) 403; (222) 305; (222) 311; (224) 156; (224) 361; (225) 269; (226) 28
Cytosol, (212) 114; (214) 62; (218) 251
Cytotoxicity, (208) 84; (218) 17
Cytotoxin, (218) 17
Cytotoxin-like basic protein, (218) 17

D_2 dopamine receptor, (209) 71
Dansyl-penicillin, (218) 126
DBMIB, (208) 317
DD-peptidase, (225) 218
Deacetyl-cephalosporin C, (225) 218
Deamidation, (221) 249
Decarboxylase, (212) 79
Decay kinetics, (217) 134
Dehydration, (209) 33
Dehydroamino acid, (222) 251
Dehydropeptide, (222) 251
Deletion analysis, (219) 70
Denervation, (223) 104
2-Deoxy-2-[^{18}F]fluoro-D-glucose, (216) 128
3-Deoxy-D-manno-2-octulosonic acid, (221) 145
Dephosphorylation, (210) 177
Depolarization, (222) 149
Dermatan sulfate proteoglycan, (213) 159
Desensitization, (214) 323; (217) 129; (217) 287; (224) 277
Desferrioxamine, (222) 246
Desiccation tolerance, (220) 113
Detergent activation, (223) 304
Detergent fractionation, (211) 94
Detergent solubilization, (214) 285
Detoxication, (224) 357
Deuteration, (218) 173
Development, (208) 460; (209) 223; (214) 45; (214) 259; (217) 260
Dexamethasone, (215) 68; (219) 56; (225) 151
Diacylglycerol, (209) 134; (214) 171; (218) 35; (218) 93; (219) 393; (221) 23; (221) 359
Diamine, (216) 155
Diamine oxidase, (219) 380
Diazirinopyridine, (212) 203
Diazofluorene, (221) 408
Dibasic sequence, (212) 307
Dicistronic transcript, (219) 181
2′,3′-Dideoxy-2′,3′-dehydrothymidine 5′-triphosphate, (219) 151
Dietary protein, (220) 231
Diethyl pyrocarbonate, (219) 301
Difference NOE, (219) 45
Differential gene expression, (219) 31
Differential scanning calorimetry, (209) 257
Differentiation, (211) 64; (211) 160; (215) 1; (215) 127; (215) 175; (218) 195; (219) 56; (221) 81; (224) 43; (225) 53
Differentiation-inducing activity, (226) 58
Differentiation induction, (222) 42
Diffusion, (209) 1
Diffusion potential, (208) 138
Difluorobenzoate metabolism, (220) 353
Dihydrofolate reductase, (217) 106; (218) 178
Dihydropyridine, (212) 127
Dihydropyridine receptor, (212) 127
1α,25-Dihydroxy-22-oxavitamin D_3, (226) 58
8α,25-Dihydroxy-9,10-seco-4,6,10(19)-cholestatrien-3-one, (218) 200
Diisopropyl fluorophosphate, (220) 89
Dimethyl pimelindiimidate, (208) 26
Dioleoylphosphatidic acid, (209) 33
Dioleoylphosphatidylcholine, (209) 33
Dioxygen, (226) 1
1,2-Dipalmitoylphosphatidylethanolamine, (224) 283
Diphenylhexatriene, (223) 381
Diphtheria toxin, (208) 217; (215) 73; (218) 271; (226) 115
Dipyrromethane cofactor, (225) 87
Direct photocrosslinking, (222) 204

Direct sequencing, (219) 343
Disk membrane, (219) 97
Disomal series, (226) 88
Displacement current, (217) 25
Dissimilatory ammonia-forming pathway, (218) 227
Distamycin A, (225) 195
Distamycin analog, (212) 297
Distance geometry, (213) 269
Disulfide bond, (212) 229; (214) 107; (215) 165
Disulfide crosslinking, (212) 254
2,5-Di(tert-butyl)-1,4-benzohydroquinone, (224) 331
Dithiothreitol, (214) 107
Diuron resistance, (208) 208
DNA, (209) 181; (209) 269; (211) 49; (214) 75; (216) 249; (220) 383; (222) 89; (226) 247
DNA binding, (221) 199; (222) 332
DNA cloning, (213) 249
DNA damage, (209) 269; (211) 49; (213) 215; (221) 110
DNA-dye interaction, (215) 95
DNA homology, (214) 75
DNA hybridization, (214) 75
DNA ligase I, (213) 345
DNA ligase II, (213) 345
DNA methylase, (215) 266
DNA methylation, (209) 223; (215) 266
DNA polymerase, (219) 151; (223) 122
DNA polymerase α, (212) 259; (216) 221; (216) 225
DNA polymerase I, (221) 194
DNA polymerase-α, (221) 48
DNA-protein interaction, (222) 332
DNA recognition, (220) 167; (225) 151
DNA repair, (212) 173
DNA repeat, (214) 87
DNA replication, (209) 277; (220) 1; (223) 122
DNA sequence, (219) 339; (220) 31; (221) 167
DNA sequence analysis, (208) 208
DNA structure, (208) 155
DNA synthesis, (220) 288; (224) 59
DNA tumor virus, (215) 345
complementary DNA, (212) 233; (223) 212
extrachromosomal DNA, (208) 386
genomic DNA, (213) 448
left-handed DNA, (213) 341
recombinant DNA, (208) 183
single-stranded DNA-binding protein, (211) 155; (220) 347
Z-DNA, (213) 341; (225) 11
cDNA, (208) 133; (208) 199; (211) 5; (211) 78; (211) 200; (212) 208; (213) 73; (213) 189; (213) 254; (213) 261; (213) 438; (217) 227; (219) 405; (219) 459; (222) 95; (222) 175; (222) 186; (223) 174; (223) 243; (224) 317; (224) 361; (225) 113; (226) 28; (226) 125; (226) 235
cDNA clone, (208) 391; (214) 317; (222) 135; (225) 159; (226) 8
cDNA cloning, (208) 77; (209) 367; (211) 160; (212) 98; (215) 100; (216) 229; (221) 415; (222) 181; (223) 69; (223) 113; (223) 169; (223) 181; (223) 340; (226) 33; (226) 91; (226) 96; (226) 176
cDNA expression, (214) 253; (222) 56; (225) 27
cDNA expression library, (220) 31
cDNA library, (209) 203; (214) 317
cDNA nucleotide sequence, (216) 234
cDNA sequence, (210) 157; (215) 233; (218) 261; (220) 36; (220) 209; (222) 139; (223) 74; (223) 138
kDNA, (213) 304
rDNA, (218) 215
18-28 S rDNA, (209) 340
DNase I, (226) 88
DNase 1, (208) 357
DNP-INT, (208) 317
Domain, (225) 163
Domain structure, (208) 199
Dome formation, (221) 315
Dopamine D_2 receptor, (224) 267
Double-helix stability, (226) 247
Down-regulation, (223) 11
Drug binding, (222) 215
Drug design, (224) 161
Drug-DNA interaction, (212) 297
Drug-enzyme interaction, (218) 178
Duchenne muscular dystrophy, (212) 313; (224) 49
Dysmyelination, (223) 417

Ecdysone, (213) 85
Echinomycin, (215) 266
EcoRI methylase, (216) 207
EcoRI restriction endonuclease, (216) 207
Ectoprotein, (226) 150
Edman degradation, (214) 50
EF-hand structure, (221) 391
EGF, (218) 143
EGF-like domain, (222) 129
EGF receptor, (219) 215
Electrical coupling, (220) 342
Electrical permeabilisation, (219) 139
Electroblotting, (211) 127; (213) 385
Electrochemical potential, (224) 201
Electrochemistry, (213) 49
Electrochromism, (219) 477
Electrogenic property, (224) 83
Electrolyte absorption, (221) 309
Electron acceptor, (211) 211; (217) 16; (220) 74
Electron carrier A_1, (215) 58
Electron donor, (211) 211
Electron gating, (213) 396
Electron microscopy, (217) 283; (219) 207; (220) 319; (221) 3
Electron spin echo, (224) 75
Electron transfer, (214) 92; (215) 63; (215) 219; (218) 209; (219) 477; (223) 376
Electron-transfer flavoprotein-ubiquinone oxidoreductase, (226) 129
Electron transport, (208) 73; (214) 370; (215) 31; (226) 347
Electron transport chain, (226) 77
Electron transport protein, (209) 261
Electronic energy level, (215) 219
Electronic structure, (208) 331
Electrophoresis, (214) 192; (217) 101
Electroporation, (219) 65
Electrotransfer, (212) 220
Ellipticine, (208) 155
Elongation factor, (223) 232
Elongation factor 1β, (208) 77
Elongation factor 1γ, (223) 181
Elongation factor 2, (208) 217; (214) 331
Elongation factor Tu, (208) 189; (217) 203; (224) 54
Elongation rate, (223) 1

Embryonic development, (213) 345
Emery-Dreifuss muscular dystrophy, (212) 313
Emphysema, (208) 183
End group analysis, (216) 195
Endocytosis, (219) 113
Endogenous lectin, (223) 330
Endoglycosidase, (212) 220
Endonexin, (216) 45
Endonuclease, (213) 215
Endonuclease defect, (214) 143
3′-Endonucleolytic cleavage, (215) 132
Endoplasmic reticulum, (218) 63; (218) 153
ENDOR, (214) 117
δ-Endotoxin, (226) 364
Energy calculation, (213) 265
Energy coupling, (219) 88; (219) 274
Energy metabolism, (221) 270
Energy profile, (216) 11
Energy transduction, (208) 138
Energy transfer, (209) 37; (213) 119; (223) 161; (224) 9
Enhancer, (221) 332; (223) 299
Enkephalin, (213) 265; (216) 118
Enkephalin analog, (208) 278; (222) 71
Enkephalinase, (216) 118
Enolase, (214) 249
Enteric bacteria, (216) 133
Enterohepatic circulation, (212) 138
Enterotoxin, (213) 249; (215) 165
Enthalpy titration, (209) 77
Enzyme activation, (208) 113; (213) 333; (219) 40; (221) 241
Enzyme activity, (209) 249; (225) 93
Enzyme assay, (219) 70
Enzyme characterization, (213) 215
Enzyme conformation, (214) 8
Enzyme crystallization, (224) 89
Enzyme domain, (220) 167
Enzyme inactivation, (225) 218
Enzyme induction, (223) 223
Enzyme inhibition, (209) 49; (214) 291; (219) 296; (224) 89
Enzyme inhibitor, (208) 113; (208) 273; (211) 144; (211) 207; (216) 229; (220) 247; (220) 271; (220) 299; (223) 267
Enzyme-inhibitor complex, (209) 9
Enzyme interaction, (214) 244
Enzyme intermediate complex, (225) 87
Enzyme kinetics, (208) 109; (212) 259; (214) 357; (216) 4
Enzyme mechanism, (219) 1; (220) 15
Enzyme metabolism, (215) 252
Enzyme precursor, (213) 329; (220) 23
Enzyme processing, (223) 227
Enzyme regulation, (209) 63; (214) 249
Enzyme specificity, (217) 180
Enzyme stability, (223) 33
Enzyme-substrate interaction, (212) 53
Enzyme synthesis, (223) 227; (225) 178
Enzymic transmethylation, (209) 362
Eosinophil, (217) 242
Epidermal growth factor, (209) 231; (213) 164; (221) 105; (222) 261; (223) 223; (225) 43
Epidermal growth factor receptor, (212) 141; (214) 219; (221) 11
5-Epimerase, (223) 207
Epinephrine, (219) 283
Epithelial transport, (221) 315
Epitope, (218) 231
EPR, (213) 453; (214) 274; (216) 140; (217) 16; (217) 57; (218) 227; (218) 283; (219) 244; (221) 245; (222) 75; (222) 171; (223) 371; (226) 129
EPR signal II_s, (217) 134
Equilibrium constant, (212) 63
Erythrocyte, (209) 254; (217) 21; (222) 167; (226) 105
Erythrocyte membrane, (219) 212
Erythropoietin binding, (211) 229
Erythrosin, (213) 401
ESEEM, (214) 117
ESR, (219) 212; (222) 246; (224) 9; (224) 283
ESR signal, (214) 279
ESR spin trapping, (221) 101
Esterase, (210) 147
Estrogen, (226) 213
Ethanethiol, (215) 21
Ethanol, (208) 292; (214) 127; (215) 295; (216) 113; (225) 201
Ethanol effect, (219) 296
Ether extraction, (220) 74
Ether lipid, (208) 51; (211) 225; (220) 220
S-Ethylcysteine, (215) 21
Ethylene glycol, (209) 33
N-Ethylmaleimide, (224) 391
Eukaryotic promoter, (214) 259
Evolution, (211) 105; (216) 17; (217) 187; (219) 17; (219) 419; (221) 391; (224) 261; (224) 297
EXAFS, (222) 275
Excitation energy transfer, (225) 59
Exercise, (217) 232
Exercise training, (208) 297
Exocytosis, (215) 339; (216) 89; (219) 326; (222) 317; (222) 358; (222) 359; (223) 25
Exon, (226) 247
Exon shuffling, (214) 1
Exon/intron structure, (217) 275
3′-Exonucleolytic cleavage, (212) 317
Exopolysaccharide, (215) 79
Expandase, (214) 357
Expert system, (214) 219
Exposed domain, (217) 269
Expression, (216) 73; (223) 174
Expression cloning, (209) 219
Expression vector, (219) 360; (222) 175
pKK233-2 Expression vector, (224) 125
Extended dimeric conformation, (213) 265
Extracellular enzyme, (220) 27
Extracellular matrix, (213) 319; (218) 167
Extracellular matrix assembly, (217) 124

F_1, (213) 381
[psi] Factor, (225) 205
FAD, (224) 121
Fast kinetics, (225) 1
Fatty acid, (208) 128; (215) 175; (219) 113; (223) 155; (226) 265
Fatty acid synthase, (220) 323; (224) 133
Fatty acid synthesis, (214) 158; (222) 159; (223) 217
Fatty acyl-CoA ester, (223) 217
Fcγ receptor, (224) 29
Fe^{2+}, (208) 331
Fe-protein, (208) 73
Fe-S center, (208) 343

Femtosecond spectroscopy, (209) 37
Fenton reaction, (213) 9
Ferredoxin, (209) 311; (215) 37; (221) 343
Ferredoxin-$NADP^+$ reductase, (215) 37
Ferricytochrome c, (221) 115
Ferriprotoporphyrin IX, (223) 251
Ferritin, (215) 41
Ferrocytochrome c oxidation, (210) 195
Fetuin, (214) 45
Fetus, (222) 256
F_1F_o, (219) 274
β-Fibrinogen, (221) 18
Fibrinogen binding, (208) 435
Fibrinogen clot, (211) 10
Fibroblast, (212) 313; (214) 81; (221) 381
Fibroblast growth factor, (213) 189; (217) 1; (224) 128
Fibroblast receptor, (218) 243
Fibronectin, (208) 418; (213) 261; (215) 269; (217) 124
Fibronectin-binding protein, (221) 381
Field-inversion gel electrophoresis, (226) 88
Filament growth, (211) 35
Filamentous growth, (214) 127
Filamin, (221) 68
Filipin, (221) 375
Flash spectroscopy, (208) 337
Flavin, (224) 406
Flavin-binding site, (218) 135
Flavodoxin, (215) 63; (224) 406
Flavoenzyme, (224) 121
Flavoprotein, (226) 129
Flexibility, (211) 185; (218) 135
Flow-force relationship, (209) 325
Flow linear dichroism, (212) 263
Fluorescence, (209) 1; (213) 119; (214) 35; (215) 83; (216) 35; (218) 247; (219) 103; (222) 154; (223) 327; (224) 9; (224) 337
Fluorescence anisotropy, (213) 443; (218) 135
Fluorescence dequenching, (221) 61
Fluorescence induction, (217) 53
Fluorescence lifetime, (224) 406
Fluorescence polarization, (223) 161; (223) 381
Fluorescence probe, (219) 233
Fluorescence quenching, (217) 169; (223) 20
Fluorescence titration, (208) 189
Fluorescent probe, (222) 47
Fluoride, (216) 107
Fluorine metabolism, (220) 353
Fluoroaluminate, (218) 87
Fluorography, (209) 71
FMN, (224) 406
Fodrin, (213) 102
Folate, (217) 106
Folding intermediate, (221) 115
Formaldehyde, (209) 362
Formaldehyde production, (216) 4
Formyl peptide, (224) 219
Forskolin, (216) 107; (217) 163; (220) 57; (220) 103; (220) 342; (221) 23; (224) 33
Forssman antigen, (219) 45
Frameshift, (222) 95
FRAP, (222) 47
Free radical, (213) 9; (216) 253; (219) 212; (220) 278; (221) 101; (224) 177
Freeze-fracture, (208) 308
Freeze-fracture electron microscopy, (209) 257; (223) 148
Fructose 2,6-bisphosphate, (209) 254; (221) 124; (222) 27; (222) 167; (225) 37; (226) 43; (226) 121
Fructose-1,6-bisphosphatase, (216) 265
Fructose-2,6-bisphosphatase, (213) 375; (224) 317
FTIR, (213) 39
Fucose, (222) 271
Fucose binding, (226) 38
Functional homology, (220) 263
Functional size, (217) 53
Fungal dimorphism, (214) 127
Fura-2, (208) 47; (209) 1; (210) 132; (216) 35; (220) 89; (221) 28; (221) 284; (221) 289; (222) 144; (223) 53; (224) 396; (225) 43
Fusion protein, (215) 269

G-binding protein, (225) 127
GABA receptor, (217) 129; (222) 125
Galactogen, (221) 139
Galactokinase, (210) 147
Galactolipid synthesis, (219) 393
Galactose operon, (219) 189
β-Galactoside binding, (214) 301
β-Galactoside-binding lectin, (223) 330
Galactosyltransferase, (208) 401
β1→3-Galactosyltransferase, (221) 139
Ganglioside, (217) 94; (221) 315; (223) 42
Ganglioside G_{M_1}, (216) 99
Gap junction, (210) 169
Gas chromatography, (226) 23
Gas phase protein microsequencing, (219) 459
Gastrin, (222) 149
Gastrointestinal disease, (208) 194
Gaucher's disease, (216) 190
GAWK, (224) 142
GC-MS, (213) 209
Gel electrophoresis, (226) 250
Gel filtration, (218) 107
Gene amplification, (212) 10
Gene cloning, (208) 183; (213) 245; (226) 241
Gene deletion, (224) 49
Gene expression, (209) 187; (209) 223; (212) 233; (215) 122; (217) 260; (221) 129; (223) 299; (225) 113; (225) 238; (226) 171
Gene expression regulation, (222) 337
Gene family, (209) 97; (217) 275; (221) 129
Gene fusion, (208) 183; (209) 83; (210) 147
Gene homology, (215) 257
Gene homostabilizing propensity, (226) 247
Gene methylation, (215) 68
Gene product, (209) 187
Gene rearrangement, (221) 189
Gene regulation, (209) 92; (226) 297
Gene transfer, (213) 245
Gene variant, (212) 233
amp Gene expression, (213) 127
CH Gene, (213) 231
c-Ha-ras Gene, (211) 23
c-myc Gene, (211) 27; (212) 159
c-ras^{Ha} Gene, (211) 27
frxA Gene, (218) 52
H-2 Gene transcription, (226) 297

Ha-rasl Gene, (218) 41
IgCH Gene, (208) 221
ND-1 Gene product, (219) 108
PRla Gene, (225) 243
pX Gene, (208) 231; (209) 187
ras Gene, (217) 253
rps12 Gene, (219) 153
SUC2 Gene, (208) 31
tat Gene, (222) 286
X Gene, (209) 289
X Gene product, (209) 289
x-lor Gene, (208) 231
Genetic variant, (208) 364
Genome sequence, (216) 217
Genomic blotting, (221) 129
Genomic cloning, (225) 243
Genomic sequence, (211) 200; (213) 221
Ghost fiber, (223) 409
Glial fibrillary acidic protein, (213) 133; (213) 138
Globin chain variant, (223) 59
Globin gene, (221) 3
Globoside, (212) 91
11 S Globulin, (224) 125
Glucagon, (208) 128; (208) 455; (218) 1; (219) 145; (219) 227; (225) 37; (226) 43
Glucagon-like peptide, (211) 169; (219) 227
Glucagon receptor, (222) 256
Glucagon switching signal, (217) 212
cyclic β-1,2-Glucan, (216) 261
β-Glucanase, (220) 27
Glucocorticoid, (220) 159; (223) 223; (226) 213
Glucocorticoid regulatory site, (211) 200
Glucokinase, (221) 375
Gluconeogenesis, (213) 209; (219) 227; (221) 277; (221) 375; (224) 201
Glucose 1,6-bisphosphate, (209) 254; (222) 167
Glucose consumption, (208) 34
Glucose paradox, (221) 277
Glucose 6-phosphate, (221) 375
Glucose-6-phosphate dehydrogenase, (211) 243
Glucose phosphorylation, (216) 128
Glucose synthesis, (220) 257; (220) 259
Glucose transport, (216) 128; (219) 5; (219) 311; (224) 224
Glucose utilization, (218) 7
D-Glucose, (213) 149; (220) 103
2-[^{3}H]Glucose, (216) 128
Glucoside, (213) 411
Glucosylceramidase, (216) 190
Glutamate, (223) 143
Glutamate dehydrogenase, (223) 37; (224) 182
Glutamate isotopomer, (212) 58
Glutamate transport, (221) 370
Glutamine, (215) 187; (225) 93
Glutamine release, (221) 370
ε-(γ-Glutamyl)lysine, (224) 104
γ-Glutamyltranspeptidase, (208) 26; (211) 133
Glutathione, (214) 62
Glutathione disulfide, (213) 34
Glutathione peroxidase, (218) 251
Glutathione reductase, (218) 135
Glutelin, (222) 135
Glutelin gene, (221) 43
Glycan hydrolase, (221) 145
Glyceraldehyde-phosphate dehydrogenase, (221) 387
Glyceraldehyde-3-phosphate dehydrogenase, (219) 233; (221) 231
Glycerophosphate dehydrogenase, (218) 47
Glycerophosphorylcholine, (214) 149
Glycine transport, (212) 87
Glycinin A_2 subfamily, (224) 125
Glycogen, (208) 34
Glycogen synthase kinase-4, (209) 175
Glycogen synthesis, (221) 277
Glycogenolysis, (221) 375
Glycogenosis, (223) 113
Glycolipid, (208) 17
Glycolipid acetylglucosaminidase, (218) 131
Glycolysis, (208) 34; (209) 254; (214) 351; (217) 163; (221) 124; (221) 375; (222) 167
Glycolytic enzyme, (225) 123
Glycophospholipid, (209) 238
Glycoprotein, (208) 369; (209) 265; (215) 209; (215) 311; (219) 224; (226) 47
Glycoprotein GPIIb/IIIa, (225) 228
Glycoprotein metabolism, (222) 271
Glycoprotein synthesis, (222) 42
140 kDa Glycoprotein, (213) 319
α_2 HS Glycoprotein, (214) 45
Glycosaminoglycan, (209) 299
Glycosomal membrane, (210) 137
Glycosphingolipid, (214) 41
Glycosylation, (217) 174; (220) 27; (225) 163
Glycosyltransferase, (208) 17
Glyoxysomal lipase, (220) 23
Glyoxysome biogenesis, (220) 23
cGMP, (208) 263; (219) 369
cGMP-activated conductance, (222) 37
cGMP binding protein, (219) 293
cGMP-binding site, (222) 37
cGMP phosphodiesterase, (208) 241; (233) 169
Gold sodium thiomalate, (208) 23
Golgi fraction, (222) 256
Golgi vesicle, (222) 175
Gonadotropin, (208) 287
Gonadotropin-releasing hormone, (225) 247
ppGpp, (223) 1
Gram negative bacteria, (226) 241
Gramicidin A analog, (216) 11
Gramicidin P, (216) 11
Gramicidin S, (220) 332; (222) 251
Granulocyte, (222) 289
Graph-theoretical method, (217) 212
Graves' disease, (211) 69
Green nonsulfur bacteria, (213) 301
Green photosynthetic bacterium, (210) 189
Growth, (226) 303
Growth factor, (210) 123; (213) 406; (220) 52
Growth hormone, (213) 155; (225) 238
Growth-plate cartilage, (220) 181
Growth substance, (208) 39
GTP, (214) 176; (217) 85; (218) 153; (219) 326; (223) 47; (226) 307
GTP-binding protein, (209) 352; (213) 353; (216) 295; (218) 63; (218) 87; (220) 15; (224) 38; (226) 91

GTP-binding site, (215) 257
GTP hydrolysis, (209) 243
GTPase, (216) 94; (224) 19
Guanidine hydrochloride, (215) 155
Guanine nucleotide, (208) 99; (212) 281; (216) 89; (217) 253; (218) 87; (223) 87; (226) 155
Guanine nucleotide analog, (215) 109
Guanine nucleotide-binding protein, (211) 137; (219) 289
Guanine nucleotide exchange protein, (208) 77
Guanine nucleotide regulatory protein, (219) 259; (221) 81
Guanosine triphosphate, (219) 97
Guanylate cyclase, (219) 375
Gum arabic, (216) 67

H^+ transport, (211) 89
HaeIII isoschizomer, (214) 305
β-β-Hairpin, (224) 372
Halometabolite biosynthesis, (216) 1
Halorhodopsin, (213) 238
HDL, (220) 371
HDL receptor, (222) 159
Heat shock, (215) 295; (220) 113; (220) 177; (221) 110; (225) 259
Heat shock response, (223) 127
Heavy atom effect, (211) 155
Heavy meromyosin, (212) 254; (223) 409
Helical structure, (221) 320
Helix dynamics, (226) 1
α-Helix, (213) 297
Heme, (213) 453; (222) 115
Heme coordination, (221) 349
Heme protein, (208) 305; (208) 325; (214) 111; (219) 244
Heme-regulated protein kinase, (208) 117
Heme regulation, (209) 162
Hemocyanin, (221) 150
Hemoglobin, (221) 161
Hemoglobin α-subunit, (213) 289
Hemoglobulin, (209) 129
Hemolysis, (209) 18; (221) 355; (223) 77
Hemoprotein, (212) 213
Heparin, (213) 159; (223) 207; (225) 109; (226) 109
Heparin cofactor II, (225) 109
Hepatic nerve, (219) 145
Hepatitis B virus, (221) 11
Hepatocyte, (208) 455; (217) 247; (219) 227; (219) 455; (223) 223; (224) 104
Hepatocyte growth factor, (224) 311
Hepatocyte heterogeneity, (221) 277; (224) 182
Hepatocyte-stimulating factor, (221) 18
Hepatotoxicity, (209) 139
Herbicide-binding protein, (208) 313; (208) 347; (211) 221
Herbicide resistance, (211) 221
Herpesvirus, (214) 153
Heterobifunctional reagent, (221) 408
Heterodimer, (224) 311
Heteroduplex, (215) 327
Heterogeneity, (209) 311; (216) 270
Heterologous system, (213) 385
Hexagonal H_{II} phase, (223) 148
Hexaheme, (218) 227
Hexamer motif, (223) 273
High-molecular-mass complex, (220) 363
Hinge, (208) 221
Hinge protein, (226) 171
Hirudin, (211) 10
Histamine, (209) 362
Histamine release, (221) 236
Histamine-N-methyltransferase, (209) 362
Histidine, (208) 73
Histidine residue, (219) 301
Histidine-rich glycoprotein, (216) 17
Histone, (220) 193; (222) 322
Histone deacetylase, (220) 79
Histone H1°, (210) 161
Histone H1° subtype, (210) 161
Histone H3, (211) 59
Histone isolation, (226) 250
Histone subtype proportion, (210) 161
Histrionicotoxin, (212) 292
HIV, (211) 17; (213) 133; (213) 138; (218) 81; (218) 231; (222) 286
HIV receptor, (213) 133; (213) 138
HNK-1 epitope, (225) 33
Hoechst 33258, (215) 95
Holocephalan, (208) 445
Homology, (214) 219; (216) 145
2D Homonuclear Hartman-Hahn spectroscopy, (212) 91
1D Homonuclear Hartmann-Hahn spectroscopy, (219) 45
Hormone action, (226) 213
Hormone precursor, (212) 307; (223) 63
Hormone processing, (218) 31; (221) 423
Hormone release, (225) 247
Hormone sensitivity, (209) 175
Hormone synthesis, (210) 185
Hornetin, (209) 18
Horseradish peroxidase, (222) 246
HPLC, (208) 147; (208) 364; (208) 445; (214) 50; (214) 351; (215) 179; (219) 426; (220) 363; (222) 42; (223) 227
HTLV-I, (208) 231; (209) 187
HTLV-II, (208) 231; (209) 289
Human foamy virus, (222) 286
Human T-cell leukemia, (209) 289
Hyaluronidase, (226) 220
Hybrid gene, (226) 364
Hybrid protein, (208) 194; (221) 226; (224) 213
Hybridization, (208) 133; (208) 158; (211) 31; (222) 191
Hybridization probe, (220) 262
Hydration, (224) 172
Hydrazone, (221) 299
Hydrogen bonding, (213) 423; (219) 17; (224) 161
Hydrogen evolution, (219) 93
Hydrogen exchange, (216) 249; (225) 11
Hydrogen fluoride, (220) 383
Hydrogen peroxide, (210) 195; (214) 269; (221) 427; (225) 139
Hydrogenase, (221) 241
Hydropathy plot, (212) 1
Hydroperoxide, (221) 349
12-Hydroperoxyicosatetraenoic acid, (213) 169
Hydrophilic photolabeling, (223) 366
Hydrophobic chromatography, (209) 311
Hydrophobic moment plot, (215) 261
Hydrophobic probe, (224) 9
Hydrophobicity projection, (215) 261
Hydroxamate, (211) 239
Hydroxide-ion translocation, (222) 235
Hydroxidemotive redox loop, (222) 235

4-Hydroxyalk-2-enal, (224) 357
Hydroxyapatite, (208) 401
β-Hydroxyasparagine, (222) 129
26-Hydroxycholesterol, (218) 77
6-Hydroxy-D-nicotine oxidase, (224) 121
2′-Hydroxyisoflavone, (214) 101
Hydroxyl free radical, (222) 246
Hydroxyl radical, (213) 9; (213) 15; (222) 75; (225) 139
Hydroxyl radical footprinting, (225) 195
Hydroxyl radical scavenger, (213) 23
Hydroxylase, (214) 357
21-Hydroxylase, (217) 65
Hydroxylation, (222) 186
N-Hydroxylation, (211) 239
9-Hydroxylinoleic acid, (209) 249
Hydroxymethylglutaryl-CoA reductase, (223) 217
β-Hydroxynorvaline, (223) 122
Hydroxyproline cluster, (221) 293
Hydroxyproline-rich glycoprotein, (209) 57
Hydroxyurea, (219) 197; (222) 171
Hyperfine interaction, (224) 75
Hyperfine shift, (222) 115; (226) 179
Hyperoxaluria, (222) 17
Hyperpolarization, (218) 247; (223) 82
Hypersensitivity, (225) 133
Hypertrehalosemic peptide, (219) 83
Hypochlorous acid, (213) 15
Hypophysis, (220) 108
Hypothalamus, (216) 123
Hypothyroidism, (223) 279
Hypoxia, (224) 4

^{125}I labeling, (215) 305; (223) 371
Icosanoid, (211) 73; (221) 155
IDL, (216) 275
IL-2 receptor, (218) 81
2-Iminothiolane, (214) 13
Immobilization, (214) 13
Immune complex, (224) 29
Immune complex disease, (213) 415
Immune electron microscopy, (226) 255
Immune system, (216) 151
Immunity, (222) 199
Immunoaffinity chromatography, (218) 159; (224) 306
Immunoassay, (212) 237
Immunoblot analysis, (224) 43
Immunoblot assay, (224) 125
Immunoblotting, (213) 448; (220) 108; (222) 17; (226) 145
Immunocytochemistry, (220) 108
Immunoelectron microscopy, (216) 207
Immunoenzyme technique, (208) 194
Immunoglobulin, (208) 221; (211) 123; (213) 231; (222) 337
Immunoglobulin deficiency, (213) 231
Immunoglobulin E, (208) 369
Immunoglobulin G3 deficiency, (208) 221
Immunoglobulin gene, (214) 87
Immunoglobulin-linked enzyme, (214) 192
Immunogold labeling, (213) 29
Immunohistochemistry, (217) 62
Immunological castration, (214) 65
Immunomodulating tripeptide, (218) 55
Immunoperoxidase electrophoresis, (212) 220
Immunoprecipitation, (212) 145; (217) 37
Immunostimulation, (218) 55
Immunotoxin, (216) 73
Incretin, (211) 169
Indirect end labeling, (208) 31
Indo-1, (216) 35; (221) 95
Indomethacin, (209) 249; (225) 103
Inducer, (208) 258
Induction, (217) 292; (221) 167
Infectious transcript, (219) 65
Infrared spectroscopy, (223) 289
Inhibitor, (210) 211; (219) 380
Inhibitor-1, (216) 291
Inhibitor protein, (213) 67
α_1-Inhibitor$_3$, (221) 55
Initiation factor, (215) 115
Initiation factor eIF-2, (208) 117; (209) 162
Initiation of DNA replication, (223) 237
Inositol lipid, (220) 52
Inositol phosphate, (208) 287; (210) 181; (210) 204; (218) 113; (220) 155; (220) 327; (222) 349
Inositol phospholipid, (208) 287; (211) 73; (218) 68; (219) 283
Inositol tetrakisphosphate, (208) 283; (214) 365; (217) 117; (219) 125
Inositol triphosphate, (219) 445
Inositol trisphosphate, (208) 283; (214) 365; (217) 85; (217) 117; (218) 87; (219) 125; (220) 302
Inositol 1,4,5-trisphosphate, (213) 95; (217) 247; (218) 153; (221) 289
Inositol trisphosphate accumulation, (223) 413
myo-Inositol 1,3,4,5,6-pentakisphosphate, (213) 278
myo-Inositol 1,4,5,6-tetrakisphosphate, (213) 278
myo-Inositol uptake, (220) 84
Insect immunity, (208) 113
Insect reproduction, (222) 345
Insertion element, (219) 254
Insertion sequence IS30, (208) 7
Insoluble fraction, (221) 68
Insulin, (208) 445; (209) 238; (212) 281; (213) 419; (214) 122; (214) 327; (215) 179; (215) 187; (216) 94; (217) 253; (220) 84; (220) 159; (220) 311; (223) 227; (224) 137
Insulin action, (224) 224
Insulin gene, (222) 332
Insulin gene expression, (223) 131
Insulin-like growth factor, (208) 439; (215) 122
Insulin-like growth factor II, (214) 259; (222) 181
Insulin-like growth factor-binding protein, (208) 439
Insulin mediator, (209) 238
Insulin receptor, (212) 141; (214) 107; (214) 219; (221) 11; (224) 198
Insulin release, (220) 103
Insulin response, (214) 327
Insulin secretion, (208) 63; (215) 9; (219) 139; (219) 311; (223) 131
Insulin synthesis, (213) 149
Integrated structural prediction, (212) 193
Intercalation, (209) 269; (225) 151
Interferon, (211) 78; (214) 153; (217) 292; (226) 53; (226) 297
Interferon-β2, (221) 18
Interferon-γ, (212) 233
Interferon function, (226) 13
Interferon induction, (226) 13

Interferon production, (208) 55; (211) 78
γ-Interferon, (221) 91
Interleukin-1, (209) 295; (212) 35; (215) 160; (223) 294; (225) 21; (226) 314
Interleukin-1 receptor, (209) 295; (215) 160; (219) 365; (225) 21
Interleukin-2, (226) 47
Interleukin-3, (208) 231
Intermediate filament, (221) 403
Intermediate transfer efficiency, (214) 244
Internal promoter, (219) 269
Internalization, (221) 105
Internalization blockade, (214) 323
Interphotoreceptor retinoid-binding protein, (208) 133
Interproton distance, (213) 269; (219) 11
Intracellular enzyme, (222) 11
Intron, (226) 247
Intron splicing, (222) 181
Inversion, (209) 57
Iodide uptake, (221) 91
Iodo-labeling, (208) 347
Iodogen, (224) 198
Ion channel, (209) 165; (212) 292; (220) 295; (221) 172; (224) 172; (226) 83
Ion channel inhibition, (225) 173
Ion channel kinetics, (222) 63
Ion flux, (217) 129
Ion pair, (224) 161
Ion-pair chromatography, (224) 128
Ionic strength effect, (220) 376
Ionomycin, (215) 183; (226) 307
Ionophore A23187, (211) 73; (212) 213; (212) 242; (225) 43
Iota toxin, (225) 48
IRCM-serine protease 1, (211) 144
Iron, (208) 292; (213) 9; (215) 151; (226) 1
Iron-binding protein, (214) 362
Iron coordination, (223) 92
Iron (II)·EDTA complex, (225) 195
Iron incorporation, (215) 41
Iron metabolism, (209) 147
Iron porphyrin, (208) 305
Iron reactivity, (208) 325
Iron-sulfur center, (216) 140; (218) 52
Iron-sulfur cluster, (214) 117
Iron-sulfur protein, (209) 311; (219) 161; (226) 129
Iron-superoxide dismutase, (221) 87; (223) 92
Iron transfer, (215) 41
Irreversible thermodynamics, (224) 201
Ischemia, (213) 23; (216) 57; (221) 101
Islet-activating protein, (213) 353; (222) 317
Isoelectric focusing, (208) 364; (224) 343
Isoenzyme, (208) 297
Isoflavone hydroxylase, (214) 101
Isoflavone oxidoreductase, (213) 324
Isoflavonoid phytoalexin, (214) 101
Isolectin, (220) 116
Isoprenaline, (222) 27
Isoproterenol, (217) 163; (218) 73
Isotocin, (220) 98
Isotope effect, (216) 4
Isotopic substitution effect, (214) 28
Isotype heterogeneity, (219) 31
Isozyme, (222) 99; (226) 176

Juvenile hormone, (213) 85
Juvenile hormone synthesis, (222) 345

K^+ channel, (208) 58; (208) 63; (209) 117; (209) 165; (215) 9; (216) 211; (219) 351; (220) 187; (223) 82; (226) 280
K^+ conductance, (217) 247
K^+ current, (208) 283; (217) 11
K^+-diffusion potential, (224) 348
K^+ stimulation, (223) 413
Kallikrein-like, (214) 317
Kanamycin, (226) 364
Karyophilic signal, (226) 217
Karyoplast, (210) 165
Ketanserin, (219) 283
K^+/H^+ antiporter, (220) 288
Kinase, (212) 281; (217) 253
Kinase II, (213) 184
Kinesin, (222) 204
Kinetic control, (221) 265
Kinetic intermediate, (224) 9
Kinetic property, (208) 297
Kinetics, (213) 128; (219) 27; (225) 247; (226) 331
T-Kinin, (218) 266; (219) 437
Kininogen, (216) 17
T-Kininogen, (218) 266
T-Kininogenase, (218) 266
Krebs cycle, (213) 209
Kringle, (224) 97
Kringle structure, (209) 111

Laccase, (215) 219; (219) 335
α-Lactalbumin, (223) 327; (223) 405
β-Lactam, (221) 167; (224) 401; (225) 218
β-Lactamase, (218) 126; (221) 179
Lactate dehydrogenase, (208) 297; (212) 203
Lactate oxidation, (219) 227
Lactation, (217) 37
β-Lactoglobulin, (221) 115
Lactose permease, (209) 325
Laminin, (217) 94
Late mitogenic event, (220) 1
Latency, (210) 137
Latent human leukocyte collagenase, (208) 23
Lateral diffusion, (222) 47
Latex particle, (213) 174
Latrunculin A, (213) 316
LC2-ATPase site communication, (220) 376
LDL, (216) 275; (218) 77; (220) 159; (220) 371
LDL modification, (218) 243
LDL receptor, (218) 77; (222) 159
Leader peptide, (213) 238; (215) 233
Leader sequence, (222) 95
Lectin, (214) 301; (215) 140; (216) 67; (216) 145; (220) 116
Lectin affinity chromatography, (209) 71
Lectin purification, (223) 330
Left-shift, (209) 129
Leghemoglobin, (213) 289
Legumin, (222) 175
Lens crystallin, (209) 107
Lens membrane, (212) 49
Lethal protein, (209) 18
Leucine, (213) 45

Leukaemia, (224) 291
Leukemogenesis, (2088) 231
Leukocyte, (209) 299; (217) 242; (217) 265
Leukocyte antigen, (210) 127
Leukotriene, (213) 169
Leukotriene A_4 synthase, (217) 265
Leukotriene B_4, (214) 167
Leukotriene C_4, (208) 26
Leukotriene E_4, (208) 26
Leukotriene synthesis, (216) 123
11,12-Leukotriene A_4, (213) 169
Li^+, (218) 47
Ligand-gated K^+ channel, (216) 104
Light activation, (221) 215
Light-arvestin complex II, (215) 25
Light-chain-specific modification, (220) 376
Light-driven chloride pump, (217) 297
Light-harvesting complex, (213) 29; (213) 245; (224) 343; (225) 59; (226) 275
Light-harvesting complex II, (221) 25; (221) 205
Light-harvesting polypeptide, (210) 189
Light regulated gene expression, (225) 159
Light regulation, (209) 87
Light scattering, (211) 35; (212) 263; (221) 253
Light/dark regulation, (213) 18
Lignin degradation, (210) 199; (220) 149
Lignin peroxidase, (210) 199; (218) 255; (220) 149
β-O-4 Lignin substructure, (218) 255; (210) 61
Ligninase, (221) 327
Limited proteolysis, (223) 402
Linear dichroism, (226) 275
Linear force-flow relationship, (224) 201
Lipase, (209) 175
Lipid, (225) 97
Lipid A, (218) 238
Lipid bilayer, (226) 115
Lipid binding, (224) 261
Lipid cofactor, (214) 339
Lipid-drug conjugate, (225) 97
Lipid flip/flop, (219) 311
Lipid interaction, (215) 73
Lipid metabolism, (208) 51
Lipid peroxidation, (208) 292; (209) 191; (209) 305; (213) 9; (214) 362; (215) 151; (220) 197; (223) 251; (224) 357
Lipid-protein interaction, (214) 181
Lipid pump, (219) 311
Lipid synthesis, (214) 158
Lipid transfer, (220) 371
Lipidosis, (219) 130
Lipoamide dehydrogenase, (218) 135
Lipocortin, (216) 45; (219) 169; (219) 176; (221) 397; (223) 267
Lipoic acid, (222) 211
Lipolysis, (208) 128; (208) 269
Lipophilic probe, (218) 22
Lipopolysaccharide, (218) 238
Lipoprotein, (218) 11; (219) 239; (224) 261; (226) 371
Lipoprotein lipase, (225) 178
Lipoprotein metabolism, (213) 221
Liposome, (216) 99; (216) 159; (219) 233; (223) 42
Liposome-induced peptide conformation, (220) 332
Lipoxin, (208) 248; (225) 103
Lipoxygenase, (208) 248
Lipoxygenase product, (216) 123; (217) 242
5-Lipoxygenase, (217) 265
12-Lipoxygenase, (213) 169
Liver pathology, (222) 17
Liver perfusion, (221) 277
Liver regeneration, (208) 418; (209) 191; (209) 197
Local anesthetic, (226) 77
Long-chain fatty alcohol, (213) 406
Long terminal repeat, (221) 332; (223) 299
Long-wavelength chlorophyll a form, (223) 165
Low-potential cytochrome, (209) 261
Low-temperature embedding, (216) 207
Low-temperature spectrophotometry, (225) 255
Low-temperature spectroscopy, (223) 165
Lubrol-WX disruption, (215) 300
Luminescence, (209) 28
Luteinizing hormone, (223) 321; (225) 247
Luteinizing hormone-releasing hormone, (214) 65; (214) 139
Lymphocyte, (216) 62; (220) 57; (222) 349; (224) 33; (224) 291
Lymphocyte mitogenesis, (223) 6
Lymphocyte transformation, (213) 199
Lymphokine, (220) 52
Lymphoma, (224) 291
Lysine, (222) 109
Lysine site, (209) 111
Lysine-Sepharose CL4B, (209) 13
Lyso-PAF, (211) 225
Lyso PAF-acether, (226) 371
Lyso-PAF analog, (211) 225
Lysosomal enzyme, (219) 130
Lysosome, (208) 352; (208) 427; (210) 165; (223) 69
Lysozyme, (223) 405

M_1 RNA, (220) 283
M_2 subtype, (209) 367
M-current, (220) 302
M-intermediate, (222) 275
α_1-Macroglobulin, (219) 37
α_2-Macroglobulin, (209) 9; (217) 101; (219) 37; (221) 55; (222) 83; (225) 223
Macrophage, (215) 223; (217) 6; (218) 200; (221) 427; (222) 341; (223) 284
Macrophage activation, (220) 93
Major groove binding, (225) 151
Malaria, (209) 23
Malate dehydrogenase, (213) 329
Malate dehydrogenase ($NADP^+$), (209) 87
Malate inhibition, (213) 18
Malignant transformation, (210) 123
Malondialdehyde, (226) 101
Maltoporin, (220) 136
Maltose-binding protein, (220) 136
Manganese, (215) 79; (221) 245
Manganese multiline, (217) 134
Mannose, (215) 140
Marker enzyme, (216) 51
Mass spectrometry, (226) 23
Mast cell, (216) 89; (221) 236; (222) 317; (226) 307
MCD, (219) 244; (226) 129
Mechanochemical coupling, (222) 204
Medicarpin, (213) 324
Megakaryocytic differentiation, (213) 89

Meiose, (226) 324
Melanocyte, (208) 17
Melanocyte-stimulating hormone receptor, (226) 134
α-Melanocyte-stimulating hormone, (226) 337
Melanoma, (208) 17
Melittin, (218) 173; (223) 148
Membrane, (222) 154
Membrane binding, (211) 127; (225) 233
Membrane-bound enzyme, (217) 269
Membrane-bound protein, (213) 438
Membrane capacity, (217) 25
Membrane conductance, (224) 208
Membrane current, (220) 302
Membrane disordering, (209) 33
Membrane energization, (218) 22
Membrane enzyme, (219) 233
Membrane fluidity, (223) 381
Membrane fusion, (217) 25; (217) 158; (222) 327; (223) 25; (224) 172
Membrane glycoprotein, (216) 151
Membrane insertion, (211) 133
Membrane permeability, (208) 43
Membrane permeabilization, (213) 401
Membrane potential, (209) 152; (213) 128; (213) 337; (214) 343; (214) 365; (219) 477; (223) 82; (223) 289; (224) 79; (224) 337; (225) 1
Membrane potential-dependent gate, (226) 270
Membrane protein, (208) 211; (213) 238; (214) 219; (219) 161; (220) 23; (221) 260; (222) 154; (226) 105
Membrane protein crystal, (226) 275
Membrane protein solubilization, (213) 283
Membrane receptor transport, (215) 323
Membrane reconstitution, (211) 1
Membrane stacking, (225) 59
Membrane surface, (223) 366
Membrane thiolation, (214) 13
Membrane transport, (216) 257
Membrane vesicle, (212) 87; (213) 45
Meninges, (216) 118
Mepacrine, (216) 51
7-Mercaptoheptanoylthreonine phosphate, (213) 123; (214) 265
N-7-Mercaptoheptanoylthreonine phosphate, (220) 358
Meromyosin, (210) 177
Mersalyl, (211) 89
Mesophyll chloroplast, (219) 331; (219) 347
Messenger ribonucleoprotein, (219) 410
[D-Ala^2]-Met-enkephalin, (217) 6
Met^5-enkephalin, (215) 215
Met-T-kinin, (219) 437
fMet-Leu-Phe receptor, (219) 289
Metabolic channeling, (224) 59
Metabolic regulation, (224) 201
Metabolic zonation, (221) 277
Metabolite channeling, (212) 45
Metachromatic leukodystrophy, (219) 130
Metacyclogenesis, (208) 379
Metal-substituted bacteriorhodopsin, (222) 275
Metalloprotein, (223) 92
Metallothionein, (217) 292; (219) 335
Metavinculin, (220) 291
Methacholine, (220) 302
Methanogenesis, (213) 123; (215) 53; (220) 358
Methanogenic bacteria, (214) 265; (220) 358
Methanol, (215) 295
Methanol oxidation, (216) 4
Methemoglobin, (221) 355
Methicillin resistance, (212) 237; (221) 167
Methionine, (224) 177
Methionine aminopeptidase, (215) 160
Methionine sulfoxide, (224) 177
4-Methoxypyridine, (212) 53
Methyl-CoM reductase, (213) 123; (214) 265; (220) 358
Methyl sugar, (209) 57
5-Methyl cytosine, (219) 464
N^6-Methyladenosine, (213) 309
Methylalanine, (220) 8
Methylamine dehydrogenase, (221) 299
γ-N-Methylasparagine, (221) 1
N-Methyl-D-aspartate, (223) 143
Methylation, (213) 297; (221) 332
Methylation analysis, (211) 175
Methylation interference analysis, (223) 273
3-Methylcholanthrene, (209) 305
cis-(+)-3-Methylfentanylisothiocyanate, (224) 272
Methyllycaconitine, (226) 357
Methylmalonyl-CoA decarboxylase, (220) 121
Methylmethanesulfonate, (212) 173
Methylmethionine, (220) 8
N-Methyl-N-nitrosourea, (212) 173
2′-O-Methyloligonucleotide, (215) 327
Methylphenylalanine, (220) 8
Methylproline, (220) 8
Mg dependence, (211) 190
Mg^{2+}, (216) 35; (216) 291; (218) 113; (219) 279; (221) 387
Mg^{2+} binding, (209) 77
Mg^{2+} effect, (208) 43
Microbody biogenesis, (213) 329
Microbore HPLC, (224) 128
Microcalorimetry, (209) 77
Microemulsion, (223) 33
Microfilament, (221) 68
Microfilament-membrane interaction, (220) 291
Microglia, (223) 284
Microheterogeneity, (209) 107; (221) 134
Microsequence, (219) 415
Microsequencing, (224) 128; (225) 215
Microsome, (208) 292; (209) 49; (209) 243; (209) 305; (211) 41; (213) 411; (215) 151; (217) 85; (220) 197; (224) 331
Microspectrophotometry, (226) 275
Microtubule, (212) 145
Microtubule assembly, (214) 226; (217) 237
Microtubule-associated protein, (217) 237; (224) 361; (226) 28
Microtubule-associated protein 1, (212) 145
Microvillar hydrolase, (208) 34
Microvillus, (225) 269
Milk whey protein, (216) 270
Minichromosome, (214) 71
Minor groove binding, (212) 297
Mismatch error, (221) 194
Mismatch repair, (214) 87
Mitochondria, (208) 427; (209) 152; (209) 197; (209) 227; (210) 142; (212) 114; (214) 62; (214) 117; (214) 158; (214) 269; (215) 300; (216) 241; (221) 270; (223) 247; (224) 251; (224) 322; (226) 77; (226) 171; (226) 265

Mitochondrial carrier family, (212) 1
Mitochondrial cytochrome b split gene, (208) 208
Mitochondrial genome, (219) 108
Mitochondrial outer membrane, (226) 83
Mitochondrial targeting, (222) 95
Mitogen, (225) 16
Mitogenic lectin, (216) 62
Mitogenic response, (220) 1
Mnemonic enzyme, (221) 194
Mobility shift assay, (223) 273
Model system, (222) 358
Modified nucleotide, (222) 89
Modulator protein, (211) 190; (216) 291
Modulator subunit, (224) 385
Molecular assembly, (208) 147
Molecular clock, (217) 184
Molecular cloning, (225) 82
Molecular design, (214) 65
Molecular evolution, (213) 301; (214) 1; (220) 167
Molecular form, (209) 206
Molecular heterogeneity, (226) 17
Molecular mass, (211) 179
Molecular mechanism, (222) 359
Molecular mimicry, (214) 236
Molecular modelling, (209) 269
Molecular orbital calculation, (219) 279
Molecular structure, (208) 331
Molybdenum, (213) 372
Monensin, (208) 34
Monoamine oxidase, (214) 291
Monoamine transporter, (222) 215
Monobasic cleavage, (211) 169
Monoclonal antibody, (208) 236; (212) 127; (212) 259; (213) 55; (214) 181; (215) 137; (215) 269; (217) 124; (218) 271; (219) 31
Monocyte, (219) 169
Monomeric actin, (217) 89
Morphine, (217) 6
Mössbauer spectroscopy, (208) 331
Motilin, (223) 74
Mucin, (215) 137
Mucin glycopeptide, (226) 23
Multi-site catalysis, (213) 381
Multigene deletion, (213) 231
Multigene family, (209) 92; (209) 107; (221) 134
Multiheme cytochrome, (209) 261
Multiple oxygenation, (208) 248
Multiple relayed COSY, (212) 91
Muramyl peptide, (220) 93
Muscarinic acetylcholine receptor, (209) 367; (214) 285
Muscarinic receptor, (216) 104; (220) 155; (226) 287
Muscle contractility, (217) 81
Muscle contraction, (209) 1; (215) 279; (217) 232; (219) 306
Mutagenesis, (208) 143
Mutant, (209) 87
Mutant structure, (218) 271
ompA Mutant, (223) 387
Mutational alteration, (208) 208
Myasthenia gravis, (226) 235
Myelin, (218) 261; (223) 417
Myelin basic protein, (211) 190
Myeloblastosis, (214) 81
Myeloperoxidase, (213) 15; (214) 111
Myocardial infarction, (214) 192
Myocyte, (223) 53
Myoglobin, (213) 289; (222) 115
Myosin, (211) 31; (212) 154; (221) 77; (225) 269
Myosin dephosphorylation, (217) 81
Myosin heavy chain kinase, (212) 154; (223) 262
Myosin-LC2 interaction, (220) 376
Myosin light chain, (215) 279; (219) 469
Myosin light chain kinase, (212) 154; (223) 262
Myosin light chain phosphorylation, (219) 469
Myosin subfragment 1, (211) 31
Myotoxin a, (224) 89
Myristic acid, (213) 428
Myxothiazol, (223) 315; (225) 6

N^6 region, (223) 197
N-linked carbohydrate, (221) 150
N-terminal analysis, (222) 6; (222) 99
N-terminal heterogeneity, (213) 385
N-terminal processing, (215) 160
N-terminal sequence, (209) 206; (218) 277
N-terminus, (220) 8; (220) 278
Na channel, (219) 355
Na^+, (214) 187; (220) 84
Na^+ channel, (214) 163
Na^+ channel toxin, (218) 59
Na^+ dependence, (211) 195
Na^+ effect, (208) 43
Na^+ jump, (219) 331; (219) 347
Na^+ pump, (215) 335; (225) 145
Na^+ transport, (214) 335
NAD, (212) 203
NADH-cytochrome b_5 reductase, (209) 49
NADH dehydrogenase, (219) 108
NADH oxidase, (215) 335
NADH oxidoreductase, (214) 269
NADP, (211) 137
NADPH-cytochrome P-450 reductase, (225) 139
NAD(P)H-dependent reduction, (219) 212
NADPH oxidase, (214) 279
Na^+/H^+ antiporter, (226) 270
Na^+/H^+ exchange, (212) 123; (215) 223; (225) 72
Nalidixic acid, (221) 110
Nascent polypeptide, (226) 255
Native fragment, (211) 127
Natural killer cell cytotoxic factor, (226) 261
NC1 domain, (208) 203
NcoI site, (224) 125
Nebulin, (224) 49
Neo-kyotorphin, (208) 253
Neomycin, (219) 445
Neoplastic tissue, (225) 113
Nerve growth factor, (208) 47; (208) 258; (223) 117; (224) 43
Netropsin, (225) 195
Neural cell adhesion molecule, (219) 135; (220) 108; (225) 33
Neurite, (223) 143; (226) 337
Neurite outgrowth, (217) 94
Neuroblastoma, (220) 302; (226) 287
Neuroendocrine system, (210) 157
Neurofibrillary tangle, (217) 62
Neurofilament, (209) 203; (221) 403
Neurohypophyseal peptide, (220) 98

Neuromedin N, (220) 243
Neuron, (210) 157
Neuron-specific enolase, (222) 139
Neuronal acetylcholine receptor, (219) 459
Neuronal differentiation, (223) 143
Neuronal nicotinic acetylcholine receptor, (212) 292
Neuronal plasticity, (226) 337
Neuronal resistance, (224) 4
Neuropeptide, (211) 105; (216) 200
Neuropeptide distribution, (220) 243
Neuropeptide K, (208) 67
Neuropeptide Y, (225) 209
Neurophysin-copeptin precursor, (217) 180
Neurotensin, (220) 243
Neurotoxicity, (209) 18
Neurotoxin, (214) 295; (214) 347; (222) 79
Neurotoxin fragment, (226) 115
Neurotransmitter, (211) 105; (224) 391
Neurotransmitter exchange, (213) 61
Neurotransmitter receptor, (222) 191
Neurotransmitter uptake, (213) 61
Neurotrophic factor, (226) 337
Neutrophil, (211) 165; (212) 242; (213) 15; (214) 181; (214) 279; (216) 31; (219) 40; (221) 289; (221) 365; (224) 219
Nicotinamide, (223) 279
Nicotine, (213) 55; (219) 459; (226) 357
Nicotinic acetylcholine receptor, (214) 253; (214) 285; (222) 56; (222) 63
Nicotinic receptor, (226) 357
Nifurtimox, (220) 197
Nitrate reductase, (209) 147; (213) 372
Nitration, (218) 287
Nitrene, (226) 134
Nitric oxide intermediate, (218) 227
Nitrite reductase, (218) 227; (219) 244
Nitro compound, (220) 197
Nitrogen fixation, (216) 1
Nitroxide free radical, (222) 246
NMB, (209) 111
NMR, (209) 265; (209) 295; (212) 91; (213) 269; (213) 278
2D NMR, (208) 94; (216) 249; (217) 106; (219) 17; (219) 426
solid-state NMR, (213) 293; (225) 183
^{13}C-NMR, (212) 58; (213) 209; (215) 79; (216) 261; (219) 5; (222) 115
^{19}F-NMR, (213) 283; (220) 353; (214) 111; (215) 88; (215) 160; (216) 4; (219) 11; (219) 45
^{1}H-NMR, (208) 211; (211) 175; (212) 53; (213) 289; (213) 391; (214) 351; (215) 311; (219) 17; (219) 239; (219) 426; (219) 464; (221) 320; (224) 9; (226) 179
2D ^{1}H-NMR, (222) 109
^{2}H-NMR, (209) 33; (218) 173
^{15}N-NMR, (217) 106
^{31}P-NMR, (208) 89; (208) 94; (213) 293; (217) 21; (225) 183; (219) 464; (223) 148
NOESY spectrum, (219) 464
Nonbilayer lipid structure, (223) 148
Noncarbohydrate moiety, (215) 79
Noncovalent interaction, (217) 101
Nondenaturing gradient polyacrylamide gel electrophoresis, (223) 255
Nonionic detergent, (223) 255
Nonsense suppression, (225) 205
Nonshivering thermogenesis, (224) 353
Noradrenaline, (208) 128; (217) 247; (219) 139; (219) 445; (221) 155
Norepinephrine, (214) 130; (223) 53; (224) 353
Northern blotting, (211) 27; (213) 448; (215) 122
Notochord, (224) 23
Noxiustoxin, (226) 280
dNTP selection, (216) 221
Nuclear matrix, (208) 451; (212) 276
Nuclear oncogene, (215) 345
Nuclear Overhauser effect, (215) 88; (215) 215; (216) 249; (219) 11; (226) 179
Nuclear protein, (209) 289; (215) 237; (223) 273; (226) 217
Nuclease evolution, (226) 63
Nucleoplasmin, (226) 109
Nucleoprotein particle, (209) 340
Nucleosomal spacing, (225) 120
Nucleosome positioning, (208) 31
Nucleotide, (226) 67
Nucleotide binding, (208) 217
Nucleotide-binding domain, (208) 1
Nucleotide-binding protein, (208) 301; (218) 222
Nucleotide-binding site, (219) 156
Nucleotide content, (219) 321
Nucleotide effect, (208) 397
Nucleotide sequence, (208) 77; (209) 213; (209) 367; (211) 160; (212) 98; (212) 168; (213) 73; (213) 301; (216) 217; (216) 229; (217) 187; (221) 415; (222) 135; (223) 169; (223) 181; (223) 340; (225) 188; (226) 8; (226) 91; (226) 96; (226) 176
Nucleotide substrate, (225) 123
Null allele, (217) 65
Nutritional effect, (220) 231

O-cycle, (219) 387
O-specific polysaccharide, (211) 175
Octadeoxyribonucleotide, (212) 73
Octopamine, (219) 83
ODMR, (211) 83
ODMR spectroscopy, (211) 155; (220) 347
Odor, (219) 224
Odor reception, (212) 225
Odor transmission, (212) 225
Okadaic acid, (217) 81
Olfaction, (212) 225
Olfactory epithelium, (219) 224
Oligodeoxyribonucleotide template, (216) 225
Oligomerization, (221) 199
Oligomycin sensitivity, (223) 309
Oligonucleotide, (208) 94; (216) 249
Oligonucleotide-directed mutagenesis, (213) 155; (218) 119
Oligonucleotide probe, (213) 221
Oligonucleotide promoter, (212) 271
Oligonucleotide substrate, (212) 267
Oligoribonucleotide, (212) 317
Oligosaccharide, (209) 265; (226) 23
Oncogene, (208) 39; (208) 231; (208) 301; (218) 222; (222) 289; (223) 243; (226) 291
Oncogene expression, (215) 127
c-Ha-ras Oncogene, (219) 70
met Oncogene, (209) 340; (209) 357
ras Oncogene, (224) 1
V-myb Oncogene, (214) 81

Oncoprotein, (222) 293
Ontogeny, (220) 201
Oocyte, (219) 75; (226) 324
Opal termination codon, (226) 227
Open reading frame, (219) 181
Opiate addiction, (217) 6
Opiate receptor, (222) 71; (222) 327; (224) 272
Opioid receptor, (208) 278
κ-Opioid receptor agonist, (223) 335
Opsonised particle size, (216) 31
Optical absorption spectrum, (214) 274
Organ spectroscopy, (215) 228
Organelle membrane, (224) 172
Organic free radical, (214) 274
Organic solvent, (213) 333; (223) 33
Organic solvent effect, (222) 163
Ornithine carbamoyltransferase, (208) 427; (224) 182
Ornithine decarboxylase, (213) 199; (215) 68
Ornithine decarboxylase mRNA, (208) 151
Orthophosphate, (213) 333
Osteoblast, (208) 263
Ouabain binding, (225) 27
Outer membrane, (211) 1; (211) 53; (223) 387
Outer membrane protein, (221) 226
Overproduction, (224) 193
Ovostatin, (222) 83
Oxaloacetate, (220) 259
Oxidase, (219) 40
β-Oxidation, (222) 21
Oxidative inactivation, (225) 233
Oxidative phosphorylation, (208) 1; (223) 247; (223) 279; (223) 395; (226) 352
Oxidative stress, (213) 9
Oxidized insulin A-chain, (219) 441
Oxo-ferryl porphyrin, (208) 305
2-Oxoglutarate dehydrogenase complex, (222) 211
Oxonol-IV, (215) 47
10-Oxo-19-nor-25-hydroxyvitamin D_3, (218) 200
Oxy-radical adduct, (221) 101
Oxygen, (215) 219
Oxygen affinity, (223) 59; (226) 343
Oxygen burst, (223) 284
Oxygen consumption, (216) 31
Oxygen evolution, (211) 215; (216) 234; (221) 245; (223) 376; (224) 71; (226) 161
Oxygen-evolving PS II particle, (226) 186
Oxygen radical, (214) 362; (221) 355; (225) 233
Oxygen saturation curve, (209) 129
Oxygenation, (221) 327
Oxyhemoglobin, (208) 331
Oxypurinol, (213) 23
Oxysterol, (218) 77
Oxytocin, (213) 419; (216) 200

P680, (223) 376
P700, (208) 343; (214) 97; (215) 58; (217) 16
P site, (213) 443
^{31}P relaxation, (217) 21
PA-II lectin, (216) 62
recombinant t-PA, (209) 13
PAF-acether, (226) 371
PAF analog, (211) 225
2D-PAGE, (223) 255
Pancreatic elastase, (225) 223
Pancreatic juice, (216) 195
Pancreatic polypeptide, (225) 209
Pancreatic secretory trypsin inhibitor, (225) 113
Pancreatic stone protein, (216) 195
Parathyroid hormone, (216) 257
Parathyroid hormone release, (213) 195
Parvalbumin, (209) 77; (214) 21
Pasteur effect, (221) 124
Patch clamp, (208) 58; (209) 165; (215) 9; (216) 89; (217) 11; (220) 295; (222) 63; (222) 317; (224) 172; (226) 83
Patch-clamp analysis, (223) 104
Pathogenesis-related protein, (225) 243
PCNA antibody, (209) 277
Penicillin, (220) 206
Penicillin-binding protein, (212) 237; (221) 167; (224) 401; (226) 150
Penicillinase, (221) 167
Penicilloyl group, (220) 206
Pepsin, (219) 1
Peptaibol, (224) 208
Peptide aggregation, (224) 208
Peptide folding, (221) 320
Peptide group, (213) 423
Peptide hydrolysis, (214) 8
Peptide initiation, (215) 16
Peptide isolation, (208) 253; (211) 105; (216) 200
Peptide mapping, (223) 59; (226) 105
Peptide processing, (224) 142
Peptide sequence, (223) 391
Peptide synthesis, (208) 253; (220) 332; (222) 251
Peptide vaccine, (218) 231
Peptide YY, (214) 57
Peptidoglycan, (226) 150
Peptidyl dipeptidase, (219) 441
Perfluoroalkyl ketone, (220) 299
Perforin, (226) 261
Perhydrohistrionicotoxin, (212) 292
Perhydroxyl radical, (220) 149
Periplasmic protein, (224) 213
Permeabilization, (219) 326
Permeabilized cell, (208) 99
Permeabilized platelet, (213) 95
Peroxidase, (208) 305; (208) 391
Peroxidative injury, (218) 251
Peroxide compound, (219) 387
Peroxisomal disorder, (222) 17
Peroxisomal enzyme, (220) 31
Peroxisome, (215) 228; (222) 21
Pertussis toxin, (211) 137; (212) 40; (214) 176; (215) 339; (216) 40; (220) 155; (220) 187; (221) 81; (222) 51; (224) 1; (225) 16
pH control, (224) 187
pH dependence, (225) 218
pH-dependent channel, (226) 115
pH effect, (215) 155
pH gradient, (209) 28
pH indicator, (208) 337; (209) 316
pH sensitivity, (219) 301
cytoplasmic pH, (215) 223; (220) 366
ΔpH, (221) 215
intracellular pH, (217) 169; (229) 288

pH_i, (222) 144
Phage D108, (222) 199
Phage λ gtll, (213) 249
Phage Mu, (208) 7; (222) 199
λgt 11 Phage library, (208) 77
MS2 Phage, (226) 255
T-even type Phage, (215) 145
Phagocytic cell, (218) 200
Phalloidin, (213) 433
Phase transition, (209) 257; (224) 283
Phaseollidin, (220) 223
Phenotypic modulation, (218) 292
Phenylalanyl-tRNA synthetase, (216) 27
Phenylalkylamine, (212) 247
Phenylarsine oxide, (224) 391
Phenylephrine, (218) 1
Phenylhydrazine, (221) 299
N-Phenyl-1-naphthylamine, (218) 22
Pheophytin, (214) 370; (226) 186
Philadelphia chromosome, (222) 293
Phorbol diester binding, (223) 11
Phorbol ester, (208) 151; (209) 134; (210) 204; (211) 151; (212) 242; (213) 89; (213) 365; (214) 122; (214) 139; (214) 171; (215) 175; (215) 223; (215) 279; (217) 69; (218) 143; (218) 238; (219) 119; (220) 57; (221) 359; (222) 345; (223) 11; (223) 15; (223) 321; (224) 33; (224) 109; (225) 233; (226) 223
Phorbol myristate acetate, (223) 284
Phorbol 12-myristate 13-acetate, (222) 27
Phosphatase, (219) 125
Phosphate, (226) 63
Phosphate carrier, (212) 1
Phosphate potential, (208) 138
Phosphate transport, (216) 257
Phosphatidic acid, (218) 93; (225) 201
Phosphatidylcholine, (213) 365; (214) 149; (219) 393
Phosphatidylethanol, (225) 201
Phosphatidylethanolamine, (214) 149; (219) 176
Phosphatidylglycerol, (209) 257
Phosphatidylinositol, (208) 47; (216) 57; (217) 45
Phosphatidylinositol anchor, (225) 273
Phosphatidylinositol bisphosphate, (218) 87
Phosphatidylinositol 4,5-bisphosphate, (223) 87
Phosphatidylinositol kinase, (211) 64
Phosphatidylinositol metabolism, (212) 159
Phosphatidylinositol 4-phosphate, (223) 87
Phosphatidylinositol phospholipase C, (212) 289
Phosphatidylinositol-specific phospholipase C, (225) 273
Phosphatidylinositol turnover, (214) 127; (226) 13
Phosphatidylserine, (209) 257; (216) 45; (218) 238; (221) 397
Phosphatidylserine liposome, (214) 181
Phosphodiester bond, (215) 247
Phosphodiesterase, (219) 97; (221) 253; (222) 37; (222) 266; (224) 19; (225) 127
2′,5′-Phosphodiesterase, (212) 267
Phosphoenolpyruvate carboxylase, (213) 18
Phosphofructokinase, (209) 254; (222) 167; (223) 113
6-Phosphofructo-2-kinase, (213) 375; (224) 317; (225) 37; (226) 43; (226) 121
Phosphoinositidase C, (224) 1
Phosphoinositide, (208) 413; (212) 123; (224) 1; (224) 137; (226) 67
Phosphoinositide breakdown, (209) 352
Phosphoinositol, (208) 413
Phospholamban, (217) 32
Phospholipase, (216) 51
Phospholipase A_2, (211) 83; (211) 119; (216) 45; (219) 169; (219) 176; (220) 187; (221) 397; (222) 11; (223) 267; (224) 137
Phospholipase C, (209) 238; (216) 57; (219) 289; (220) 187; (224) 219
Phospholipase D, (225) 201
Phospholipid, (208) 51; (209) 257; (219) 113; (219) 169
Phospholipid asymmetry, (219) 311
Phospholipid dependence, (214) 181
Phospholipid membrane, (218) 173
Phospholipid metabolism, (218) 68
Phospholipid methyltransferase, (209) 238
Phospholipid monolayer, (213) 49
Phospholipid vesicle, (224) 337
Phosphopantothenoylcysteine, (212) 79
Phosphoprotein, (209) 63; (209) 181; (216) 83; (222) 261; (223) 25
Phosphoprotein pp69, (222) 261
Phosphoprotein secretion, (213) 164
Phosphorylase kihase, (220) 36
Phosphorylation, (208) 117; (208) 269; (208) 352; (208) 455; (209) 63; (209) 162; (210) 177; (212) 281; (213) 18; (213) 85; (213) 365; (214) 249; (215) 241; (215) 279; (216) 211; (217) 32; (217) 74; (217) 237; (217) 253; (217) 287; (218) 1; (218) 195; (219) 321; (220) 57; (221) 77; (221) 199; (221) 309; (224) 325; (225) 251
Phosphorylation domain, (218) 81
Phosphorylation site, (209) 175; (212) 103; (215) 16; (218) 1
Phosphoserine, (215) 21
Phosphotyrosine, (222) 293
Phosphotyrosine antibody, (222) 104
Photoactivable group, (221) 408
Photoaffinity analog, (209) 157; (223) 395
Photoaffinity label, (212) 247; (213) 179
Photoaffinity labeling, (209) 71; (212) 203; (213) 359; (226) 134
Photocycle, (209) 44; (209) 316; (217) 297; (222) 275; (226) 139
Photoinhibition, (214) 370; (215) 31; (226) 186
Photointermediate, (225) 255
M Photointermediate, (209) 44
R350 Photointermediate, (209) 44
Photon activation, (213) 401
Photooxidation, (223) 77
Photophosphorylation, (226) 352
Photoreaction, (225) 255
Photoreactive phospholipid, (223) 366
Photoreceptor, (219) 97; (219) 293; (224) 19; (225) 127
Photoregulation, (224) 287; (224) 343
Photorepair, (225) 133
Photosynthesis, (209) 83; (211) 94; (213) 29; (213) 119; (213) 245; (214) 28; (214) 92; (214) 97; (215) 37; (216) 7; (216) 234; (219) 321; (221) 211; (223) 371; (224) 71; (224) 75; (226) 352
Photosynthetic bacteria, (213) 245; (219) 88
Photosystem I, (208) 343; (214) 97; (215) 37; (215) 58; (217) 16; (217) 283; (220) 74; (221) 260; (224) 75; (225) 59
Photosystem I particle, (218) 52
Photosystem II, (208) 347; (209) 83; (209) 181; (211) 94; (211) 99; (211) 215; (212) 103; (213) 241; (215) 31; (216) 7; (217) 53; (219) 207; (220) 67; (223) 371; (223) 376; (224) 71; (224) 343; (225) 59
Phototrapping, (218) 283
o-Phthaldialdehyde, (219) 233

Phycobiliprotein, (217) 279; (221) 1
Phycobilisome, (215) 209
Phylloquinone, (208) 343; (215) 58; (217) 16
Phylogenetics, (221) 189
Physico-chemical characterization, (212) 193
Physiological regulation, (216) 27
Physiological variant, (219) 431
Phytoalexin, (213) 324
Phytochrome, (224) 287
Phytochrome isoprotein, (225) 215
Phytoene synthesis, (210) 211
Phytoene synthetase, (210) 211
P_i, (213) 375; (221) 270
Picosecond charge separation, (214) 28
Picosecond spectroscopy, (213) 119; (213) 241; (223) 161
Pigment-protein complex, (215) 171
Pigment-protein interaction, (214) 92
Pinealocyte, (208) 133
Planar bilayer, (211) 1
Planar lipid bilayer, (224) 208
Plant gene, (224) 287
Plant respiration, (226) 352
Plant storage protein, (224) 125
Plasma, (215) 311
Plasma membrane, (208) 269; (212) 289; (216) 94; (217) 169; (220) 366; (222) 358; (223) 87; (224) 193; (226) 291
Plasma protein, (213) 254; (220) 129
Plasmid, (215) 335
Plasmid pBR322, (211) 49
Plasmid pKK223, (220) 47
Plasmid pUC18, (220) 47
λPlasmid vector, (208) 183
Plasminogen, (209) 111; (213) 254; (214) 187
Plasminogen activator, (208) 273; (209) 213; (214) 187; (223) 361
Plasminogen activator inhibitor, (209) 213
Plastid, (210) 211
Plastocyanin, (222) 109; (225) 159
Plastocyanin a, (226) 17
Plastocyanin b, (226) 17
Platelet, (210) 132; (213) 179; (214) 176; (215) 183; (219) 283; (222) 144; (222) 299; (223) 47; (224) 311; (225) 228
Platelet-activating factor, (208) 51; (211) 225; (220) 220; (221) 305
Platelet activation, (212) 123; (212) 213
Platelet-derived growth factor, (213) 89; (223) 243
Point mutation, (208) 183; (211) 221
Polarized fluorescence, (223) 409
Poliovirus, (214) 153
Poly(A) sequence, (214) 313
Poly(ADP-ribose) polymerase, (212) 73; (215) 252
Polyacrylamide gel electrophoresis, (211) 78
Polyamine, (213) 365; (219) 380
Polychlorobiphenyl, (220) 231
Poly(dA-dT), (208) 94
Poly[d(G-C)], (213) 341
Polydisperse size, (208) 386
Polyelectrolyte, (225) 77
Polyethylene glycol, (209) 243; (217) 85
Poly (I) · poly(C), (226) 13
Poly(L-lysine), (220) 143
Polymer level epimerization, (212) 199
Polymorphonuclear leukocyte, (221) 95; (224) 29; (225) 103
Polymyxin B, (225) 173
Polyoma, (223) 347
Polypeptide, (211) 94
Polypeptide degradation, (210) 142
Polypeptide separation, (218) 277
33 kDa Polypeptide, (224) 71
Polyphosphoinositide, (214) 339
Polysaccharide B, (216) 261
Polysaccharide lyase, (223) 207
Polysialylation, (225) 33
Polyunsaturated fatty acid, (215) 228
Pore-forming protein, (211) 53; (220) 136
Porin, (211) 1
Porphobilinogen deaminase, (225) 87
Positive regulation, (226) 241
Posttranscriptional regulation, (209) 187
Posttranslational modification, (216) 200; (221) 1; (222) 129
Posttranslational processing, (210) 45; (213) 385; (213) 329; (214) 50
Posttranslational protein processing, (208) 369
Potato virus X, (214) 308
Potential barrier, (225) 255
H^+-PPiase, (215) 47
Pratensein, (214) 101
Prazosin binding, (212) 149
Precursor, (218) 31
Precursor sequence, (209) 92; (225) 159
Predictive method, (213) 297
Pregnancy, (226) 220
Prenyltransferase, (220) 223
Preproapolipoprotein C-II, (213) 221
Preproinsulin, (213) 149
Preproinsulin mRNA, (223) 131
Preprotein, (212) 168
Presequence, (226) 171
Presynaptic membrane, (219) 219; (222) 51
Pretranslational control, (208) 55
Primaquine, (214) 291; (216) 51; (223) 77
(+)-Primaquine, (221) 325
Primary cultured hepatocyte, (224) 311
Primary electron donor, (224) 75
Primary reaction, (213) 241
Primary specificity, (209) 235
Primary structure, (212) 98; (215) 285; (219) 161; (221) 87; (222) 211; (223) 92; (224) 128; (225) 188; (226) 17
Primer extension, (208) 208; (220) 61
Priming effect, (208) 55
Pro-enkephalin, (211) 144
Pro-opiomelanocortin, (211) 144
Proalbumin processing, (219) 75
Processing, (212) 307
Processing enzyme, (211) 144
Processivity, (221) 194
Procollagen assembly, (217) 216
Procollagen propeptide, (217) 216
Type I Procollagen, (212) 229
Prodrug, (225) 97
Proenzyme, (223) 69
Proenzyme activation, (220) 271
Progastrin, (210) 185
Progesterone, (223) 321; (226) 213
Prohormone precursor, (224) 142

Prohormone processing enzyme, (217) 180
Proinsulin, (212) 307
Proinsulin synthesis, (215) 179
Prokaryote gene, (220) 262
Prolactin, (212) 220
Proliferation, (215) 1; (217) 1
Proline, (208) 211
Prolyl endopeptidase, (209) 235
Promoter, (219) 70; (219) 254; (225) 264; (226) 241
Promoter mutation, (219) 189
Promoter region, (222) 337
c-H-ras Promoter, (225) 264
lac Promoter, (220) 47
Proofreading, (221) 194
Propeptide region, (226) 33
Prophenoloxidase, (208) 113
Propylthiouracil, (209) 49
Prosome, (219) 410
Prostacyclin, (222) 27
Prostaglandin, (213) 174; (217) 6; (220) 371; (225) 43
Prostaglandin E_2, (212) 35
Prostate-specific antigen, (225) 168
Prosthetic group, (221) 299
Protamine, (215) 237; (220) 193
Protease, (221) 231
Protease inhibitor, (214) 187
Protein, (212) 276; (218) 52
Protein A-gold labeling, (216) 207
Protein acetylation, (224) 198
Protein activator, (216) 190
Protein bioregulator, (223) 402
Protein blotting, (220) 126
Protein-bound polyamine, (220) 236
Protein catabolism, (216) 265
Protein complex, (213) 391
Protein conformation, (226) 364
Protein crystallography, (214) 8; (224) 97
Protein degradation, (209) 227; (210) 165; (221) 231; (224) 182
Protein disulfide isomerase, (217) 216
Protein evolution, (220) 193
Protein export, (221) 226; (224) 213
Protein family, (214) 301
Protein fluorescence, (215) 155
Protein folding, (220) 15; (221) 115; (223) 327
Protein fragment, (216) 270; (221) 320
Protein-free diet, (215) 13; (216) 287
Protein gene product 9.5, (210) 157
Protein glycosylation, (208) 34
Protein homology, (212) 1
Protein hydration, (213) 423
Protein import, (209) 152
Protein interaction, (215) 261; (219) 189
Protein iodination, (224) 198
Protein isolation, (221) 381
Protein kinase, (208) 352; (208) 423; (209) 162; (209) 219; (212) 103; (213) 149; (215) 16; (215) 279; (216) 211; (220) 47; (221) 33; (221) 403; (222) 279; (223) 97; (223) 131; (223) 217; (224) 325; (224) 377
Protein kinase C, (208) 39; (209) 134; (210) 132; (210) 169; (211) 151; (211) 207; (212) 242; (214) 122; (214) 139; (214) 171; (214) 249; (214) 339; (216) 22; (217) 69; (217) 227; (217) 237; (218) 35; (218) 195; (218) 238; (219) 119; (219) 215; (220) 271; (220) 311; (221) 23; (221) 365; (222) 279; (222) 299; (222) 305; (223) 11; (223) 15; (223) 138; (223) 212; (224) 33; (224) 109; (225) 233; (226) 13; (226) 125; (226) 223; (226) 324
Protein kinase F_A, (211) 190; (216) 291
Protein kinase family, (218) 81
Protein labeling, (210) 137; (212) 163
Protein-lipid interaction, (211) 83; (216) 99; (223) 148
Protein methylation, (220) 8
Protein micelle, (222) 1
Protein NMR, (223) 191
Protein mobility, (209) 111
Protein modelling, (224) 161
Protein movement, (225) 127
Protein phosphatase, (211) 190; (221) 415; (223) 340; (226) 176
Protein phosphorylation, (210) 169; (212) 141; (214) 331; (215) 21; (215) 25; (217) 85; (219) 219; (220) 52; (220) 143; (221) 205; (221) 211; (224) 109; (226) 109
Protein precursor, (209) 227; (221) 423
Protein processing, (224) 213; (224) 261
Protein purification, (219) 426
Protein S, (222) 186
Protein secretion, (217) 174; (222) 261; (226) 53
Protein sequence, (209) 107
Protein sequence comparison, (224) 149
Protein sequencing, (212) 68
Protein structure, (208) 211; (213) 269; (213) 283; (214) 226; (219) 11; (223) 402
Protein structure prediction, (224) 149
Protein surface, (222) 109
Protein synthesis, (208) 77; (208) 117; (210) 173; (212) 317; (215) 16; (215) 115; (215) 187; (215) 257; (216) 257; (217) 37; (217) 203; (221) 110; (223) 181; (224) 54
Protein synthesis inhibition, (225) 53
Protein synthesis initiation factor 2, (215) 109
Protein translocation, (218) 63
Protein transport, (213) 329
Protein turnover, (208) 427
Protein-tyrosine kinase, (208) 451
14-3-3Protein, (219) 79
A* Protein, (218) 119
C-Protein, (208) 123
CTL-specific Protein, (208) 84
D1 Protein, (209) 83; (216) 7
D2 Protein, (209) 83; (216) 7; (223) 371
G-Protein, (208) 241; (208) 413; (211) 113; (212) 281; (213) 199; (214) 226; (215) 109; (215) 339; (216) 94; (219) 97; (220) 155; (220) 187; (220) 327; (221) 253; (222) 51; (222) 191; (222) 317; (224) 1; (224) 43; (224) 219; (224) 365; (226) 67; (226) 155
G-Protein subunit, (216) 104
G_s-Protein, (217) 287
8.3 kDa Protein, (212) 103
16 kDa Protein, (216) 234
23 kDa Protein, (216) 234
32 kDa Protein, (223) 267
33 kDa Protein, (211) 215; (214) 181; (226) 161
36 kDa Protein, (221) 381
38 kDa Protein, (217) 74
47 kDa Protein, (226) 161
60 kDa Protein, (221) 68
87 kDa Protein, (220) 311
M Protein, (208) 435
N-Protein, (218) 7

3-orf Protein, (218) 81; (222) 286
Q_B Protein, (208) 313
ras Protein, (217) 74
S-100 Protein, (214) 35
T-Protein, (216) 133
TonB Protein, (208) 211
VPg Protein, (215) 247
Proteinase inhibitor, (215) 274; (223) 174
Proteinase yscF, (218) 31
Proteoglycan, (209) 299
Proteolipid protein, (223) 417
Proteoliposome, (219) 387; (223) 155
Proteolysis, (208) 113; (209) 139; (210) 142; (211) 185; (215) 115; (216) 253; (219) 455; (220) 278; (221) 231; (222) 6; (225) 163; (225) 215
Proteolytic activation, (223) 15
Proteolytic degradation, (220) 214
Proteolytic fragment, (218) 167
Proteolytic susceptibility, (220) 376
Prothoracicotrophic hormone, (219) 419
Prothrombin, (209) 111
Prothrombin fragment 1, (224) 97
Prothymosin α, (218) 107; (226) 217
Proton conductivity, (213) 67
Proton-deuterium exchange, (221) 241
Proton electrochemical-driven Na^+ pump, (226) 270
Proton gradient, (212) 323; (224) 79
Proton pump, (208) 143; (208) 337; (209) 316; (216) 155; (219) 387; (226) 139
Proton pumping, (226) 1
Proton transfer, (219) 22
Proton translocation, (215) 53; (217) 169
Proton translocation stoichiometry, (208) 138
Protonation change, (213) 39
Protonmotive force, (208) 138; (209) 325; (219) 88
Protonmotive potential, (224) 79
Proto-oncogene, (223) 97; (225) 264
c-fos Proto-oncogene, (222) 289
Protoplast, (212) 114; (212) 133
Protoplast infection, (221) 337
Proximal histidine, (213) 289
PS II particle, (217) 134
Pseudoazurin, (218) 209
Pseudogene, (225) 243
Pseudomembranous colitis, (225) 82
Psychotropic effect, (211) 119
Pterocarpan biosynthesis, (213) 324
Pterocarpan synthesis, (214) 101
PTH release, (208) 99; (218) 113
Pulsed quenched-flow, (226) 331
Pump current, (224) 83
Purification, (212) 247; (213) 215; (215) 105; (215) 140; (216) 67; (218) 277; (220) 227
Purple membrane, (208) 337; ; (209) 316; (216) 155; (216) 241; (218) 287; (223) 289; (226) 139
Pyranine, (208) 337
Pyrazine-binding protein, (212) 225
Pyrazofurin, (224) 59
Pyrethroid, (219) 355
Pyridoxal 5′-phosphate, (225) 109
Pyridylamino derivative, (209) 265
S-Pyridylethylation, (212) 68
Pyrimidine de novo synthesis, (224) 59
Pyrimidine salvage pathway, (224) 59
Pyrithiamine, (220) 84
H^+-Pyrophosphatase, (224) 348
(+)-trans-2-Pyrrolidinyl-N-methylcyclohexylamine, (223) 335
Pyrrolnitrin, (209) 321
Pyrroloquinoline quinone, (214) 236; (214) 274; (221) 299
Pyruvate, (212) 79
Pyruvate kinase, (218) 1
Pyruvate oxidation, (211) 239
Pyruvate transport, (219) 331; (219) 347

Quantum yield, (223) 187
Quench flow, (217) 129
Quinone, (225) 139
Quinone-iron complex, (226) 347
Quinone protonation, (226) 347
Quin-2, (208) 47; (209) 169; (211) 44; (211) 195; (212) 119; (212) 123; (213) 195; (213) 401; (214) 130; (220) 89; (221) 23

Radiation inactivation, (225) 145
Radioimmunoassay, (208) 26; (214) 50
Radioiodinated ligand, (224) 267
Radiosequence analysis, (213) 385
Radish mosaic virus, (215) 247
Raman spectroscopy, (219) 202; (221) 37
Rb efflux, (219) 451
$^{86}Rb^+$ transport, (225) 27
Reaction center, (208) 347; (213) 241; (214) 28; (214) 92; (216) 7; (216) 245; (217) 16; (218) 283; (220) 67; (221) 221; (221) 260; (223) 371; (224) 71
Reaction center complex, (225) 1; (226) 161
Reaction center insertion, (215) 171
Reaction mechanism, (217) 265; (219) 27; (225) 139; (226) 63
Reannealing, (213) 433
Rearrangement, (224) 291
Receptor, (211) 17; (211) 73; (211) 113; (211) 165; (212) 281; (213) 81; (216) 94; (217) 253; (218) 189; (219) 224; (219) 259; (219) 355; (224) 251; (226) 213
Receptor binding, (219) 369
Receptor binding assay, (226) 287
Receptor-binding peptide, (211) 17
Receptor evolution, (211) 200
Receptor heterogeneity, (222) 71
Receptor internalization, (214) 323
Receptor-mediated endocytosis, (216) 159
Receptor sequestration, (212) 149
Receptor subtype, (226) 287
V_1 Receptor, (210) 181
Recombinant glucagon secretion, (212) 302
Recombinant IgE Fcε fragment, (224) 306
Recombinant interleukin-1, (219) 365
Recombinant lymphokine, (226) 47
Reconstitution, (214) 335; (216) 83; (217) 287; (218) 287; (222) 154; (222) 327; (225) 67
Recycling, (221) 105
Redox-linked conformation, (213) 396
Redox potential, (213) 372
Redox potentiometry, (217) 57
Redox reaction, (209) 87
Refolding kinetics, (224) 9
Regiospecific cleavage, (226) 232

Regiospecificity, (221) 327
Regulation, (213) 375; (215) 31; (215) 83; (221) 179; (224) 193; (226) 155
Regulatory enzyme, (216) 133
Regulatory subunit, (209) 219
Reinnervation, (223) 104
Release, (219) 445
Release factor, (226) 227
Remazol yellow GGL, (211) 123
Renal failure, (220) 307
Renin, (220) 299
Repair fidelity, (217) 184
Repair regulation, (225) 133
Reperfusion, (221) 101
Reperfusion damage, (213) 23
Repetitive epitope, (224) 23
Repetitive sequence, (208) 386
Repetitive transient potential, (219) 351
Replication, (223) 347
Repressor, (221) 179; (222) 199
Reserpine, (222) 215
Reserve carbohydrate, (225) 259
Resonance energy transfer, (219) 233
Resonance Raman spectroscopy, (208) 305; (217) 297
Resonance Raman spectrum, (214) 92
Respiration, (217) 169
Respiratory chain, (218) 22
Restrained molecular dynamics, (213) 269
Restricted tissue specifity, (224) 23
Restriction endonuclease, (213) 304; (222) 89
Restriction enzyme, (214) 305
Restriction fragment length analysis, (213) 448
Restriction fragment length polymorphism, (208) 221; (217) 65; (221) 129
Reticulocyte, (219) 410
Reticulocyte lysate, (212) 317; (226) 227
Reticuloendothelial system, (223) 42
Retinal, (213) 113; (218) 287
Retinal cyclic GMP cascade, (220) 15
Retinal isomerization, (217) 297
Retinal protein, (217) 297
Retinoic acid, (208) 151; (224) 43
Retinoid, (214) 167
Retrovirus, (223) 299
Reverse transcriptase, (223) 352
Reverse transformation, (224) 377
Reversible Michaelis-Menten mechanism, (208) 109
Reversible synthesis, (225) 53
Rh antigen, (226) 105
Rhesus polypeptide, (226) 105
Rheumatoid arthritis, (214) 327
Rhodopsin, (208) 241; (213) 113; (219) 97
Ribonucleoprotein, (209) 52
Ribonucleotide reductase, (219) 197; (222) 171
Ribose-5-phosphate isomerase, (212) 45
Ribosomal protein, (220) 126; (220) 227; (224) 65
Ribosomal protein L12, (220) 43
Ribosomal protein S6, (213) 85
Ribosomal subunit, (208) 373
30 S Ribosomal subunit, (220) 319
70 S Ribosomal particle, (209) 104
Ribosome, (208) 321; (209) 52; (209) 104; (213) 443; (215) 115; (216) 83; (220) 126; (220) 319; (225) 205; (226) 255
Ribosome binding, (212) 15
Ribosome binding site, (213) 155
70 S Ribosome, (219) 343
Ribozyme, (219) 279; (220) 283
Ribulose 5-phosphate, (212) 45
Ribulose-bisphosphate carboxylase, (221) 215
Ribulose-1,5-bisphosphate carboxylase, (209) 63
Ribulose-1,5-bisphosphate carboxylase-oxygenase, (224) 287
Ribulose-5-phosphate kinase, (212) 45
Ricin, (208) 373; (215) 155; (216) 99; (219) 51
Ricin A chain, (216) 73
Rieske Fe-S protein, (208) 317; (225) 67
RNA, (214) 308; (214) 313; (226) 63; (226) 255
RNA blot hybridization analysis, (209) 367; (223) 104
RNA dependence, (209) 284
RNA-dependent RNA polymerase, (221) 337
RNA hydrolysis, (215) 327; (226) 232
RNA oligomer polymorphism, (219) 464
RNA polymerase, (209) 284; (212) 21; (212) 28; (215) 83; (218) 163; (219) 189; (224) 257
RNA polymerase III, (219) 400
RNA processing, (214) 143; (223) 63
RNA-protein complex, (215) 247; (224) 54
RNA-protein interaction, (220) 126
RNA sequence, (210) 153
RNA sequencing, (211) 221
RNA splicing, (220) 61
RNA synthesis, (214) 71
5 S RNA, (208) 373; (217) 197; (220) 126
5 S RNA-binding protein, (220) 126
5 S RNA-containing particle, (223) 232
5 S RNA gene, (217) 197
7 SL RNA, (212) 15
double-stranded RNA, (214) 308; (214) 313
subgenomic RNA, (214) 308
T_7 RNA polymerase, (212) 271
mRNA, (208) 67; (208) 379; (213) 149; (213) 189; (215) 68; (219) 75; (221) 81; (222) 191; (223) 117; (224) 54; (224) 104
mRNA stability, (220) 177; (224) 353
mRNA synthesis, (225) 53
mt RNA synthesis rate, (213) 144
rRNA expression, (219) 269
rRNA gene, (209) 223
rRNA operon, (219) 269
5 S rRNA, (209) 52; (212) 15; (213) 301; (217) 191; (221) 189
5 S rRNA synthesis, (214) 143
16 S rRNA, (219) 343; (220) 283
23 S rRNA, (218) 215
23 S rRNA structure, (218) 215
tRNA binding, (220) 283
tRNA-containing particle, (223) 232
tRNA cycle, (223) 1
tRNA processing, (215) 132
tRNA synthesis, (214) 143
aminoacyl-tRNA, (208) 189; (217) 203
aminoacyl-tRNA-binding site, (215) 257
Seryl-tRNA, (226) 227
uncharged tRNA, (223) 1
Valyl-tRNA synthetase, (220) 363
RNase, (208) 373; (209) 335
RNase A, (221) 320; (225) 183

RNase H, (215) 327; (226) 232
RNase P-like enzyme, (215) 132
RNP structure, (213) 309
U snRNP, (213) 309
Rod outer segment, (211) 35; (219) 293; (221) 253
Rod outer segment disk, (213) 113
Rod photoreceptor cell, (208) 133
Rotational correlation time, (218) 135
Rotenone, (219) 108
RPLC, (215) 21
Rubredoxin, (208) 73; (215) 63
Ryanodine, (221) 119

S_1 nuclease mapping, (221) 43
S state, (211) 215
SAA protein, (218) 11
Saccharide binding, (219) 51
Safranine O, (224) 337
Salt adaptation, (220) 98
Salt effect, (225) 77
Sarcomere structure, (224) 49
Sarcoplasmic reticulum, (217) 32; (221) 119
Sarkosyl, (214) 71
Saturation transfer, (208) 89; (226) 179
Schiff base, (213) 113
Scleroderma, (215) 331
Scleroderma antiserum, (209) 231
Sculpin, (214) 57
SDS, (211) 78
SDS electrophoresis, (213) 359
SDS-PAGE, (209) 71; (222) 17
SERRS, (222) 120
Second messenger, (208) 47; (208) 413; (220) 52
Secondary binding site, (212) 83
Secondary electron donor, (208) 347
Secondary structure, (208) 1; (211) 179; (212) 15; (213) 297; (213) 301; (213) 423; (217) 191; (219) 202; (220) 283
Secondary structure analysis, (212) 1
Secondary structure prediction, (218) 231; (220) 167
Secretagogue stimulation, (215) 179
Secretin, (215) 88
Secretion, (208) 357; (209) 169; (211) 44; (213) 189; (213) 195; (215) 183; (223) 47; (224) 14; (226) 307
Secretion-blocking agent, (214) 135
Secretogranin, (224) 142
Secretory granule, (215) 339
Sedimentation equilibrium, (218) 107
Seed storage protein, (226) 145
Segmented analysis, (221) 189
Selectivity, (213) 265
Selenium, (214) 62; (218) 251
Seminal plasma protease, (225) 168
Seminalplasmin, (218) 97
γ-Seminoprotein, (225) 168
Senescence, (208) 313
Senile plaque, (217) 62
Sensory rhodopsin, (225) 255
Sequence comparison, (224) 65
Sequence conservation, (222) 139
Sequence homology, (208) 203; (208) 231; (209) 107; (211) 200; (211) 243; (212) 193; (213) 438; (214) 301; (216) 17; (216) 281; (217) 187; (218) 59; (219) 419; (220) 167; (221) 73; (221) 87; (221) 134; (221) 391; (221) 415; (222) 99; (222) 286; (223) 92; (223) 232
Sequence recognition, (222) 89
Serine esterase, (208) 84
Serine phosphate, (221) 403
Serine protease, (219) 22
Serine protease inhibitor, (224) 14
Serine proteinase, (223) 352
Serotonin, (212) 40; (219) 283
Serotonin-S_2 receptor, (213) 359
Serpin, (209) 213
Serum-free culture, (220) 163
Serum ultrafiltrate, (215) 41
Severe combined immunodeficiency disease, (208) 431
Sex hormone, (220) 129
Sex hormone-binding globulin, (215) 100; (220) 129
Sex pheromone, (218) 31; (221) 423
Sexual cell fusion, (215) 323
Shape determination, (211) 53
β-Sheet, (209) 111; (213) 297
Shift correlation, (208) 94
Sialic acid, (208) 418; (223) 42
Sidechain conformation, (223) 191
Siderophore, (209) 147; (211) 239
Sigma factor, (224) 257
Signal peptide, (212) 168; (224) 213
Signal recognition particle, (211) 133; (212) 15
Signal sequence, (211) 133; (218) 63
Signal transduction, (208) 413; (208) 423; (209) 352; (213) 199; (221) 81; (221) 95; (222) 191; (222) 305; (223) 87
Signal 1, (215) 58
Signal II_{slow}, (223) 371
Single chain, (216) 145
Single-channel conductance, (220) 136
Single-channel recording, (226) 280
Single-site catalysis, (212) 63; (222) 32
Single-strand cleavage mechanism, (209) 269
Singlet oxygen, (211) 49; (213) 401
Site-directed mutagenesis, (209) 295; (216) 79; (222) 56; (223) 294; (226) 150
Site labeling, (223) 391
Site-specific cleavage, (215) 327
Site-specific mutagenesis, (220) 347
Small subunit, (209) 63
Sodium azide, (213) 381
Sodium fluoride, (210) 204
Solar energy conversion, (219) 93
Solar UV, (225) 133
Solution conformation, (215) 88
Solvent isotope effect, (219) 27
Somatic cell hybrid, (217) 42
Somatic mutation, (214) 87
Somatomedin, (214) 259; (222) 181
Somatostatin, (218) 73
Somatostatin-28, (214) 57
Somatostatin receptor, (216) 104
Somatotropin, (208) 269
Southern blotting, (213) 304; (213) 448
Special pair, (224) 75
Specific trypsin cleavage, (216) 195
Specificity, (220) 299
Spectral characteristic, (219) 387

Spectral hole burning, (223) 165
Spectrin, (213) 102; (213) 184; (219) 311; (221) 73; (224) 156
Spectroscopic study, (219) 472
Spermidine, (220) 79
Spermine, (220) 79; (226) 109
Sperm-zona interaction, (226) 38
Sphingomyelin, (223) 42
Sphingosine, (212) 91
Spin echo, (214) 351
Spin labeling, (212) 203
Spin probe, (224) 283
Spin transfer, (219) 5
Spin trapping, (222) 75
Splicing, (213) 309; (215) 269; (223) 417; (226) 63
Sporulation, (224) 257
Stacking interaction, (211) 155; (220) 347
Standard structure, (224) 372
Staphyloccocal α-toxin, (211) 127
Stark effect, (221) 221
Starvation, (211) 41
State 1–state 2 transition, (208) 308; (215) 25
Statistical distribution, (223) 187
Steady state, (226) 343
Steroid, (226) 213
Steroid-binding domain, (215) 100
Steroid-binding protein, (219) 405
Steroid hormone, (225) 151
Steroidogenesis, (219) 113
Stigmatellin, (208) 317
Stimulatory effect, (208) 258; (217) 1
Stimulus response, (219) 259
Stoichiometry, (211) 94; (212) 87
Stopped flow, (215) 83; (221) 115; (221) 349
Stopped-flow kinetic intermediate, (223) 327
Stopped-flow spectrometry, (216) 35
Storage protein, (221) 43
β-Strand, (224) 372
Streptavidin-Sepharose, (215) 305
Stress, (215) 295
Stress response, (225) 259
Stringent response, (223) 1
Structural analysis, (219) 360
Structural homology, (208) 301; (212) 193; (220) 209; (220) 263
Structural model, (226) 88
Structural-reporter signal, (209) 265
Structure, (219) 207
Structure-activity, (209) 122
Structure-activity relationship, (220) 332; (222) 251
Structure analysis, (217) 283
Structure determination, (211) 175; (221) 325
Structure effect, (220) 376
Structure modeling, (219) 419
Structure prediction, (214) 65; (214) 219; (215) 145; (221) 37
Subcellular fractionation, (216) 51
Submandibular gland, (218) 266
Submitochondrial particle, (214) 269
Substance K, (208) 67
Substance P, (208) 67; (221) 236
Substrate analog, (212) 83
Substrate recognition, (220) 283
Substrate specificity, (214) 335; (219) 441; (220) 323
Substrate stereochemistry, (220) 121
Subtilisin nagarse, (225) 215
Subunit, (225) 145
Subunit composition, (220) 116
Subunit sequence, (218) 277
Subunit structure, (211) 179; (221) 134
60 S Subunit, (216) 83
α-Subunit, (214) 107; (217) 275; (223) 366
β-Subunit, (208) 1
Succinate, (214) 357
Succinate pathway, (216) 140
Succinate dehydrogenase, (213) 385; (216) 140; (216) 241
Succinylation, (222) 211
Sucrase-isomaltase, (208) 460
Sugar structure, (209) 265
Sugar transport, (220) 201
Sulfate, (221) 150
Sulfate reduction, (215) 63
Sulfhydryl group, (208) 73; (211) 89; (212) 68
Sulfhydryl reagent, (211) 89; (224) 391
Sulfite-reductase, (221) 343
Supercoiling, (215) 83
Superoxide, (209) 134; (213) 15; (214) 167; (214) 171; (226) 285
Superoxide anion, (217) 158; (223) 284
Superoxide assay, (210) 195
Superoxide dismutase, (211) 211
Superoxide generating enzyme, (214) 279
Superoxide generation, (214) 269
Superoxide production, (216) 31
Suppressor tRNA, (226) 227
cdc25 Suppressor gene, (222) 279
Surface charge, (216) 155
Surface-enhanced Raman scattering, (213) 113
Surface potential, (213) 39
Surfactant, (223) 33
SV40 large T antigen, (221) 199
Synapsin I, (213) 184
Synaptic vesicle, (222) 51
Synaptosome, (213) 61; (216) 57; (218) 189; (219) 296
Synchrotron radiation, (208) 321
Synexin, (217) 25
Synthetic antigen, (208) 435
Synthetic enzyme, (210) 147
Synthetic gene, (208) 194
Synthetic peptide, (221) 172; (222) 266; (224) 156
Synthetic peptide substrate, (223) 15
Synthetic substrate, (209) 235

T-cell, (224) 291
T-cell receptor, (218) 87; (219) 360
T lymphocyte, (225) 16
Tachykinin, (208) 67
Tail fiber protein, (215) 145
Tamm-Horsfall glycoprotein, (226) 314
Tandem promoter, (212) 21; (219) 189
Taurine, (224) 4
Taurocholate transport, (213) 34
Teleost, (219) 227
Temperature dependence, (208) 89; (221) 245
Terbium, (218) 97
Terbium binding, (225) 223
Terminal deoxynucleotidyltransferase, (213) 204
Termination substrate, (219) 151

Ternary complex, (209) 162; (217) 203
Ternary model, (225) 109
Tertiary granule, (217) 158
Tertiary structure, (208) 1; (209) 52; (213) 269; (219) 419
Tetanus toxin, (213) 337; (226) 115; num); (226) 115
Tetrabenazine, (222) 215
Tetracycline, (213) 443
Tetradecanoyl phorbol acetate, (226) 324
12-O-Tetradecanoyl 13-acetate, (225) 43
N,N,N′,N′-Tetramethyl-p-phenylenediamine, (223) 155
Tetranitromethane, (218) 287
Thermal stability, (215) 115; (215) 165
Thermal stability map, (226) 247
Thermodynamic stability, (211) 179
Thermogenin, (224) 353
Thermoluminescence, (211) 215
Thermotolerance, (220) 113
Thesaurin a, (223) 232
Thiamine pyrophosphate, (211) 239
Thiazoline ring formation, (226) 319
2-Thiocytidine, (208) 189
Thiol activation, (218) 266
Thiol ester, (221) 55; (222) 83
Thiol-liposome, (214) 13
Thiol proteinase, (211) 23
Thiophosphorylation, (208) 117
Thioredoxin, (209) 87; (222) 353
Thioridazine, (222) 21
Three-dimensional crystal, (220) 319
Three-dimensional electron microscopy, (211) 53
Three-dimensional peptide structure, (225) 209
Three-dimensional solution structure, (219) 11
Three-dimensional structure, (218) 55
Thrombin, (211) 10; (214) 176; (218) 68; (222) 27; (223) 47; (225) 72; (225) 109; (225) 228
Thrombin inhibitor, (211) 10
Thrombin specificity, (222) 6
Thromboxane, (225) 103
Thromboxane receptor, (213) 179
Thromboxane synthesis, (213) 95
Thylakoid, (215) 25; (215) 37; (221) 205; (221) 215
Thylakoid chlorophyll fluorescence, (208) 308
Thylakoid membrane, (209) 28; (213) 29; (225) 59
Thylakoid membrane organization, (215) 31
Thylakoid protein kinase, (226) 155
Thymidine kinase, (209) 191; (217) 292; (223) 122
Thymidine kinase gene, (212) 173
Thymocyte, (213) 401; (224) 104
Thymosin α_1, (218) 107
Thyroid, (208) 391; (215) 316; (221) 91
Thyroid autoantibody, (211) 123
Thyroid hormone, (219) 135; (224) 251
Thyrotropin, (210) 204; (215) 316; (226) 223
Thyrotropin receptor antibody, (211) 69
Tight junction, (221) 359
Tightly bound nucleotide, (219) 156
Time-resolved fluorescence, (224) 406
Time-resolved light scattering, (208) 241
TIMP, (209) 9
Tissue culture, (224) 396
Tissue specificity, (221) 129; (222) 332
Tissue-type plasminogen activator, (208) 273
Titanium dioxide, (219) 93
Titin, (224) 49
TLC, (209) 362
Tobacco mosaic virus, (219) 65
α-Tocopherol, (209) 191
Topoisomerase I, (209) 231
Topoisomerase II, (213) 304
Toxin, (208) 236
Toxin A, (225) 82
Toxin mutant, (215) 73
α-Toxin, (224) 277
β-Toxin, (214) 163
Tracer kinetics, (216) 128
Trans splicing, (210) 153
Transacylase, (220) 323
Transcript termination, (212) 21
Transcription, (212) 21; (212) 28; (212) 208; (212) 271; (215) 331; (219) 65; (219) 400; (221) 332; (225) 120
Transcription factor, (222) 337
Transcription factor IIIA, (217) 197
Transcription initiation, (215) 83
Transcription regulation, (213) 144
Transcription vector SP6, (215) 132
Transcriptional complex, (214) 71
Transcriptional control, (212) 233; (224) 287
Transcriptional enhancer, (218) 41
Transducin, (220) 15; (222) 266; (224) 19
Transesterification, (226) 72
Transfection, (212) 173; (221) 332; (223) 347
Transferrin, (215) 41; (216) 159; (222) 271
Transformation, (214) 81; (219) 400; (223) 237
Transforming growth factor β, (222) 261
Transglutaminase, (220) 236; (224) 104
Transhydrogenase, (219) 88
Transient kinetics, (213) 396; (224) 83
Transit peptide, (216) 234
Translation, (220) 23
Translation regulation, (214) 331
Translation termination, (225) 205
Translational accuracy, (223) 1
Translational efficiency, (213) 155
Translational inhibitor, (209) 162
Transmembrane arrangement, (217) 269
Transmembrane segment, (222) 56
Transmembrane signaling, (216) 104; (220) 217
Transpeptidation, (212) 163
Transposable bacteriophage, (222) 199
Tn3 Transposase, (208) 7
Trehalose, (220) 113; (225) 259
Triiodothyronine, (220) 163; (223) 279
Trimethylammonium diphenylhexatriene, (223) 381
Triton X-100, (226) 72
Triton X-114, (213) 107
Tropomyosin, (218) 139
Tropomyosin-troponin, (208) 123
Trypanothione, (221) 427
Trypsin digestion, (226) 166
Trypsin protecting activity, (219) 37
Trypsinogen, (211) 185
Tryptic cleavage, (211) 127
Tryptic digestion, (217) 269
Tryptic proteolysis, (212) 254

Tryptophan, (211) 83
Tryptophan 5-monooxygenase, (219) 79
Tryptophan promoter, (212) 233
TSH receptor, (215) 316
Tubulin, (209) 23; (213) 184; (218) 102; (219) 31; (222) 311
Tubulin gene, (208) 379
Tubulin isotype, (219) 103
α-Tubulin, (217) 260
Tubulinyl-tyrosine, (219) 31
Tuftsin, (226) 303
Tumor differentiation, (215) 122
Tumor necrosis factor, (211) 179; (226) 314
Tumor progression, (208) 17
Tumor promoter, (214) 339; (221) 359; (223) 299
Tumor promotor, (208) 455
β-Turn, (224) 372
β-Turn conformation, (213) 265
Turnover rate, (213) 144; (222) 32
Two-dimensional crystal, (211) 53
Two-dimensional sheet, (209) 104
Type X collagen, (220) 181
Tyrosinase, (219) 335
Tyrosine kinase, (209) 357; (212) 141; (222) 293
Tyrosine 3-monooxygenase, (219) 79
Tyrosine protein kinase, (215) 241; (222) 104; (223) 6
Tyrosine residue, (219) 51
Tyrosine synthesis, (216) 133
Tyrosylsulfotransferase, (211) 234

U46619, (213) 179
U50,488 synthesis, (223) 335
Ubiquinol-cytochrome C_2 oxidoreductase, (213) 128
Ubiquinol:cytochrome c reductase, (219) 161
Ubiquitin, (209) 92; (220) 278
UDP-L-arabinose-hydroxyproline-O-glycosyltransferase, (221) 293
UDP-sugar transport, (208) 407
Ultrasonic relaxation spectrometry, (219) 22
Ultraviolet crosslinking, (219) 410
Ultraviolet difference absorbance technique, (208) 23
Ultraviolet irradiation, (221) 110
Uncoupled mutant, (224) 365
Uncoupler, (226) 77; (226) 265
Uncoupling, (224) 353
Uncoupling protein, (211) 89; (212) 1; (226) 166
Uni-site catalysis, (213) 381; (218) 222
Unidirectional flux, (208) 89
Unsaturated fatty acid, (221) 33; (221) 289
Urate, (219) 145
Ureogenesis, (224) 201
Urokinase, (208) 273; (214) 187; (223) 361
Uromodulin, (226) 314
Uronic acid, (215) 79

V_1 receptor, (218) 93
Valinomycin, (224) 337
Vanadate, (224) 89
Vanadium, (216) 1
Vapor-phase modification, (212) 68
Variant surface glycoprotein, (221) 37; (225) 53
Vasoconstriction, (213) 174
Vasopressin, (210) 181; (216) 200
Veratryl alcohol, (220) 149; (221) 327
Very long-chain fatty acid, (214) 158; (222) 21
Vesicle-melittin transition, (218) 173
Vestitone, (213) 324
Vimentin, (222) 311
Vinblastine, (209) 23
Vinculin, (213) 428; (220) 291
VIP, (211) 151; (218) 73
VIP receptor, (211) 151
Virus replication, (223) 352
Virus-membrane fusion, (221) 61
Viscosity, (219) 97
Vision, (211) 35
Visual signal transduction, (220) 15
Visual transduction, (219) 293
Vitamin B12, (220) 358
Vitamin D, (212) 138; (226) 58
Vitamin D metabolism, (218) 200; (220) 307
Vitamin K, (222) 353
Volatile anesthetic, (221) 28
Voltage clamp, (214) 163; (214) 295; (214) 347
Voltage-dependent channel, (211) 1; (224) 208
Voltage gating, (226) 115
Voltage-sensitive Ca^{2+} channel, (216) 40

Warfarin, (222) 353
Water content, (223) 33
Water-soluble enzyme, (226) 150
Water suppression, (216) 249
Weaning, (214) 41
Western blotting, (208) 439; (212) 237
Wheat histone gene, (223) 273

X-linkage, (220) 36
X-linked muscular dystrophy, (212) 313
X-ray absorption, (208) 325
X-ray analysis, (221) 325; (223) 335
X-ray diffraction, (208) 301; (212) 49; (225) 123
X-ray structure, (211) 185
Xanthine, (225) 97
Xanthine antagonist, (209) 122
Xanthine oxidase, (213) 23; (222) 246

Yeast, (219) 339

Z variant, (216) 79
Zero-field splitting, (211) 155; (220) 347
Zero-phonon transition, (223) 165
Zinc chelate-Sepharose CL4B, (209) 13
Zn^{2+} effect, (214) 35
Zona binding, (226) 38

the names and initials of the author(s), year of publication, title of the book, name and initials of editor(s) in parentheses, volume, edition, inclusive page numbers, and name and location of the publisher. No part of the references should be underlined; 'and' should be used instead of '&'; authors' surnames should always be in lower case and precede authors' initials. Citations as 'unpublished' and 'personal communication' should be placed in the main text. *Examples of journals [1] and books [2,3]:*

[1] Diller, R., Stockburger, M., Oesterhelt, D. and Tittor, J. (1987) FEBS Lett. 217, 297–304.

[2] Gomperts, B.D. and Fewtrell, C.M.S. (1985) in: Molecular Mechanisms of Transmembrane Signalling (Cohen, P. and Houslay, M.D. eds) Mol. Aspects Cell. Regul. vol. 4, pp. 377–409, Elsevier, Amsterdam, New York.

[3] Robson, B. and Garnier, J. (1986) Introduction to Proteins and Protein Engineering, Elsevier, Amsterdam, New York.

3. Preparation of Figures

3.1. All line figures (including graphs) should be clearly drawn in black ink on white paper and machine-lettered to allow for reduction. Graph paper should be ruled in pale blue and any grid lines to be shown should be in black ink. All figures should be designed to economise on space. Half-tone figures should not be included in line drawings. Figures will be returned to authors only if requested when the manuscript is submitted.

3.2. *Nucleic and amino acid sequences* should be submitted as sharp, clean prints already reduced as much as possible to fit within a single-column (7.6 cm) or double-column width (16.2 cm), while retaining perfect legibility (e.g. to avoid confusion between the letters C and G). If a word-processor is used to generate the sequences, a letter-quality printer should be employed; matrix-printed characters do not reproduce.

3.3. Original half-tone figures should be very sharp, high-contrast photoprints and be designed to display relevant information only, avoiding disproportionately large labelling of lanes. Line drawings should not be included in half-tone figures.

3.4. *Symbols:* The following symbols can be typeset and should be used in preference to non-standard forms:

△ ▽ ◇ ○ ⊙ ⊖ ⊕ □ ☆ ▲ ▼ ◆ ● ■ ★ × +

3.5. When preparing figures, particularly graphs, we urge authors to follow the suggestions of H.G. Hers ((1984) Nature 307, 205). In particular:

3.5.1. Graph with more than one line: identify each by appropriate labelling.

3.5.2. Figure containing several panels with the same axes, usually denoted a,b, etc: indicate on each panel its experimental specificity.

3.5.3. Use one symbol for the same experimental condition in all comparable figures in the article.

3.5.4. If several symbols necessary: use the open circle (suggesting nothing added) for the control, e.g. experiment without (○) or with (●) an addition or treatment.

3.5.5. Label axes as precisely as possible: e.g. 'Time after drug addition' rather than 'Time'. Also express results in mol rather than cpm or absorbance units, sparing readers the task of conversion. If results are given in percent, define 100% in standard units in the legend.

3.5.6. *Legend:* should be as short as possible, and could often be restricted to the title since, ideally, the figure would be intelligible without referring to the legend.

3.6. *Colour reproduction:* When these are essential to the understanding of a paper, figures may be reproduced in colour, at the authors' own expense. This will, however, lengthen the publication time. Half-tones: first page, Dfl. 2,700; subsequent pages, Dfl. 1,600. Line figures (e.g. computer graphics): black + 3 colours, Dfl. 1,800; black + 2 colours, Dfl. 1,250; black + 1 colour, Dfl. 700.

4. Ethics of Experimentation In scientific investigations in people the steps taken to avoid unjustified risk or discomfort should be stated. These should include peer review of the proposed investigation and informed consent by the subjects being investigated. Similarly the steps taken to protect animals from pain should be stated explicitly.

5. Proofs Proofreading will be undertaken by the Publisher. *Unless strictly necessary* and specifically requested on the title page or by the Editor, proofs will not be sent to authors. Proofs will be supplied to authors only for checking of typesetting accuracy – no change to the accepted manuscript will be allowed at this stage (see 2.9 above). Sending of proofs to authors will of course delay publication of those papers; to minimise this, authors should respond within 24 hours after receipt of their proofs.

6. Requests for Permission to Reproduce Material from Published Articles Authors wishing to reproduce figures, tables and excerpts of text (not exceeding 250 words) from articles published in *FEBS Letters* for *non-commercial purposes* may do so providing the original publication is acknowledged accordingly and the authors' approval is obtained.

7. Reprints On acceptance of a paper the submitting author will receive an order form on which all authors' reprints of that article may be ordered, including the 50 free reprints. In addition, the author should supply supplementary material as requested under (a)–(d) on the form. The intact reprint order form should be returned as soon as possible to: *FEBS Letters*, Elsevier Science Publishers BV, PO Box 1527, 1000 BM Amsterdam, The Netherlands.

8. Editorial Matters Address all queries concerning accepted manuscripts to: *FEBS Letters*, Elsevier Science Publishers BV, PO Box 1527, 1000 BM Amsterdam, The Netherlands **(Telex 18582 ESPA NL)**, specifying editor's code and manuscript number.

Notes to Authors

Editorial Policy Papers should be short, complete and essentially final reports. Papers not sufficiently sustained by experimental details will not be published. The over-riding criterion is that a paper must be of sufficient immediate importance to the work of other investigators to justify urgent publication. *FEBS Letters* is not a repository for material which, though sound, does not meet this criterion.

Subject Coverage Protein Chemistry, Enzymology, Biophysical Chemistry, Nucleic Acids, Protein Synthesis, Biochemical and Molecular Genetics, Morphogenesis, Cell Biology, Metabolism, Immunochemistry, Natural Products.

Submission Procedure Research Letters may be sent to any of the Editors. *The authors should submit their papers to the Editor who is the closest to their field of interest (rather than geographically): this accelerates the reviewing process.* The full addresses of the Editors are given on page 2 of the cover (see 2.2–2.4 below). Papers may be submitted in English, French or German, with an abstract in English. *Authors should include 3 self-addressed sticky labels* with their manuscript to facilitate editorial handling *and provide a telex number* on the manuscript to enable the Editor/Publisher to contact them on queries. Authors may *not* re-submit to other Editorial Board Members a paper that has already been rejected.

Submission of a paper to FEBS Letters is understood to imply that this has not been published in part nor is being considered elsewhere, and that all authors agree to the submission.

In addition to Review Letters, Discussion Letters, Meeting Reports, Commentaries and Hypotheses, Mini-Reviews of 2000 words and less than 15 references may be submitted in essay format to Professor H.R.V. Arnstein (to whom copies of books for review should also be addressed). Reviews should be brief, topical and of interest to the general reader as well as to specialists. Sufficient information should be given in the Introduction to explain the background for non-specialists. Discussion Letters and Hypotheses should present novel ideas or re-interpretation of established observations but should be based on sound data and not be excessively speculative.

1. General Arrangement of Research Letters The first page should carry the title, authors' names and affiliations (with full postal addresses), an abstract, up to six specific keywords and appropriate footnotes. The abstract should be less than 100 words and summarize the research problem and pertinent findings; the keywords should reflect significant factors of the investigation as a whole (see 2.7 below). On subsequent pages the Introduction (1.) should be followed by numbered sections: 2. Materials and Methods, or 2. Experimental; 3. Results; 4. Discussion. When appropriate these may be combined. Acknowledgements should precede References and not be included as footnotes. Notes Added in Proof should appear before Acknowledgements. Figure legends and tables (see 3 below) should appear on separate sheets after References. Appendices should appear before Acknowledgements and have references continuing the main list.

2. Preparation of Text

2.1. Manuscripts must be typewritten, *double spaced with wide margins,* on one side of the paper only.

2.2. The original manuscript and figures, and two copies of both, should be submitted to facilitate refereeing. Unless requested otherwise, only the top copy of a manuscript will be returned to the author if a paper is rejected.

2.3. The manuscript should not be longer than about six pages, double-spaced typewritten, including the figure legends, tables and references. The Editors will only accept longer papers when there are compelling reasons. Any repetition of information in the text and figures should be avoided.

2.4. Papers may be returned to authors for shortening and language correction. Revised versions returned more than two months after the original date of receipt will be considered as resubmissions and will carry only the date of receipt of the revised version.

2.5. The author(s) for correspondence and full postal address should be indicated in a footnote on the title page.

2.6. If the paper is a dedication, part of a numbered series or has been reported in part previously, this should be indicated in a footnote on the title page.

2.7. *Keywords:* Authors are requested to consult the annual cumulative subject index of *Biochimica et Biophysica Acta* or the list of subject headings from Index Medicus for preferred synonyms and standard abbreviations. A maximum of six keywords should be selected and included with the submitted manuscript. American spelling will be used and plural terms avoided where possible. The list submitted may be amended to ensure that index entries are consistent in the master index. General terms such as enzyme, membrane, etc. should not be used unless qualified, e.g. membrane phosphorylation, enzyme activation. Terms indicating the origin of the material studied such as rat, *E. coli,* etc. should only be used as sub-keywords within parentheses, e.g. (Rat heart) (Bovine uterus) (CHO cell). Example: Game, S., Holman, G. and Eisenthal, R. (1986) FEBS Lett. 194, 126–130.

Title: Sugar transport in *Trypanosoma brucei:* a suitable kinetic probe

Keywords: Sugar transport; D-Glucose; 1,5-Anhydro-D-glucitol; 1-Deoxy-D-glucose; *(Trypanosoma brucei)*

2.8. *Nomenclature and abbreviations:* Authors should follow internationally agreed rules as set out in 'Information for Contributors' to *Biochimica et Biophysica Acta* (1982) Elsevier Science Publishers BV, PO Box 1345, 1000 BH Amsterdam, The Netherlands, and in 'Biochemical Nomenclature and Related Documents' (1978), The Biochemical Society Book Depot, PO Box 32, Commerce Way, Colchester CO2 8HP, England. Abbreviations should be expanded in a footnote on the title page.

2.9. Typescripts should be carefully checked before submission to obviate alterations after acceptance.

2.10. *References:* These will be numbered in square brackets, e.g. [7], or [11–13,17], in order of citation in the text. The Harvard system of citation will not apply. The list of references will be printed at the end of the paper. References to journal articles should contain the names and initials of all authors, year of publication in parentheses, abbreviation of the journal title according to current practice, volume number and inclusive page numbers. Articles may be cited as 'in press' only if they have been accepted for publication. References to books should include